

68 to 1

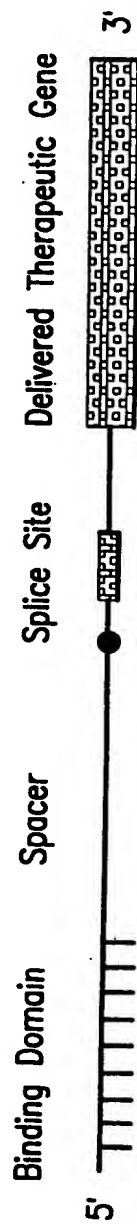


FIG. 1A

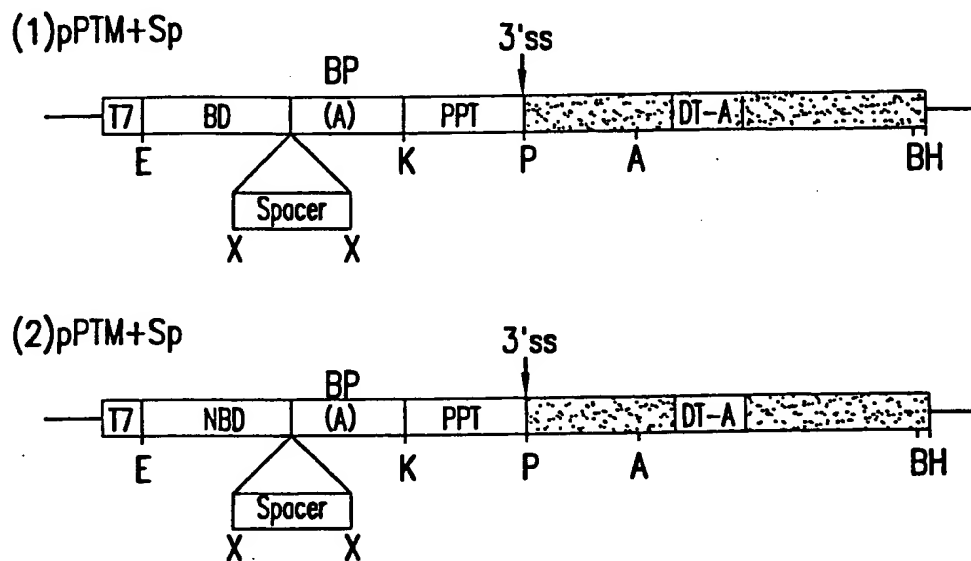


FIG.1B

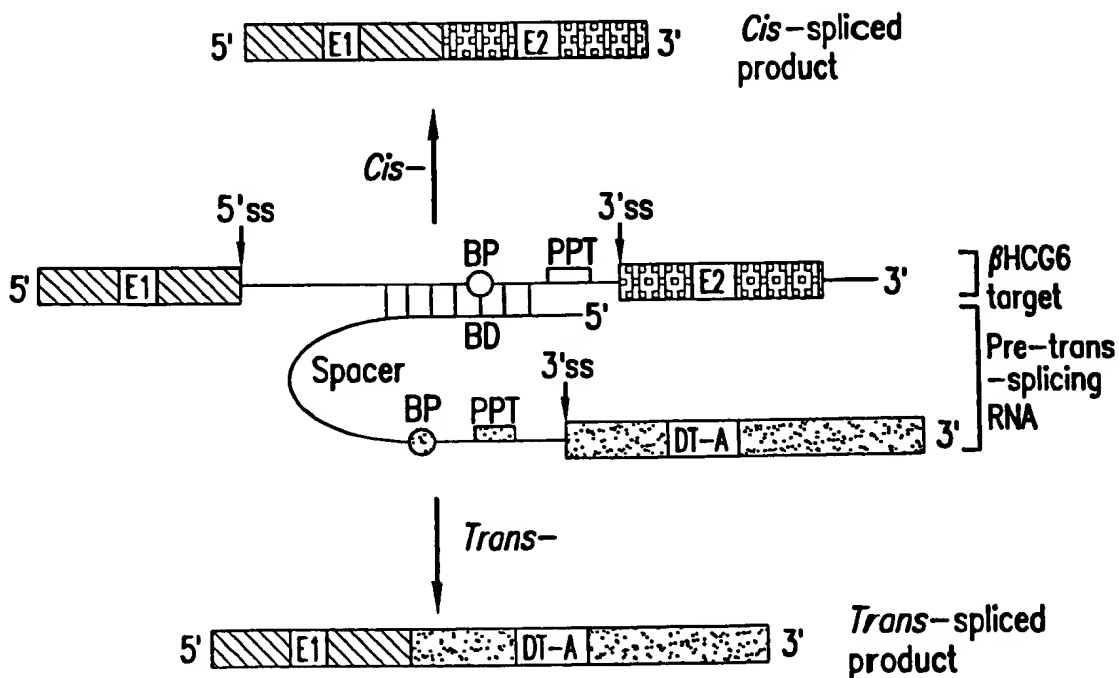


FIG.1C

054449-03001

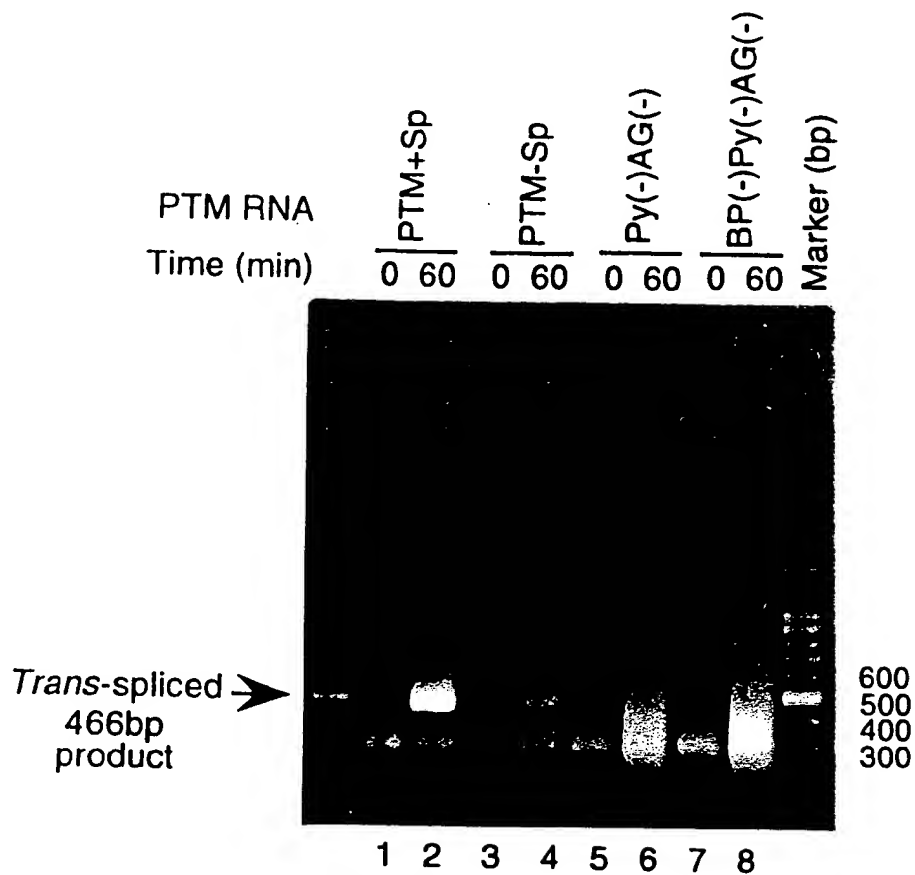


FIG.2A

106290-264T4660

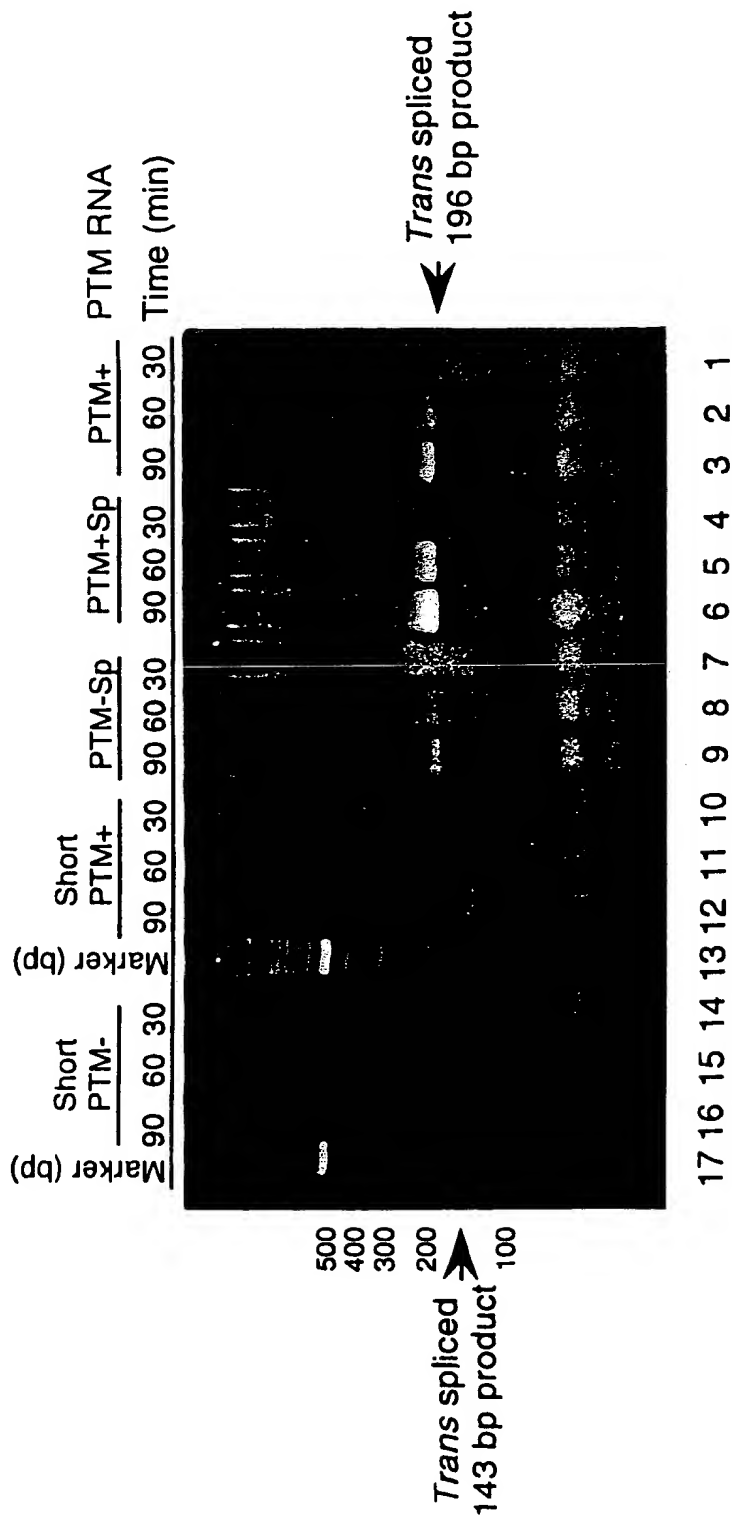


FIG.2B



5 of 89

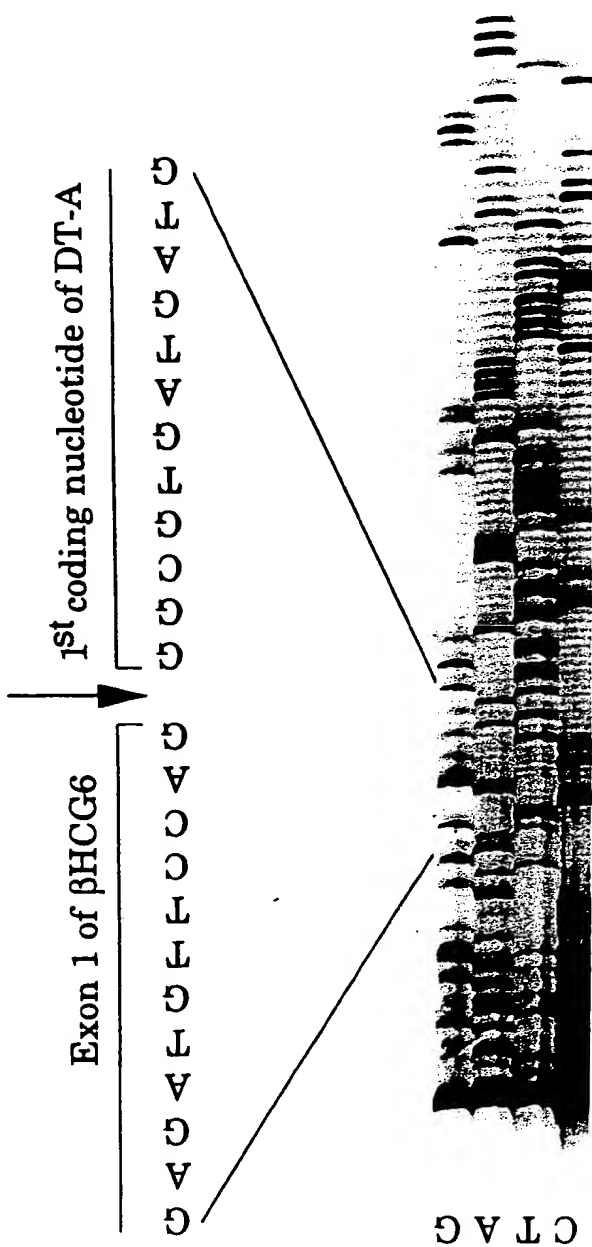
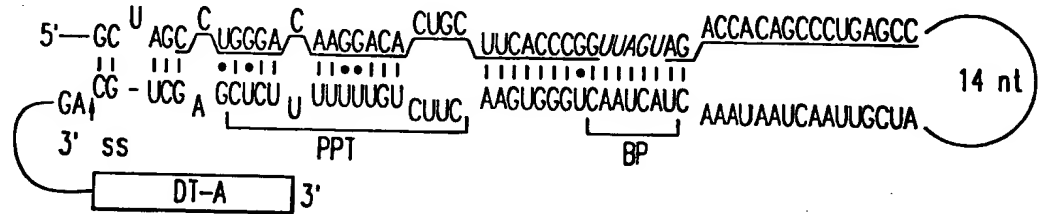
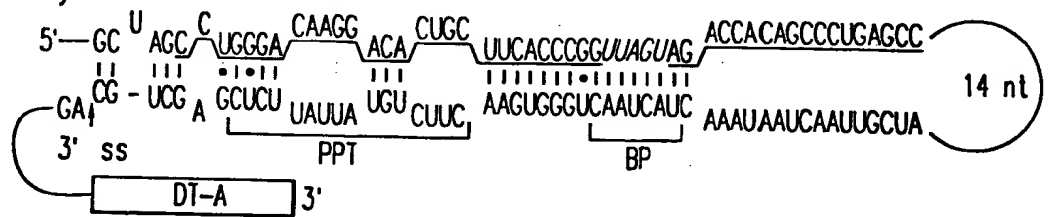


FIG. 3

## 1. PTM+SF:



## 2. PTM+SF-Py1:



## 3. PTM+SF-Py2:

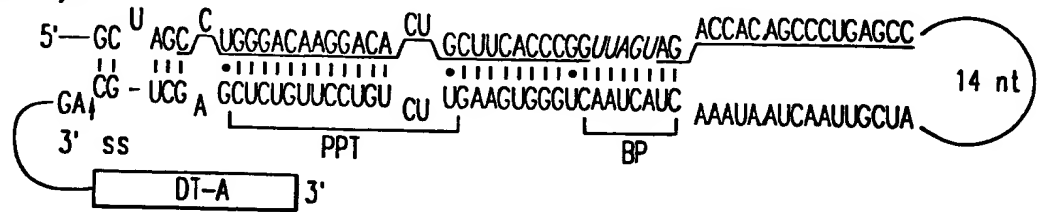


FIG.4A

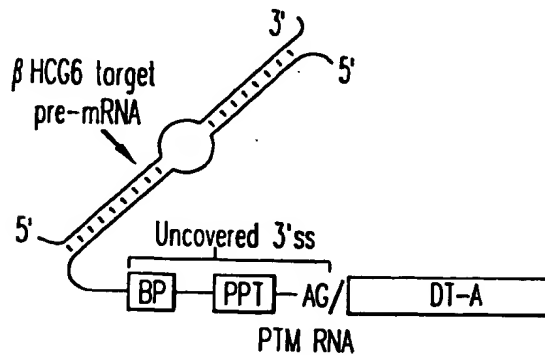


FIG.4B

105280"264T460

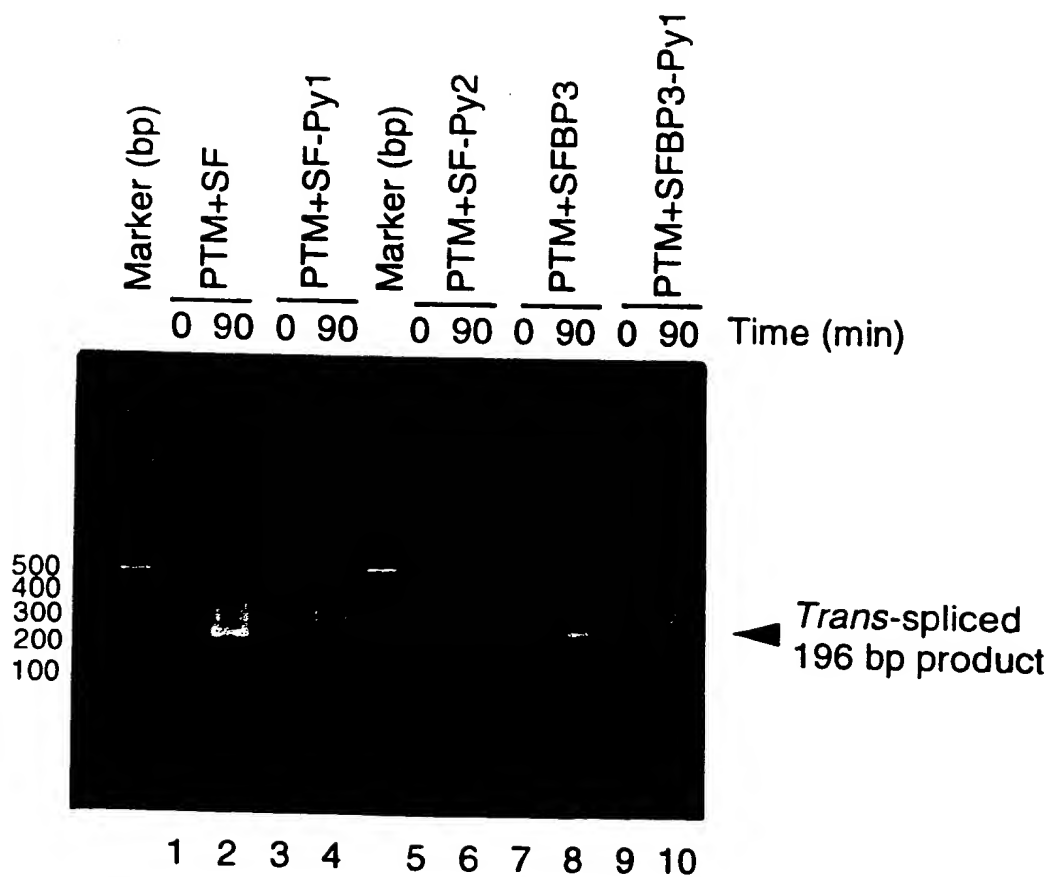


FIG.4C

8 of 8

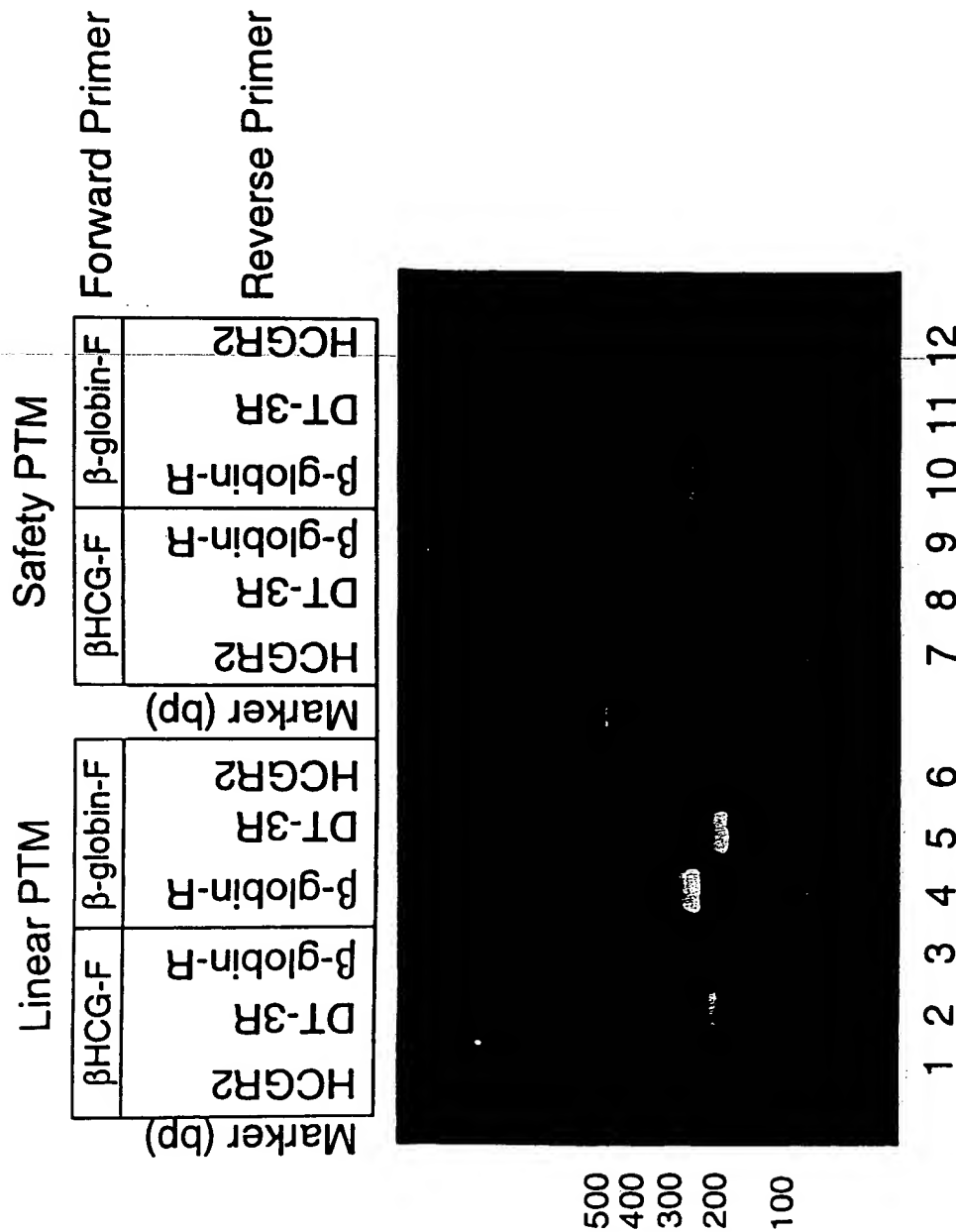


FIG.5

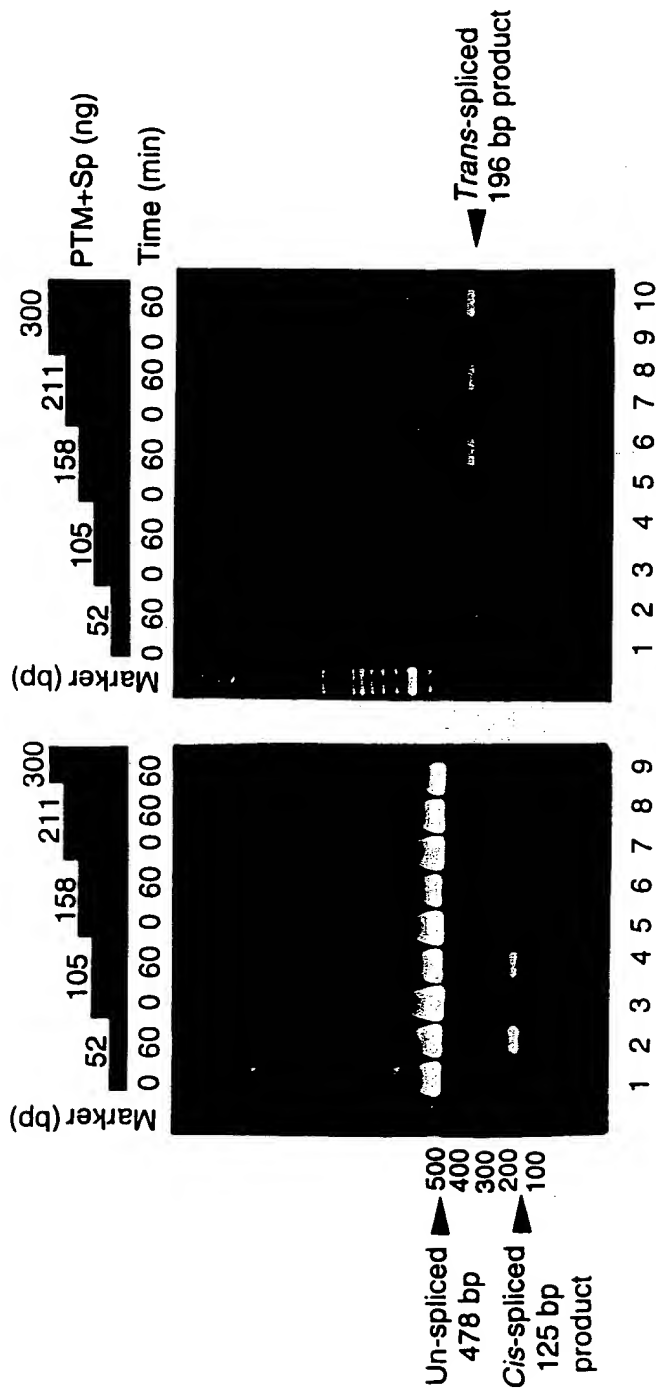


FIG. 6B

FIG. 6A

10 of 89

105290" 264T460

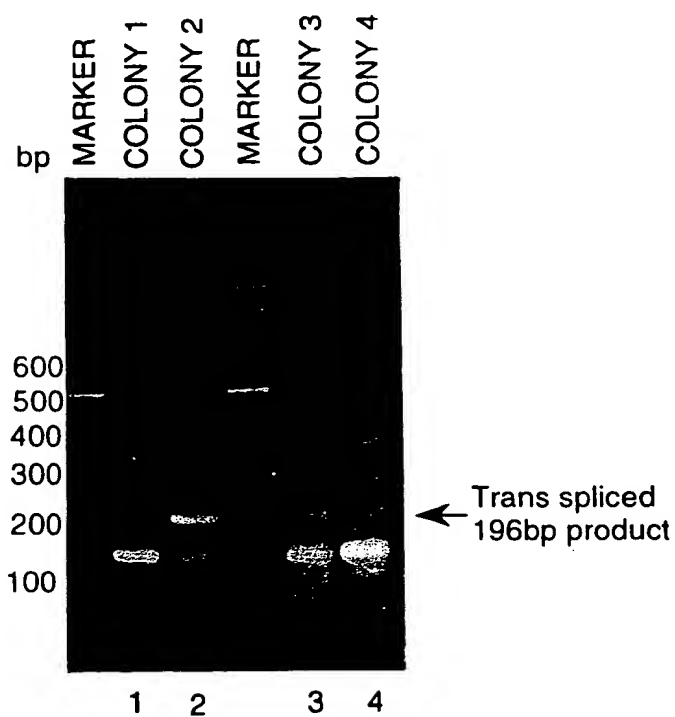


FIG.7A

68 80 11

EXON 1 OF  $\beta$ HOG6 ↓  
 5'-CAGGGACGCCACCAAGGATGGAGATGTTCCAG-GGGCTGATGATGTTGTT  
 ↓ 1ST CODING NUCLEOTIDE OF DT-A  
 GATTCCTTAAATCITTTGTGATGGAAAACITTTCTTCGTACCACGGGACTA  
 AACCTGGTTATGTAGATTCATTCAAAA-3'

FIG.7B

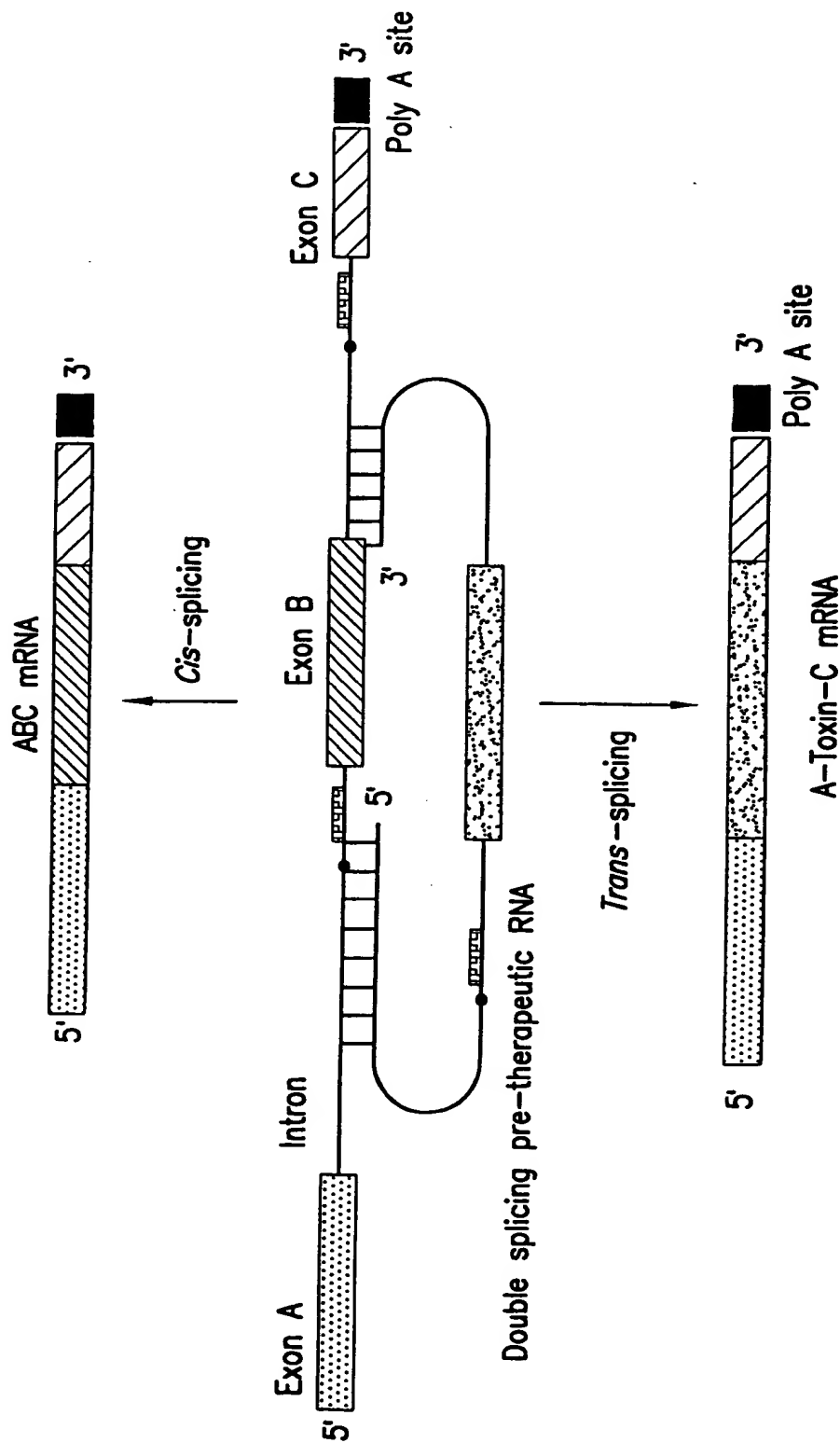
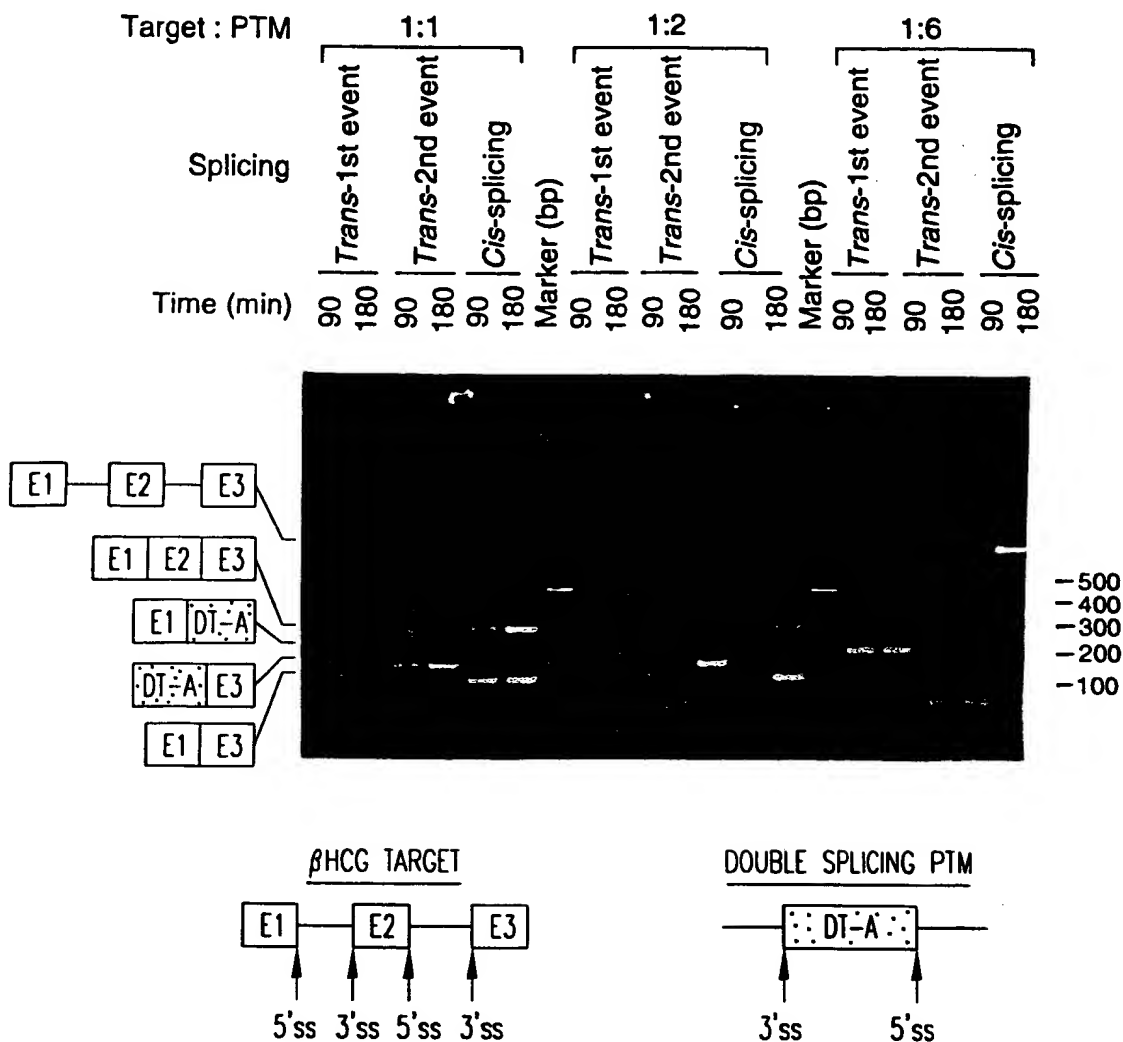


FIG. 8A





Cis-spliced products

E1 E2 E3 = NORMAL *cis*-SPLICING (277bp)

E1 E3 = Exon SKIPPING (110bp)

Trans-spliced products

E1 DT-A = 1st EVENT, 196bp. *Trans*-SPLICING BETWEEN 5' ss OF TARGET & 3' ss OF PTM.

DT-A E3 = 2nd EVENT, 161bp. *Trans*-SPLICING BETWEEN 3' ss OF TARGET & 5' ss OF PTM.

FIG.8B

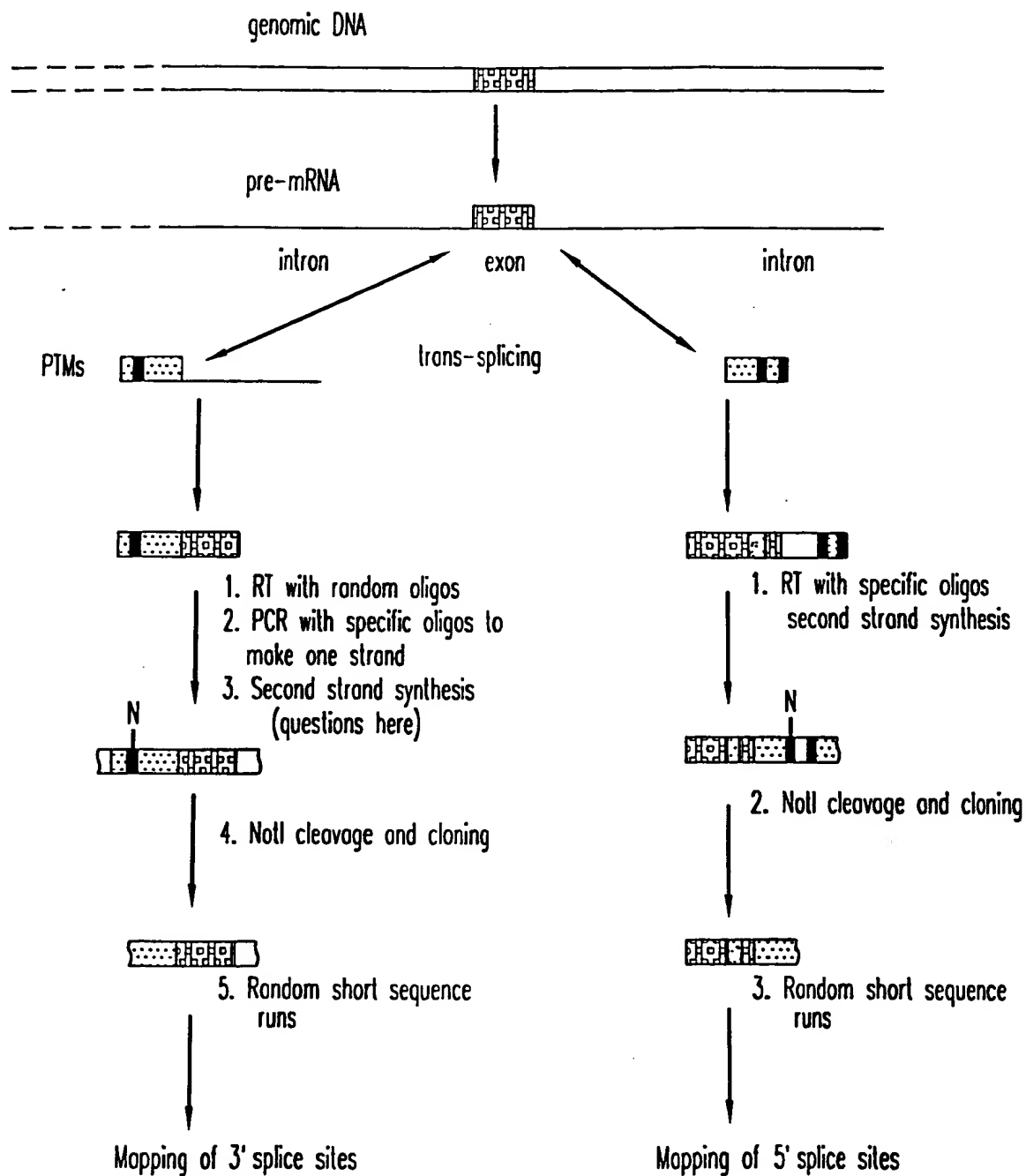
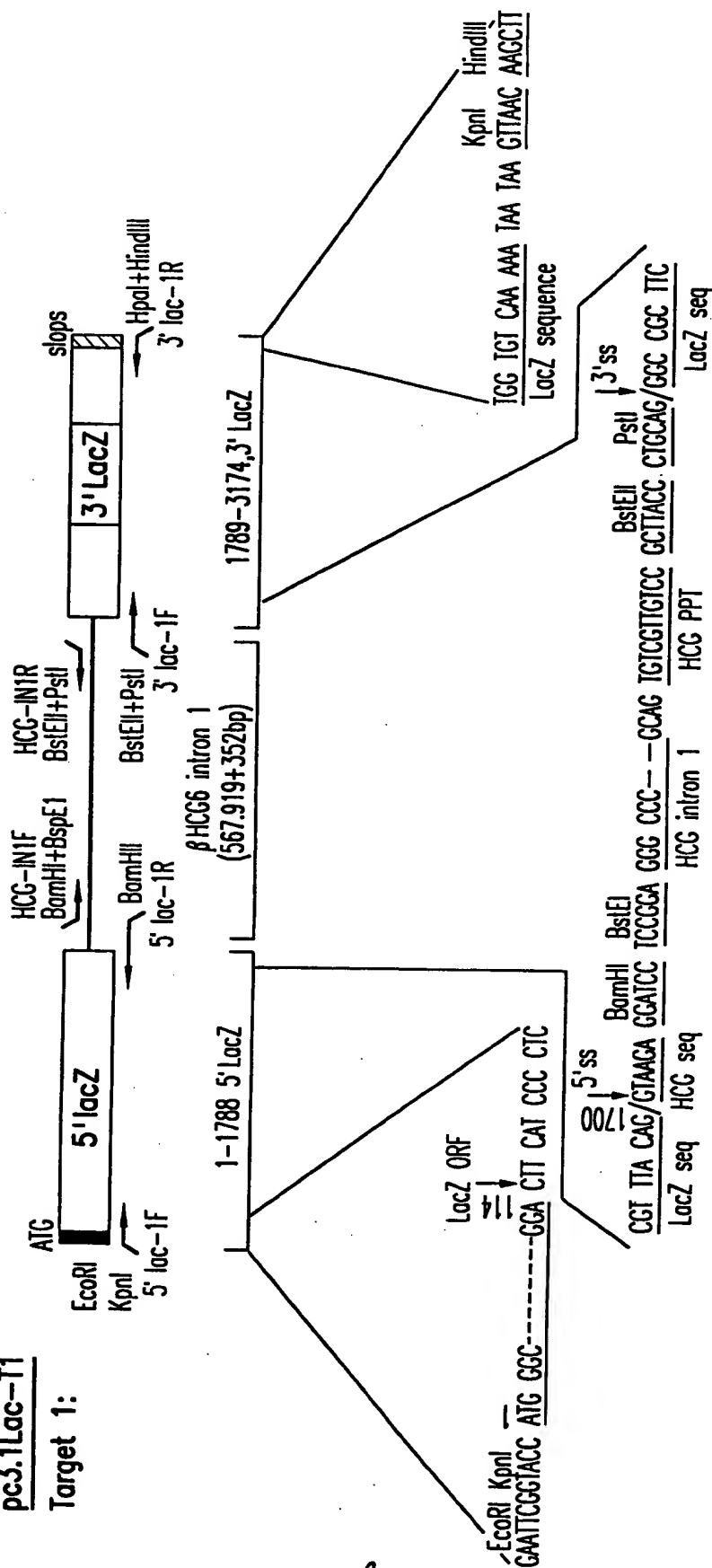


FIG.9

pc3.1Lac-T1

Target 1:



PTMs

pc3.1PTM2:

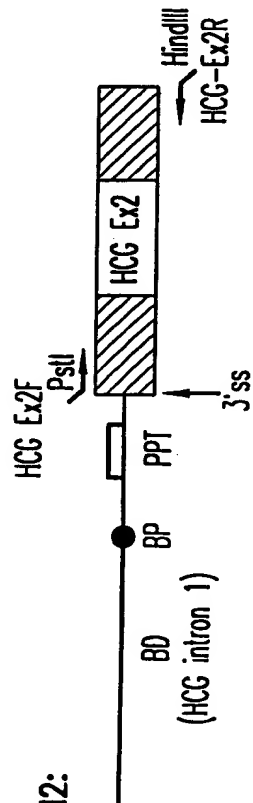
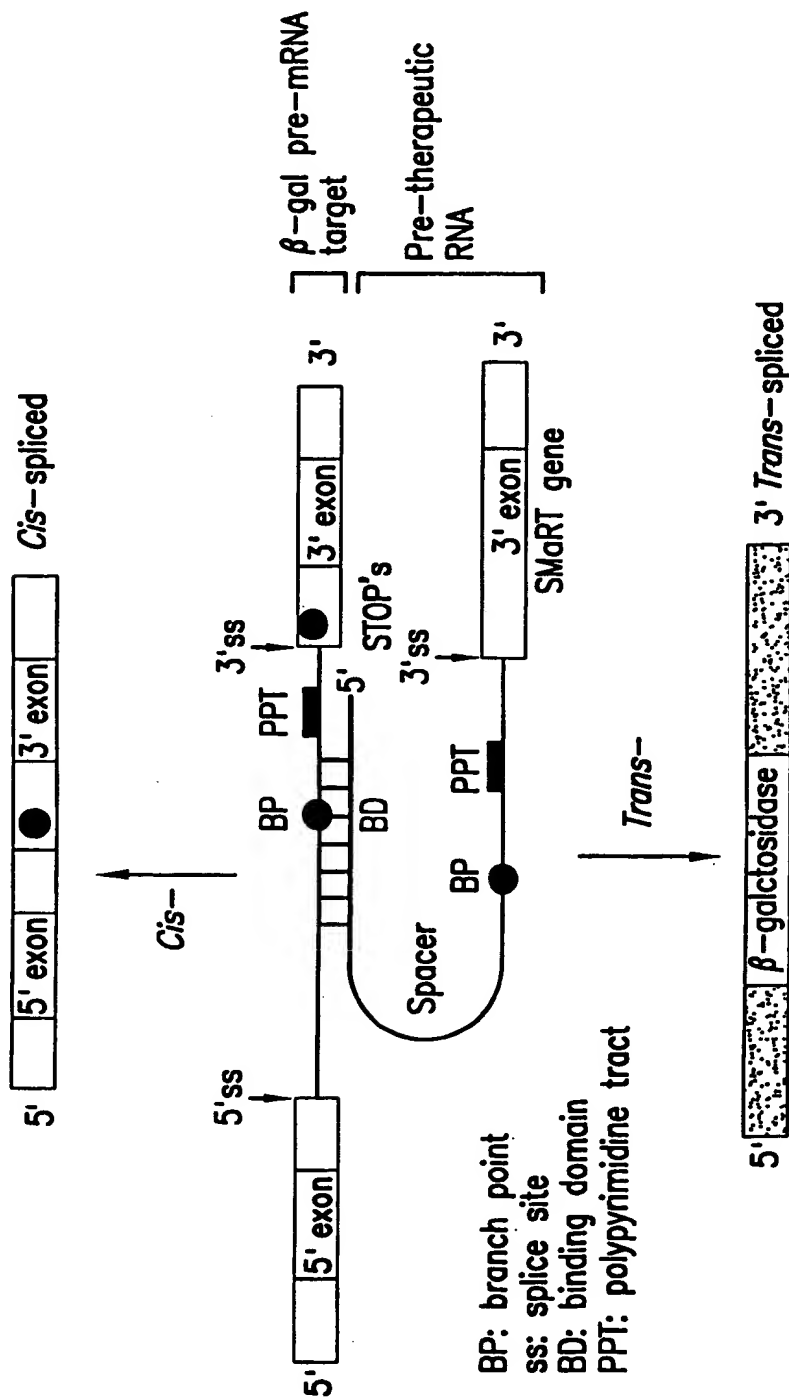


FIG.10A

15 of 89



68 2 71

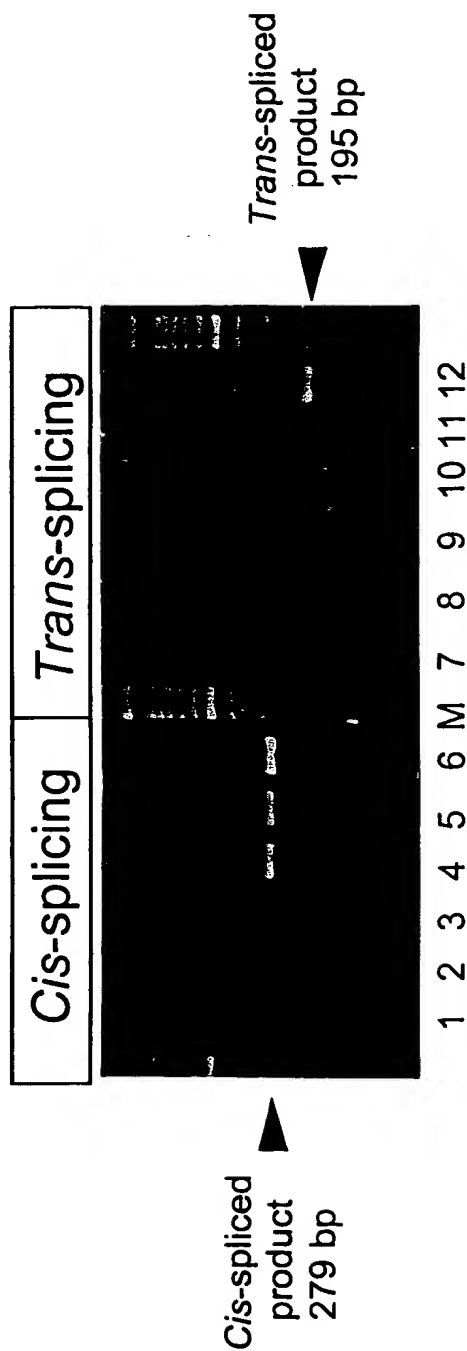


FIG.11A

094492082901

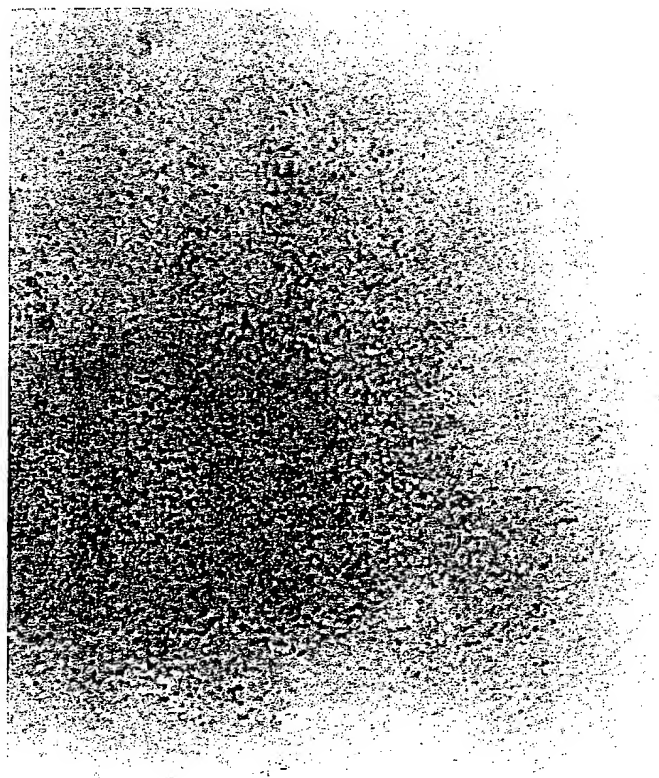


FIG.11B

19 of 89

09041492-082901

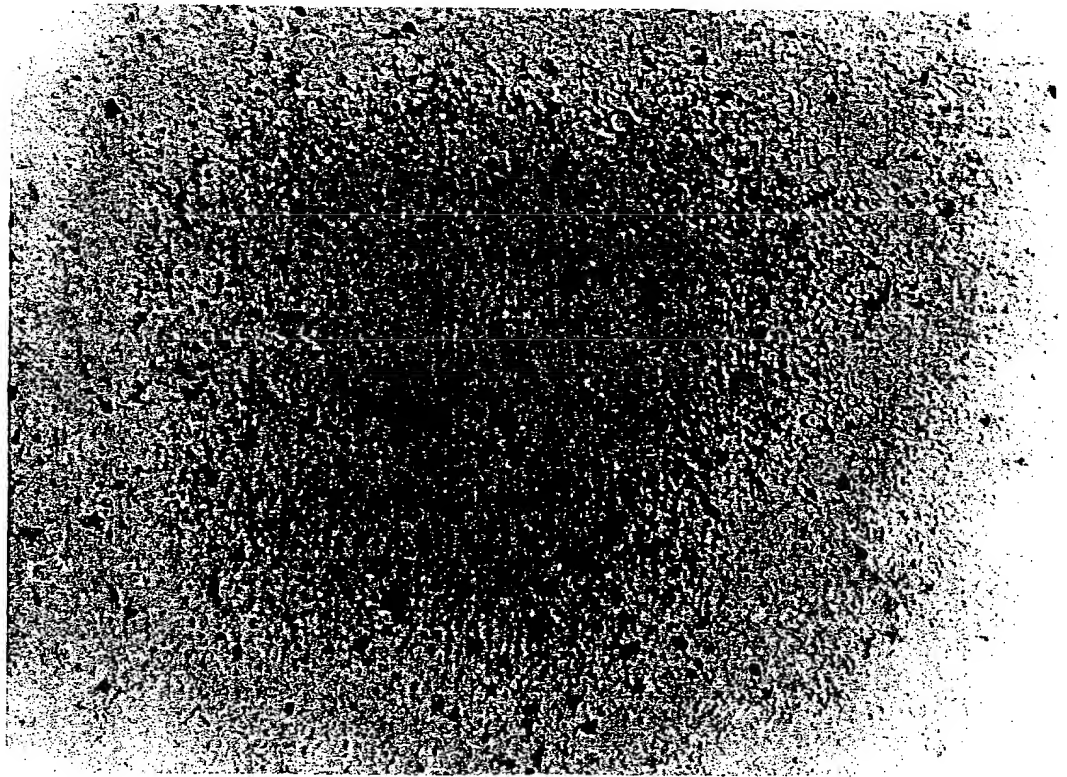


FIG.11C

094149.082901  
FO6280"264T4650

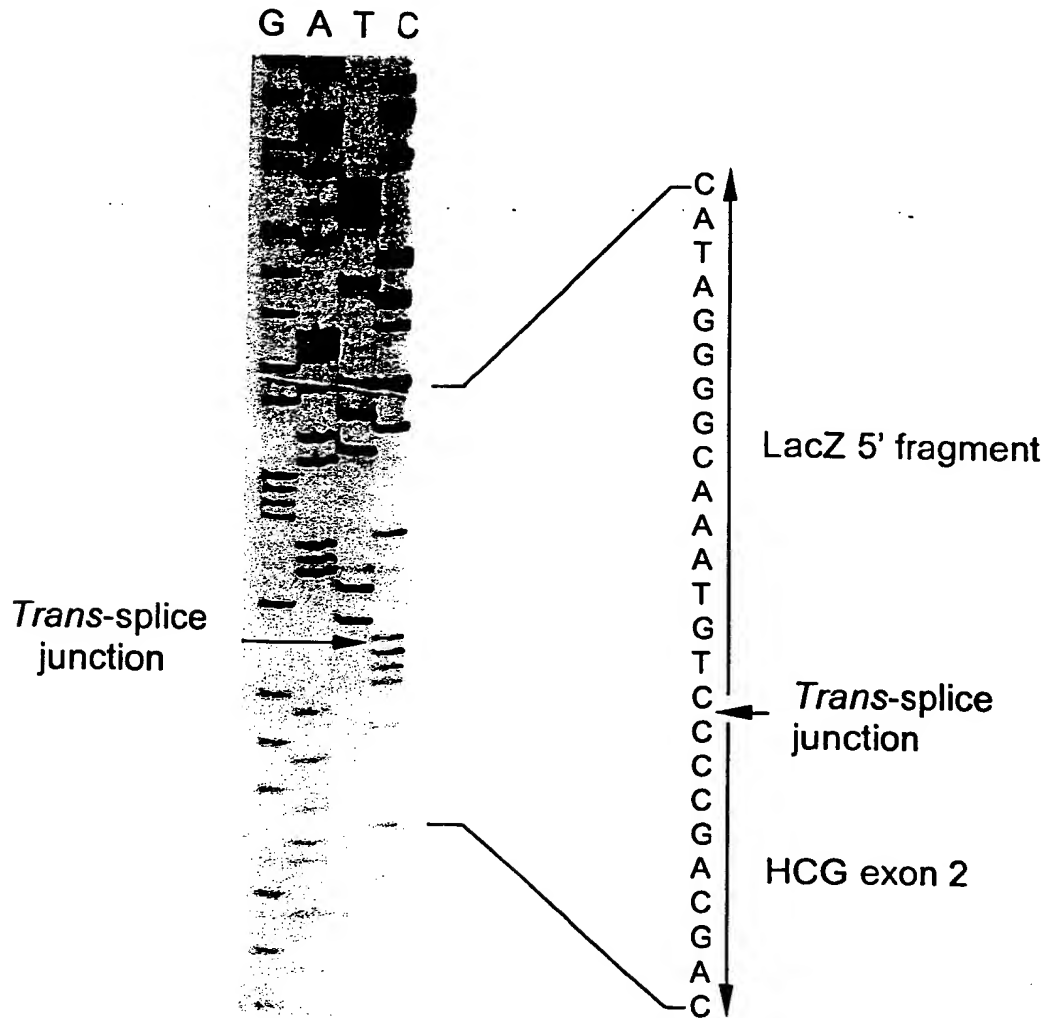


FIG.12A

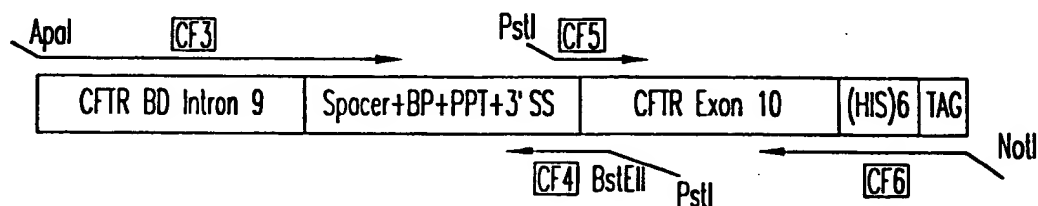


1. NUCLEOTIDE SEQUENCES OF THE *cis*-SPICED PRODUCT (285 bp):  
 BioLac-TR1  
GGCTTCGCTACCTGGAGAGACGCGCGCGCTGATCCTTTGCGAATACGCCCCACGCGATGGGTACAGTCTTG  
 GCGGTTTCGCTAAATACITGCCAGGCGTTTCGTCAGTATCCCGTTTACAG/GGCGGCTTCGCTAATAATG  
 GGACTGGGTGGATCAGTCGCTGATTAAATAATGATGAACGCGCAACCGTGGTTCGGCTTACGGCGGTGATTT  
 TGGCGATACGCGGAACGATCGCCAGTTCTGTATGAACGGTCTGGTCTTTGCGACCGCACCGCATCCAG  
 Lac-TR2  
 GCGCTTCGCTACCTGGAGAGACGCGCGCGCTGATCCTTTGCGAATACGCCCCACGCGATGGGTACAGTCTTG  
 CCGTTCGCTAAATACITGCCAGGCGTTTCGTCAGTATCCCGTTTACAG/GGCGTTCGCTTCGCTGCTGCT  
 Splice junction  
 HGR2  
GAGCATGGGCGGACATGGGCATCCAAGGAGCCACTTCGGCCACGGTCCCG

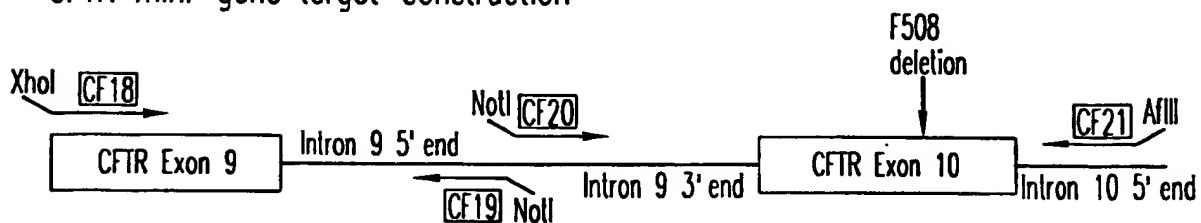
2. NUCLEOTIDE SEQUENCES OF THE *trans*-SPICED PRODUCT (195 bp)  
 BioLac-TR1  
GGCTTCGCTACCTGGAGAGACGCGCGCGCTGATCCTTTGCGAATACGCCCCACGCGATGGGTACAGTCTTG  
 CCGTTCGCTAAATACITGCCAGGCGTTTCGTCAGTATCCCGTTTACAG/GGCGTTCGCTTCGCTGCTGCTGCT  
 Splice junction  
 HGR2  
GAGCATGGGCGGACATGGGCATCCAAGGAGCCACTTCGGCCACGGTCCCG

FIG.12B

CFTR Pre-therapeutic molecule (PTM or "bullet")



## CFTR mini-gene target-construction



## Trans-splicing Repair

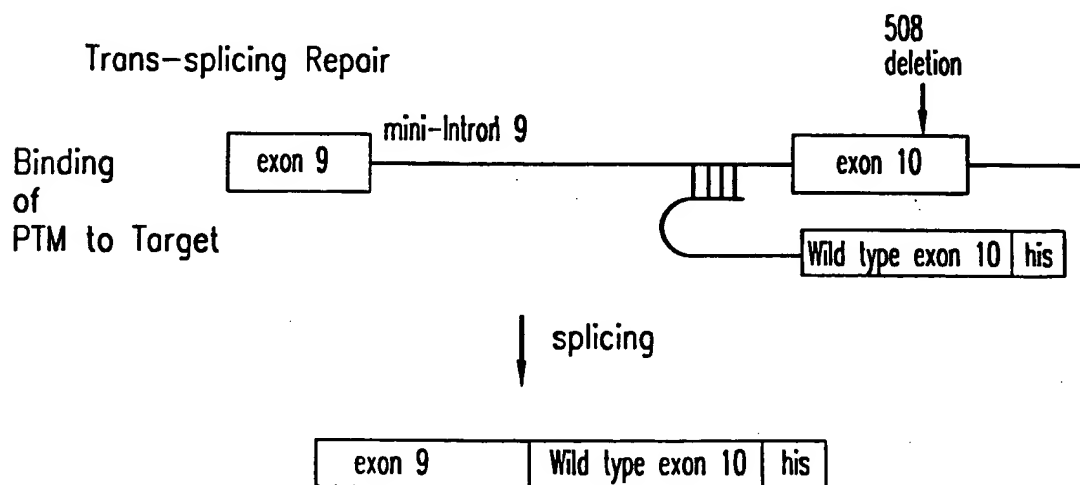
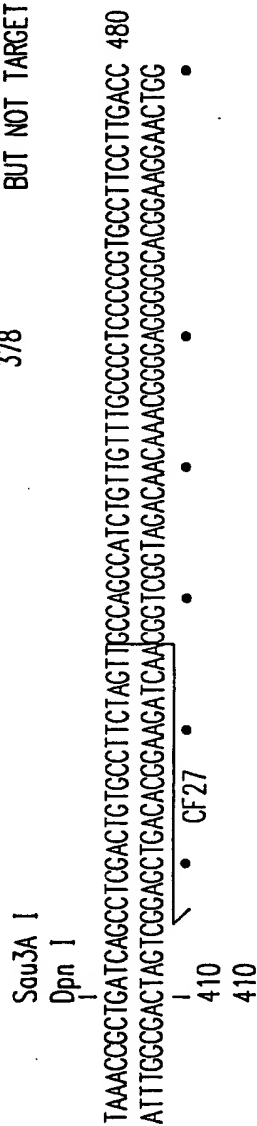
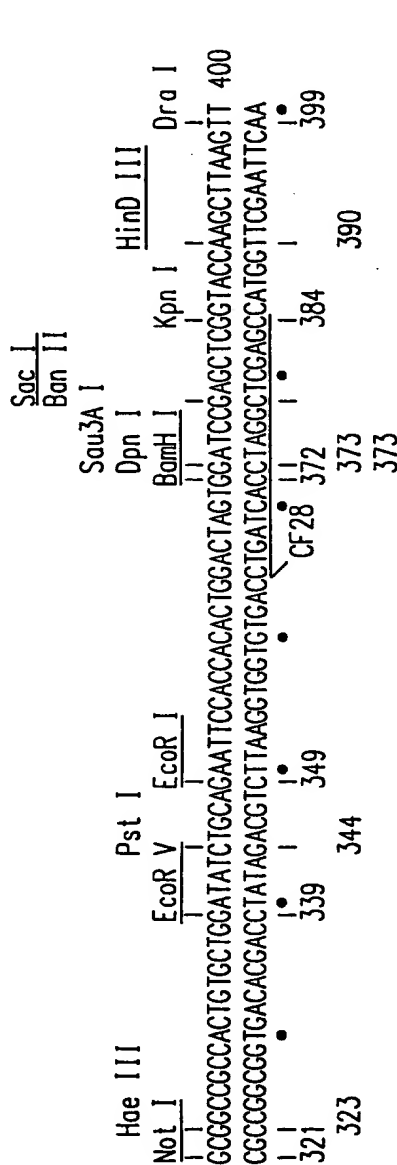


FIG. 13







CTGGAAGGTGCCACTCCAC 500

GACCTTCCACGGTGAGGGTG

Restriction Endonucleases site usage

Acc I	-	EcoR I	1	Nde I	-	Sau96 I	2
Apa I	1	EcoR V	1	Nhe I	1	Sca I	1
Apal I	-	Hae II	-	Not I	1	Sma I	-
Avr II	-	Hae III	2	PfIM I	-	Sph I	1
BamH I	1	HinC II	-	Pst I	2	Spl I	-
Ban II	2	HinD III	1	Pvu I	-	Ssp I	-
Bbe I	-	Hinf I	-	Pvu II	-	Stu I	-

FIG.15B



+

F508 deletion

CFTR Target  
(mini-gene)

CFTR Exons 1-9

CFTR Exons 10-24

Mini-intron 9  
(~0.6kb)

Cotransfect PTM and target molecules in HEK 293 cells  
and detect repaired CFTR mRNA by RT-PCR.

Repaired  
CFTR mRNA

CFTR Exons 1-9

Exons 10-24 CFTR

(His) 6 TAG

FIG.16

20250420 20250420

Double Splicing  
PTM

CFTR BD intron 9	Spacer+BP+PPT+3'SS CFTR exon 10	Spacer+BP+PPT+5'SS CFTR BD intron 10
------------------	---------------------------------	--------------------------------------

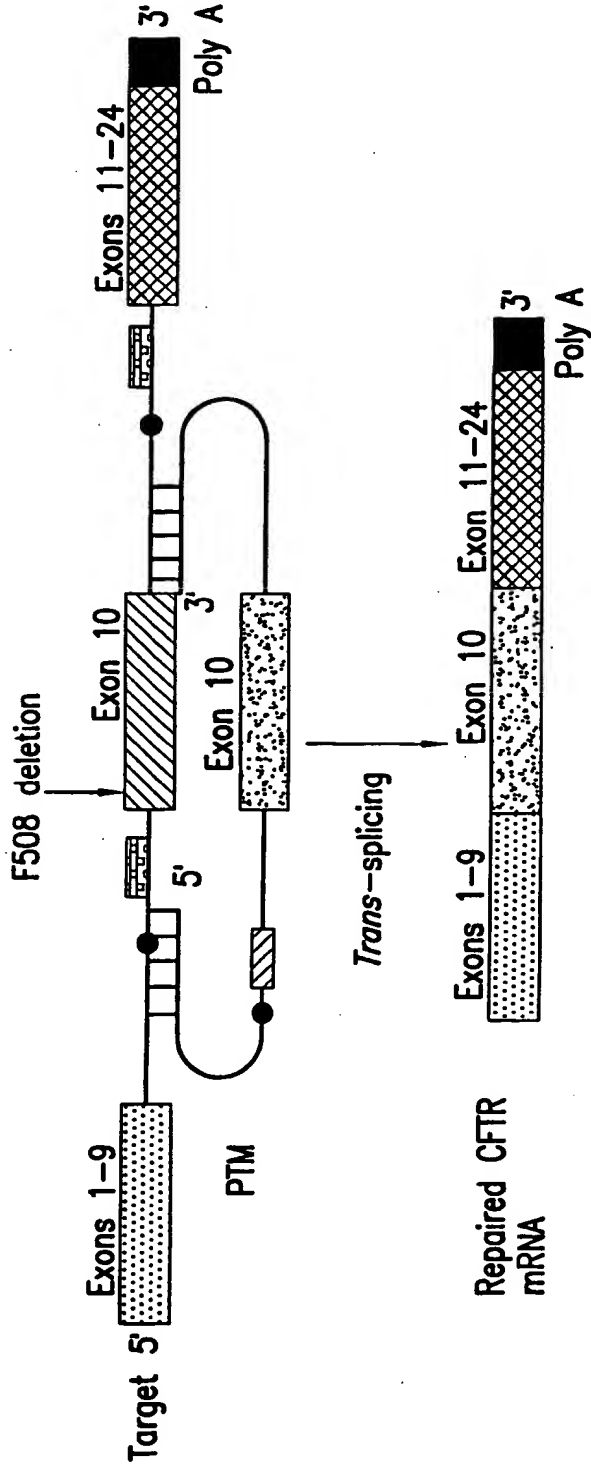


FIG.17

# Double Trans-splicing Specific Target

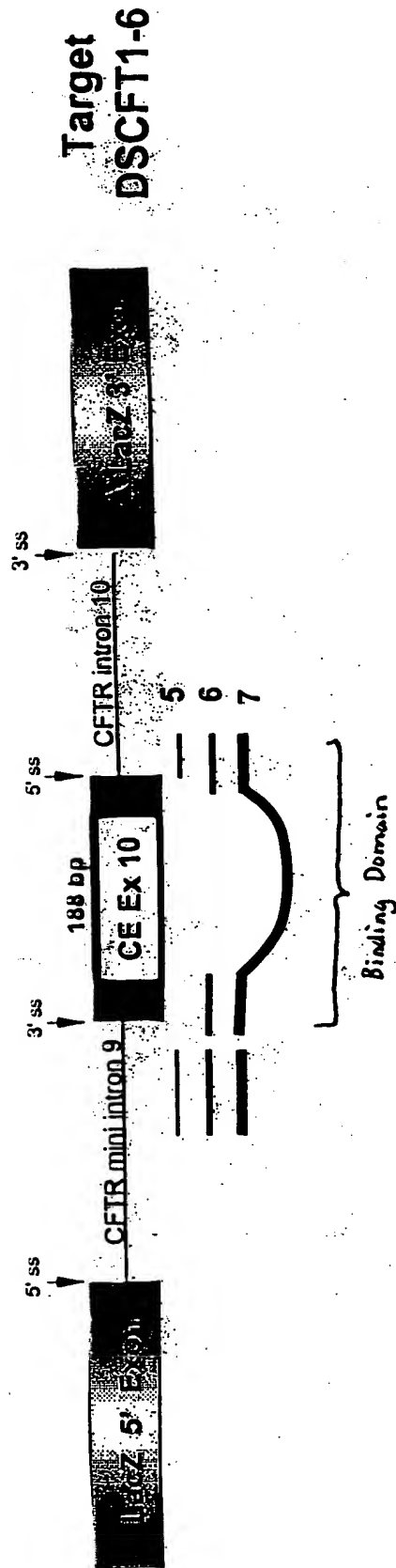


Figure 18



## Double Splicing PTMs



**DSP™-5**

**DSP™-6**

**DSPTM-7**

**PTM with 260 bp BD  
masking both the ss &  
the entire CFTR Ex10**

# Double Trans-splicing $\beta$ -Gal Model

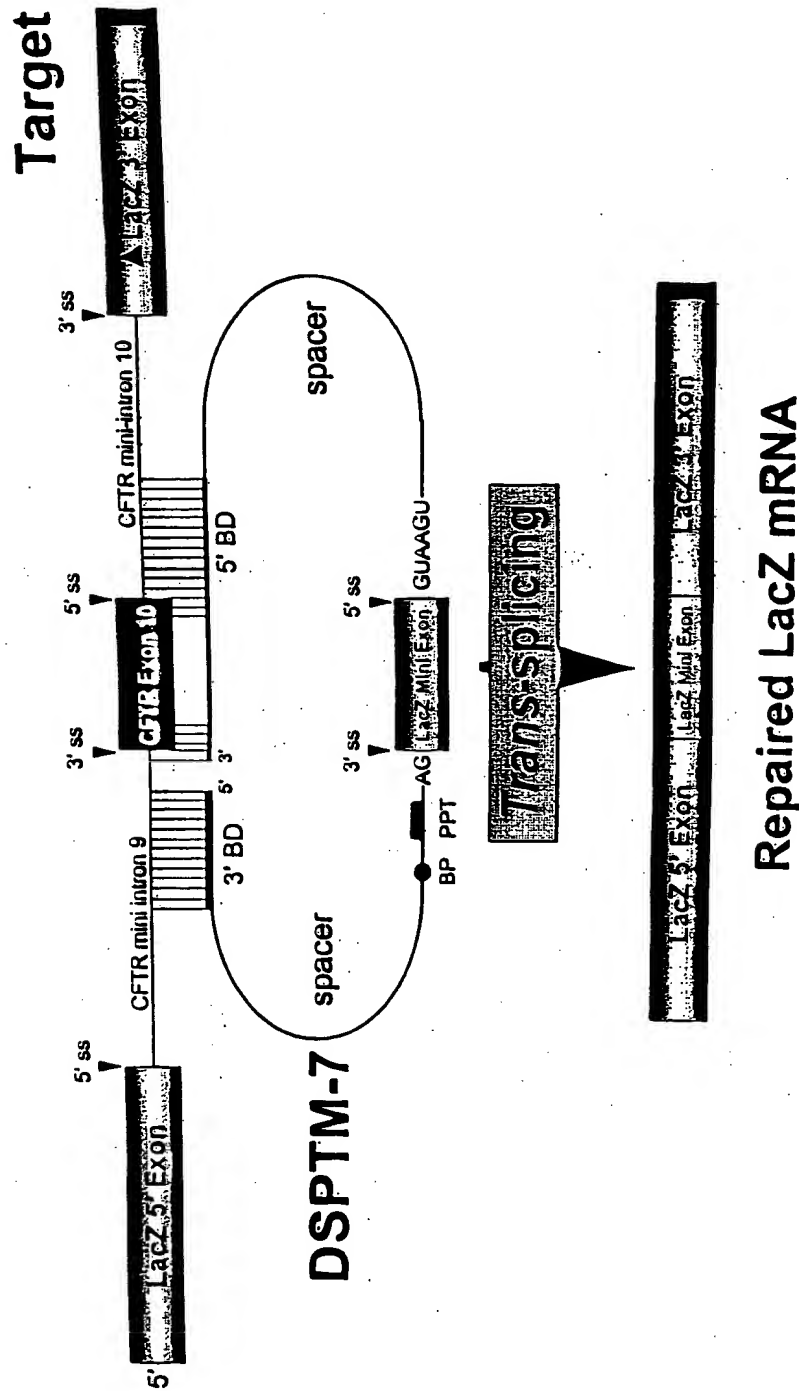
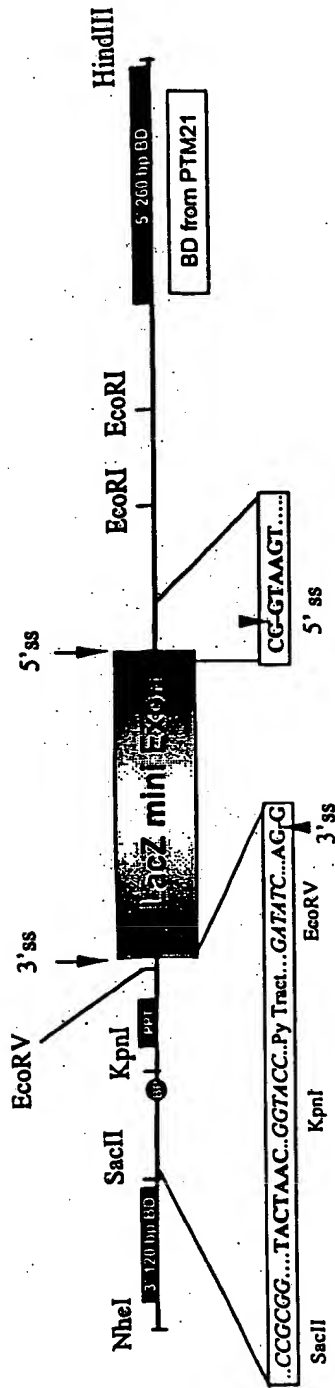


Figure 20

# Important Structural Elements of DSPTM-7: (Double splicing PTM with all the necessary splice elements i.e. has both 3' and 5' functional splice sites and the binding domains)



(1) 3' BD (120 BP) : GATTCACCTTGCTCCAAATTATCATCCTAAGCAGAAGTGATATCTTATTGTAAAGATTCTATTAACTCATTGATTC  
AAAATATTAAATACTTCCTGTTTCATACTCTGCTATGCAC

(2) Spacer sequences (24 bp): AACATTATTATAACGTTGCTCGAA

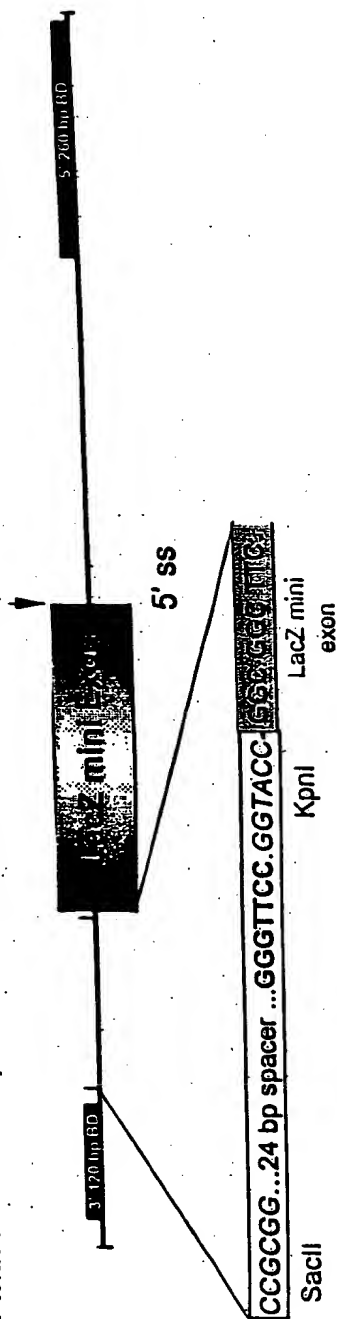
(3) Branch point, pyrimidine tract and acceptor splice site: TACTAAC T GGTACC TCTTCTTTTTTTTTT GATATC CTGCAG GGCGGG  
BP KpnI PPT EcoRV LacZ mini exon

(4) 5' donor site and 2<sup>nd</sup> spacer sequence: CTAAGG GTAAAGT GTTATCACCAGATATGTGCTAACCTGATTCTGGGCTTCGATACG  
5' ss LacZ mini exon  
CTAAGATCCACCGG

(5) 5' BD (260 BP) : TCAAAAAGTTTTCACATAATTTCTTACCTCTTCTTGAA77CATGCTTTGATGACGCTTCTGTATCTATATTCATTGGAA  
ACACCAATGATTTTCTTTAATGGTGCTGGCCTGGCATAATCCTGGAAAACGTGATAACACAATGAAATCTTCCACTGTGCTTAA  
AAAAACCCCTCTGAA77CTCCATTCTCCCATATCATCATTAACACTGAACCTCTGGAAATAAAACCCATCATATTAACTCA  
TTATCAAAATCACGC

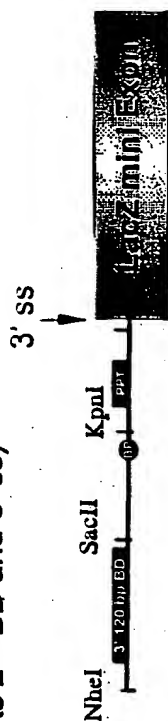
Figure 21

**DSPTM8** : (▲ 3' ss: 3' splice elements i.e. BP, PPT & AG dinucleotide has been deleted and replaced with random sequences, but still has the functional 5' splice site)



Mutants

**PTM29** (lacks 2<sup>nd</sup> BD and 5' ss)



**PTM30** (lacks 1<sup>st</sup> BD and 3' ss)



Figure 22

# Accuracy of Double Trans-splicing Reaction

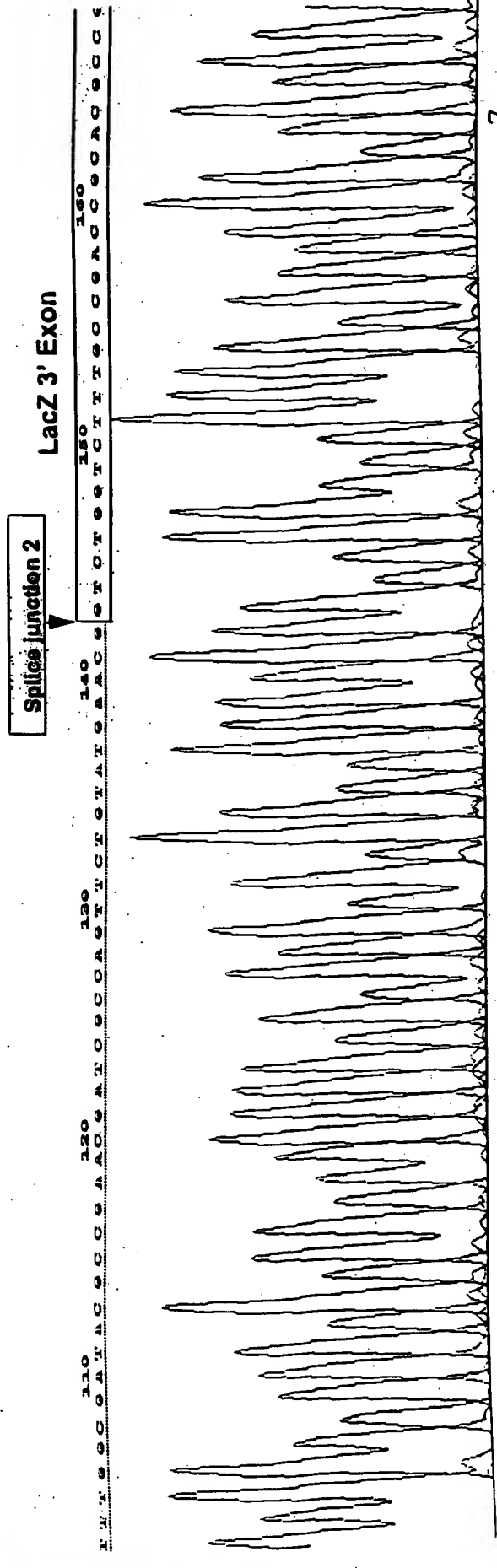
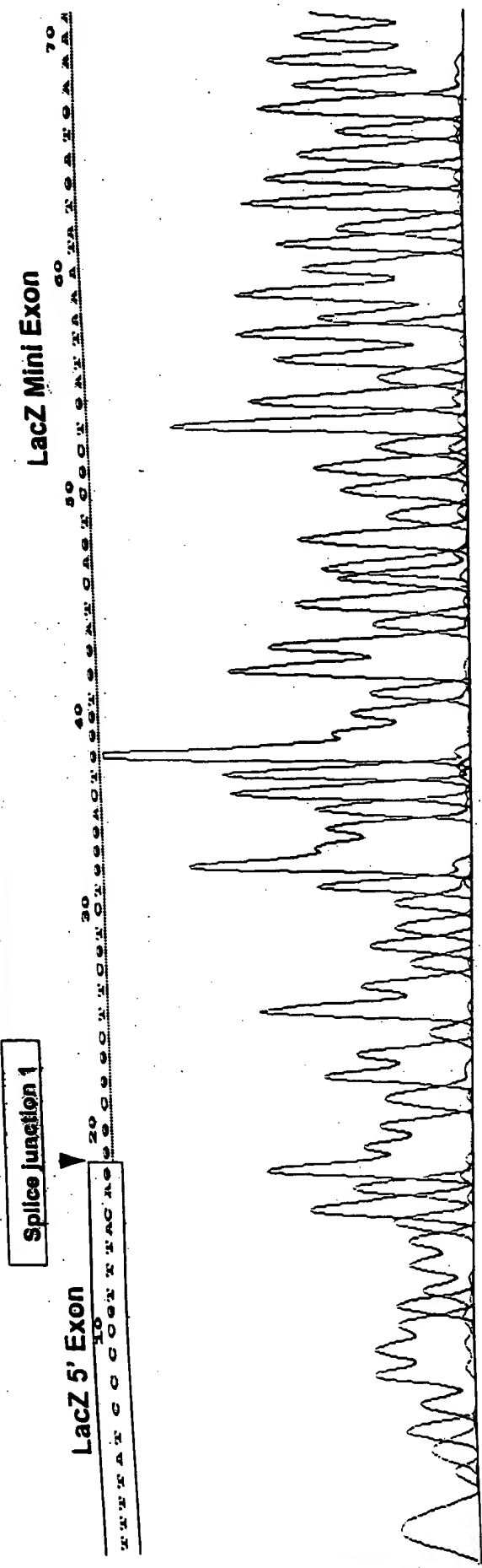


Figure 23

# Double Trans-splicing Produces Full-length Protein



β-gal →  
(120 kDa)

1 2 3 4 5 6 7

Lane 1: DSCFT1.6 Target alone 25 μg  
 Lane 2: DSPTM7 25 μg  
 Lane 3 Target + PTM #6 25 μg  
 Lane 4: Target + PTM #9 25 μg  
 Lane 5: Delta 3' splice mutant alone 25 μg  
 Lane 6: Target + Delta 3' ss 25 μg  
 Lane 7: Target+PTM29+30 (mutants) 25 μg

Figure 24

# Restoration of $\beta$ -Gal Function by Double Trans-splicing

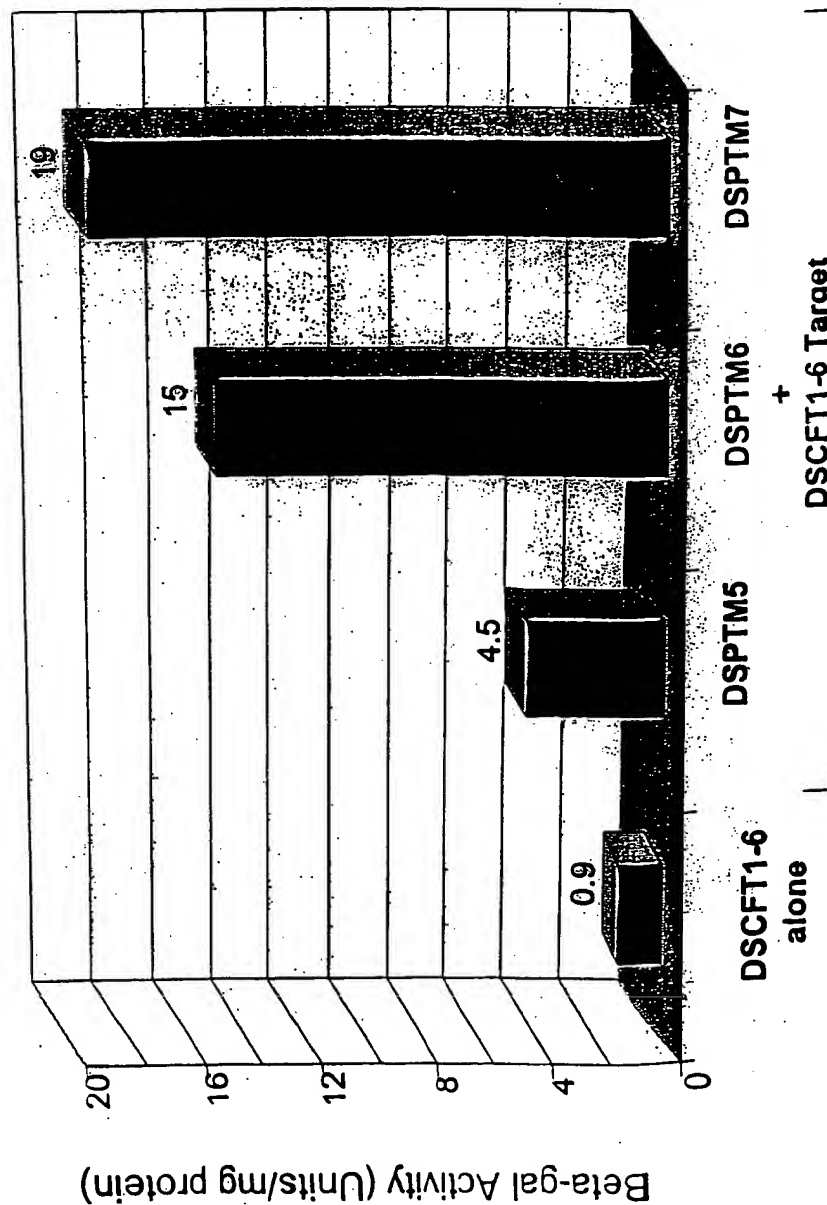


Figure 25

# Restoration of $\beta$ -gal activity is due to double RNA trans-splicing events

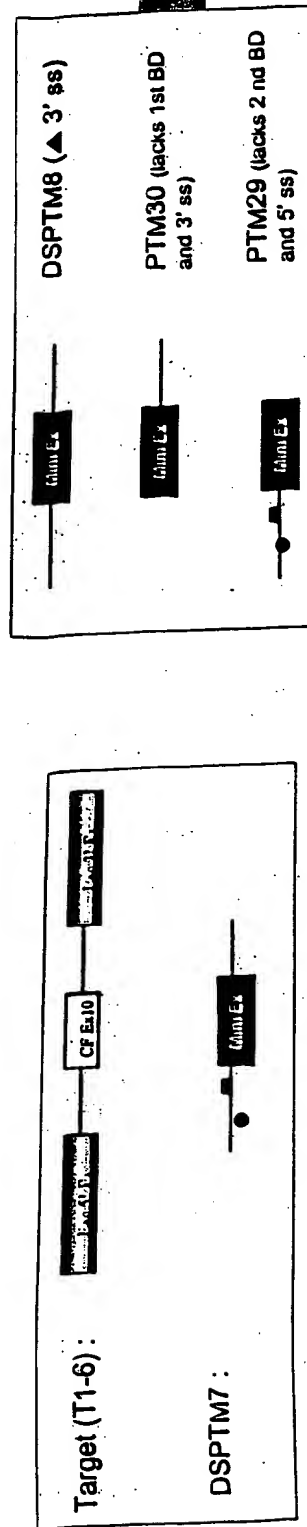
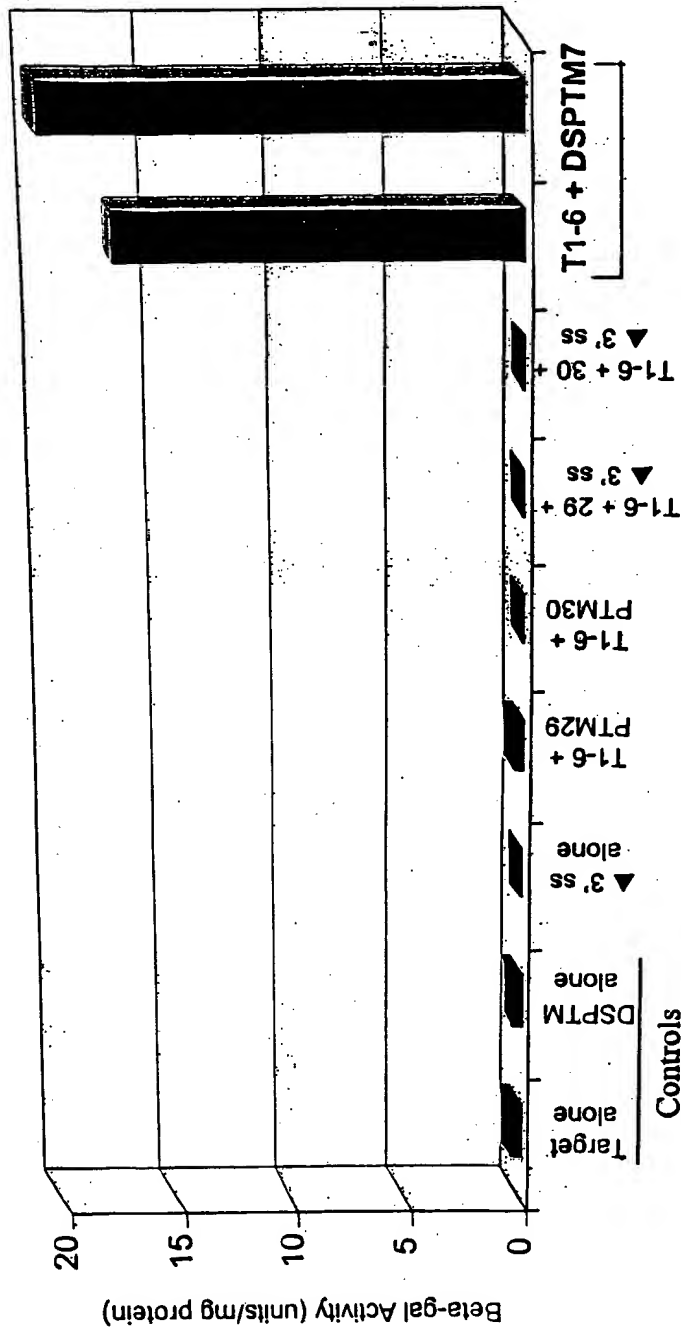
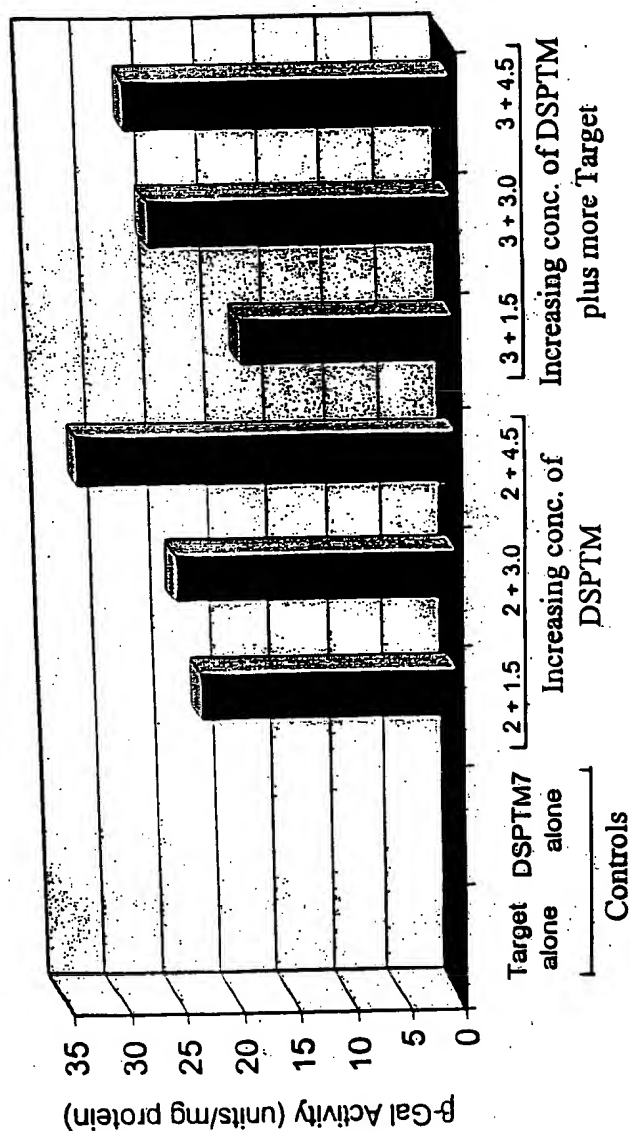


Figure 26



# Double Trans-splicing: Titration of Target & PTM



The current level of beta-gal activity due to double trans-splicing is ~ 1-1.5% of the best single splice model (3' exon replacement)

Figure 27

**DSCFT1-6 (Specific Target):**



**DSHCGT1 (Non-specific Target):**

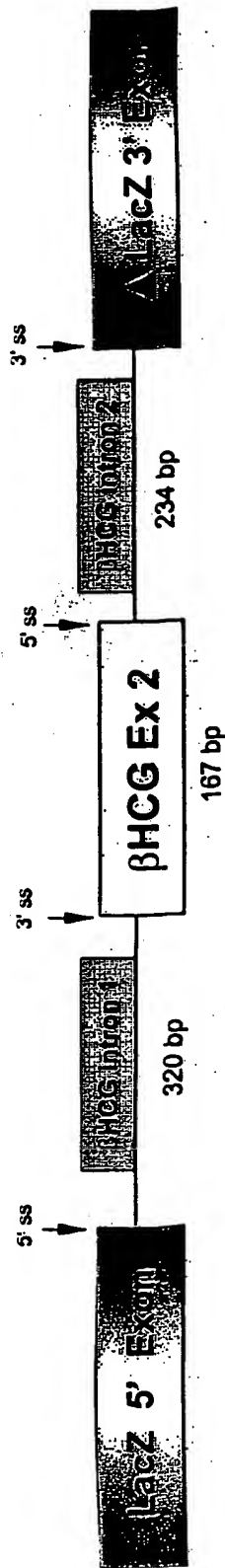


Figure 28

106280-2644650

## Specificity of double *trans*-splicing Reaction

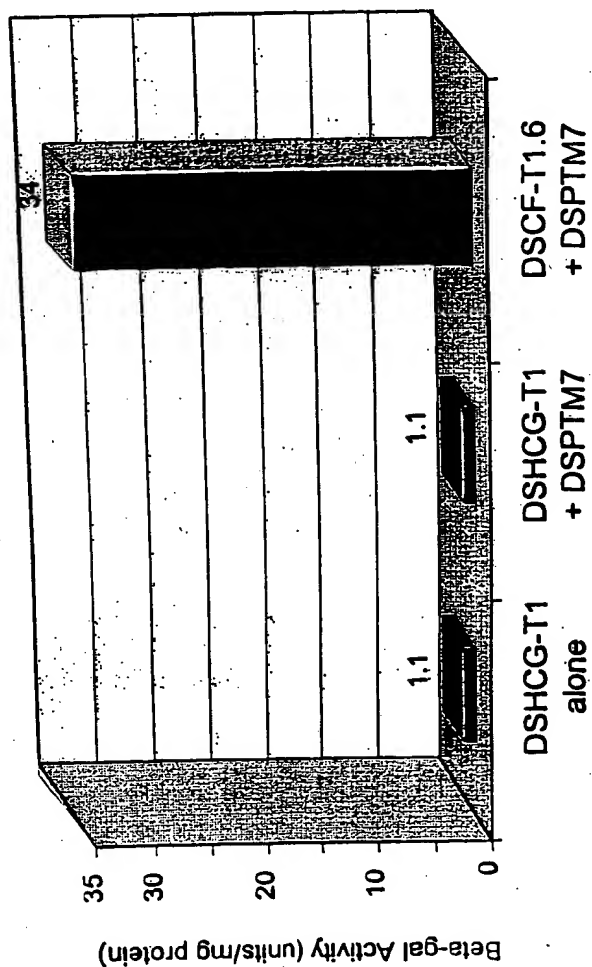


Figure 29

Repaired full length CFTR mRNA

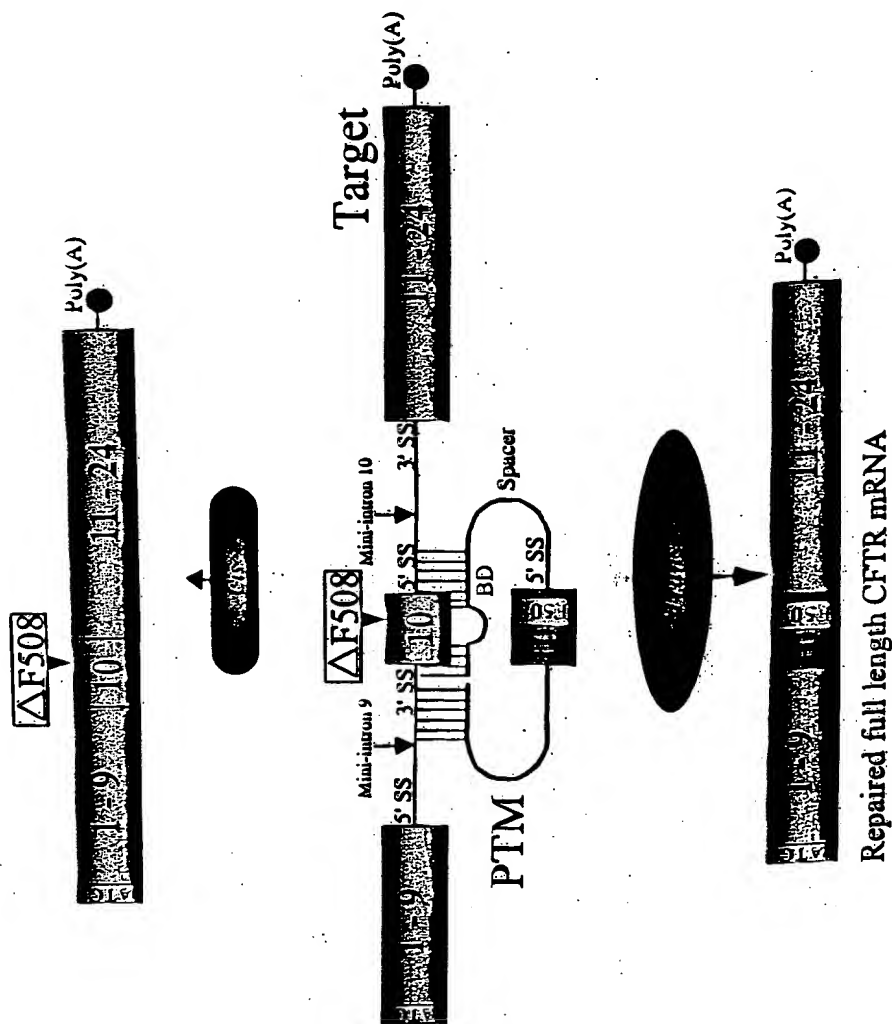
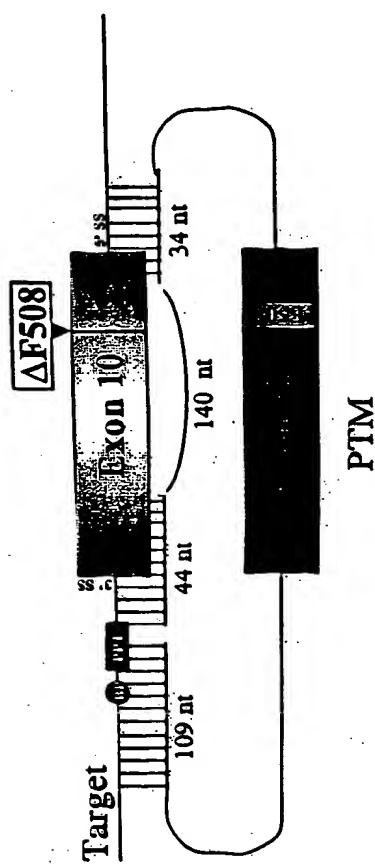


Figure 30

INTRONIN

PTM with a long binding domain masking two splice sites and part of exon 10 in a mini-gene target.



ACGAGCTTGCTCATGATCATGGCGAGTTAGAACCAAGTGAAGGCAAGATCAAAATTCCTCG  
GCCGCATCAGCTTTTCAGCCCAATTTCAGTTGGATCATGCCCGGTACCATCAAGGAGAACATAT  
CTTCGGCGTTCAGTTACGACGAGTACCGCTATCGCTCGGTGATTAAGGCCCTGTCAGTTGGAGGAG

MCU in exon 10 of PTM  
88 of 192 (46%) bases in PTM exon 10 are not complementary to its binding domain (bold and underlined).

Figure 31

INTRONN

# Sequence of a double *trans*-spliced product

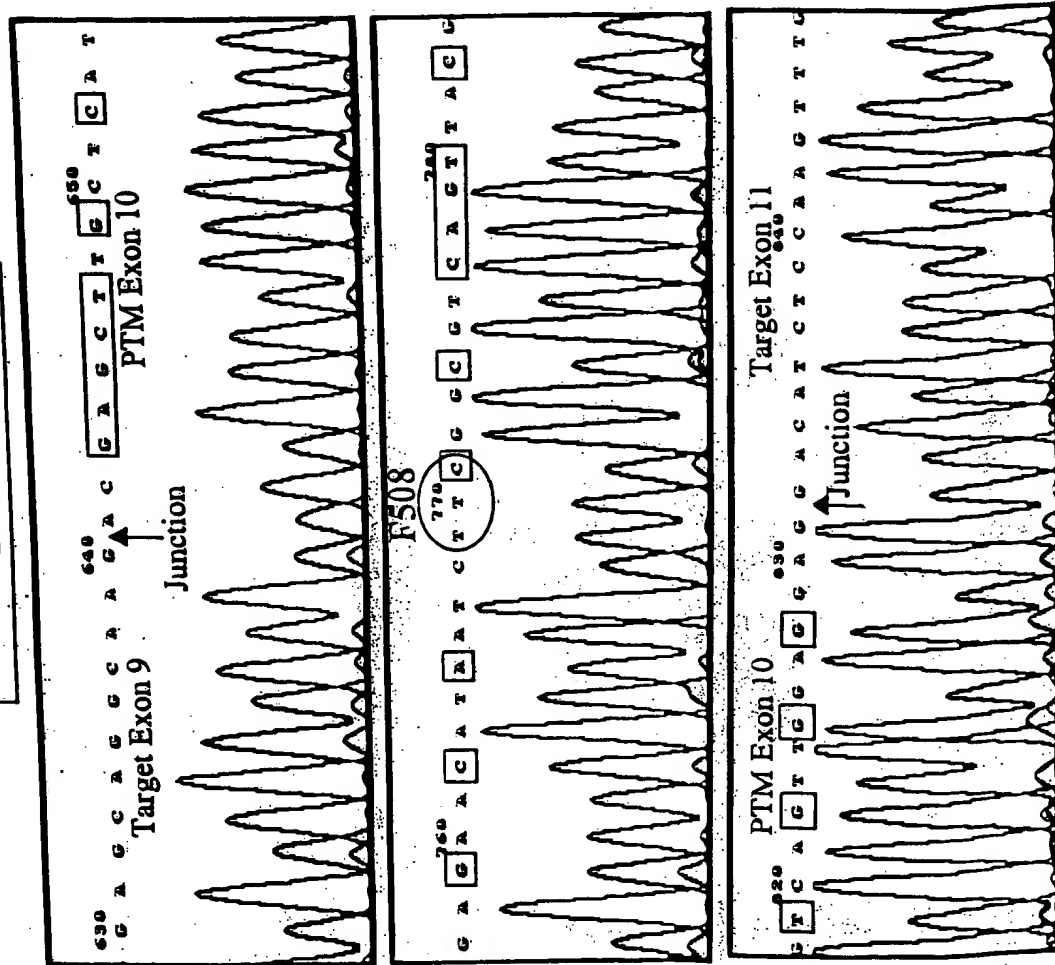


Figure 32

—

b8 fo r77

# CFTR Repair: 5' Exon Replacement

Schematic diagram of a PTM binding to the splice site of intron 10 of a mutant gene target

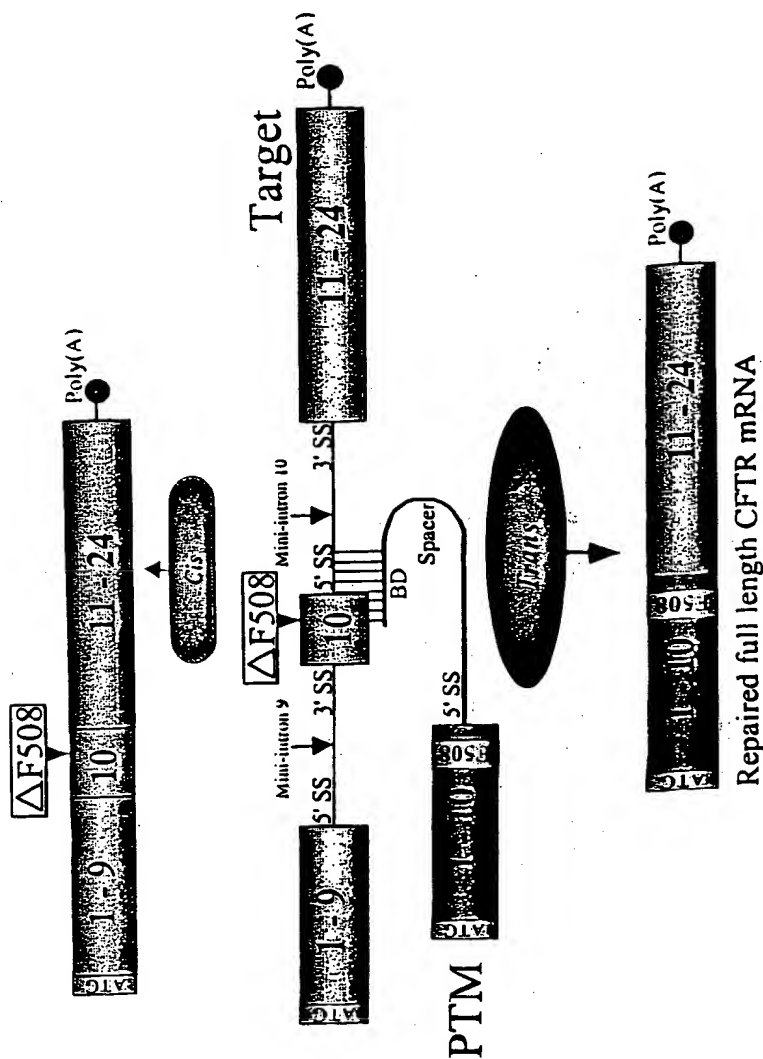
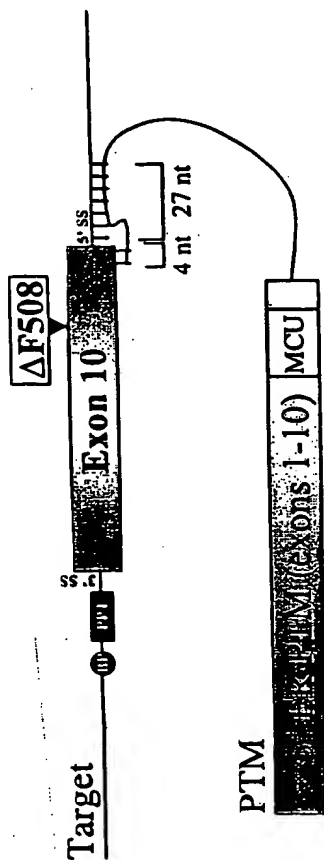
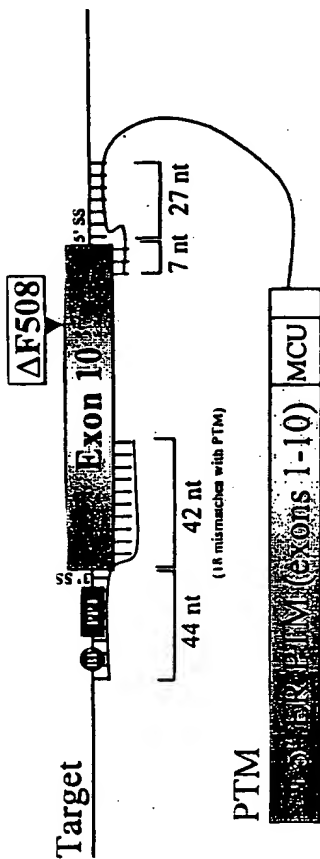


Figure 33

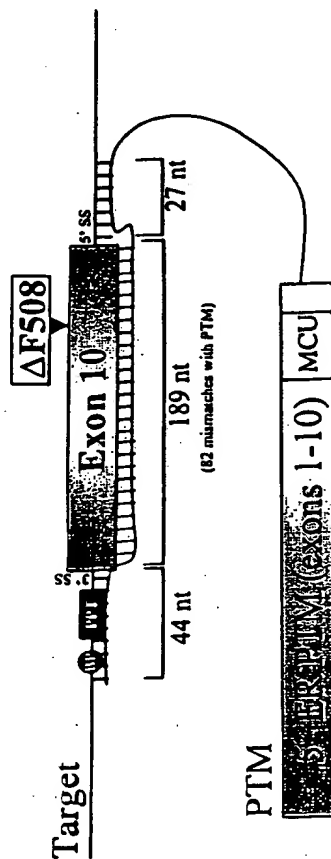
20250-264450



PTM with a short binding domain masking a single splice site in a mini-gene target.



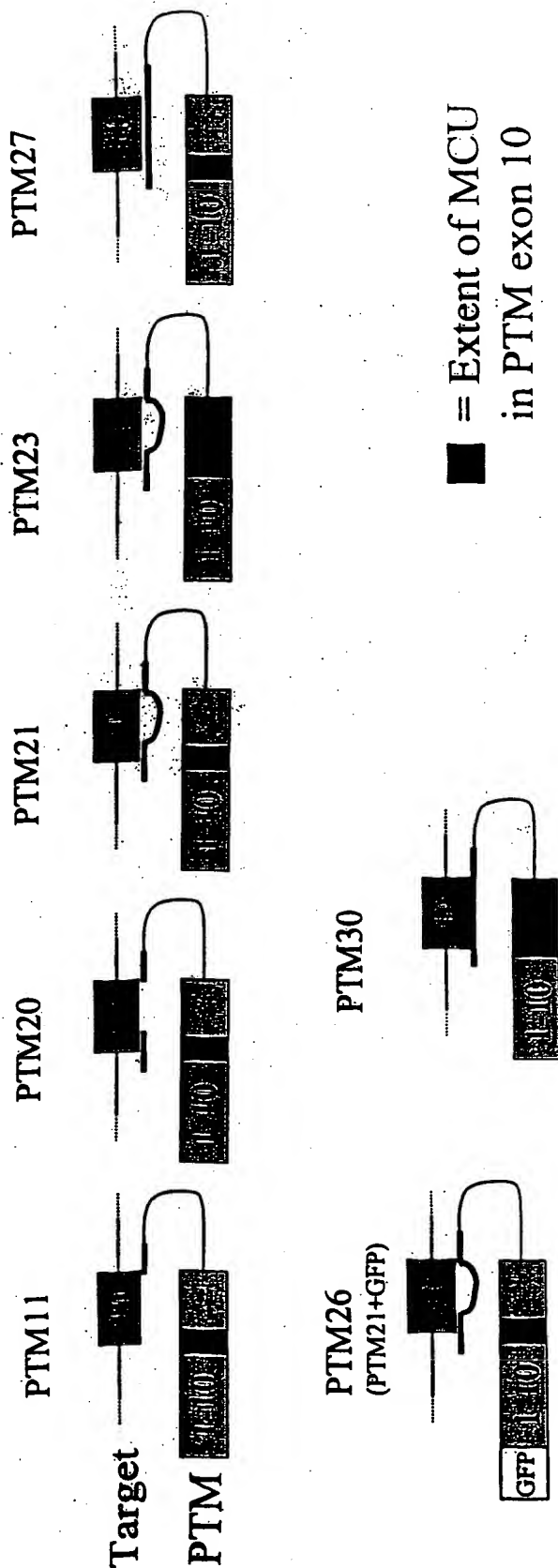
PTM with a long binding domain masking two splice sites in a mini-gene target.



PTM with a long binding domain masking two splice sites and the whole of exon 10 in a mini-gene target.

Figure 34





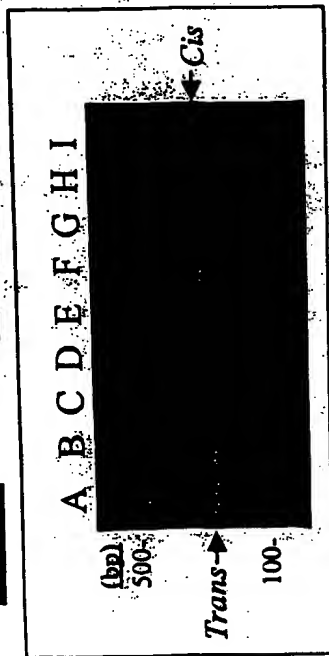
MCU in exon 10 of PTM  
88 of 192 (46%) bases in PTM exon 10 are not complementary to its binding domain.

ACGAGCTTGCTCATGATCATGGCCGAGTTAGAACCAAGTGAAGGCAAGATCAACATTCGG  
GCCGATCAGCTTTTGCAGCCAAATTCAGTTGGATGATGCCGGTACCATCAAGGAGAACATAAT  
CTTCGGGCTCAGTTACGACGAGTACCGCTATCGCTCGGTGATTAAGGCCCTGTCAGTTGGAGGAG

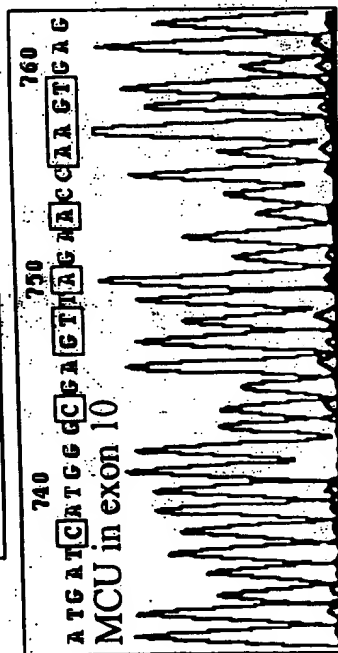
Figure 35

# INTRONIN

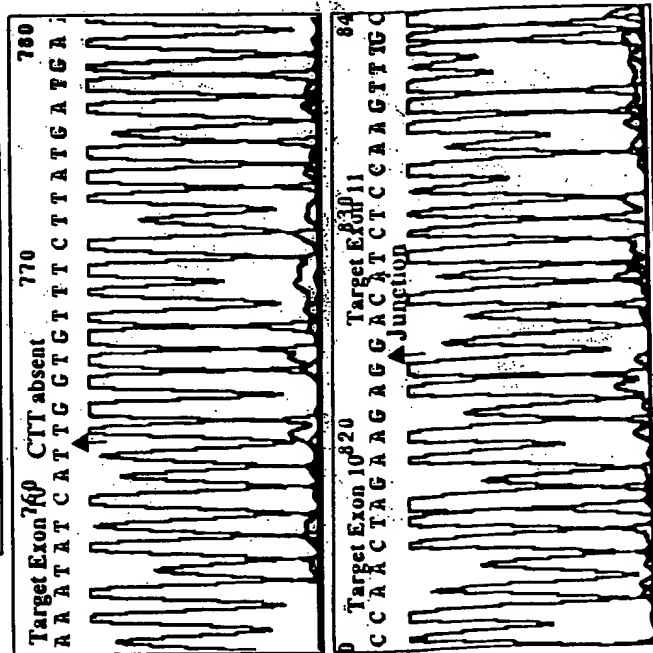
PlM Target



**B.**  
Trans-spliced product  
[Primers CF93 + CF111]



**A.**  
Cis-spliced product  
[Primers CF1 + CF111]



A

lacZCF9m

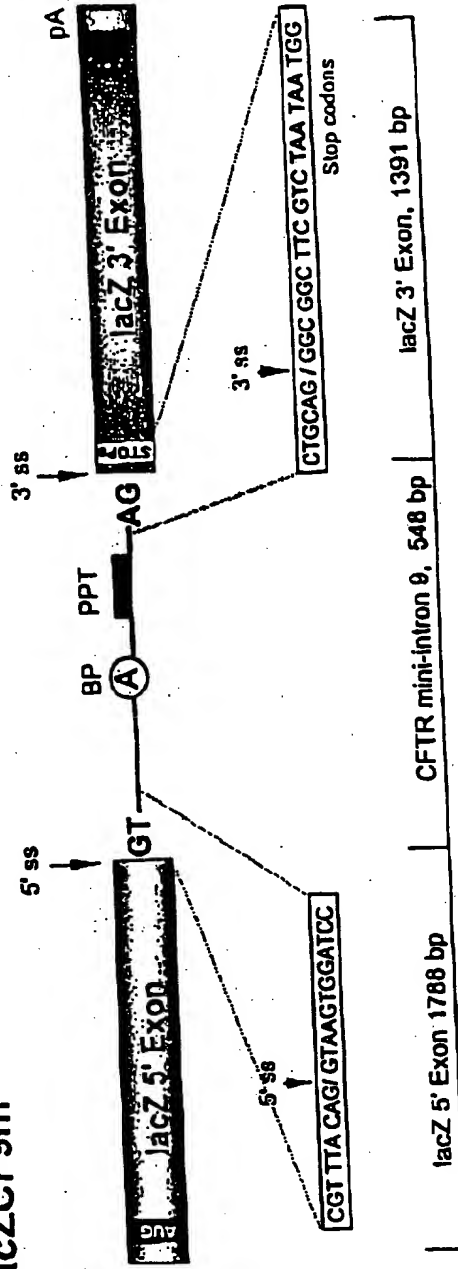


Figure 37 A

47 of 89

B

48 of 89

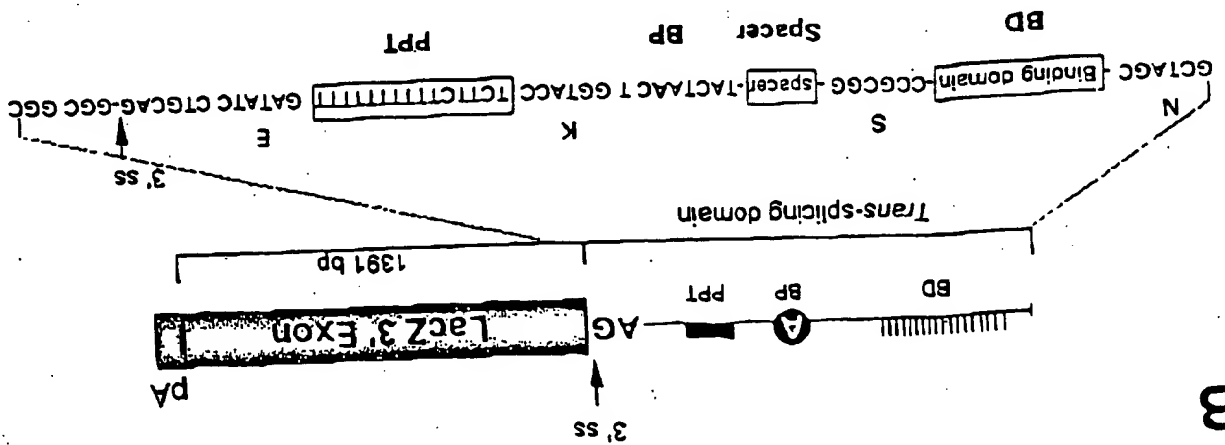
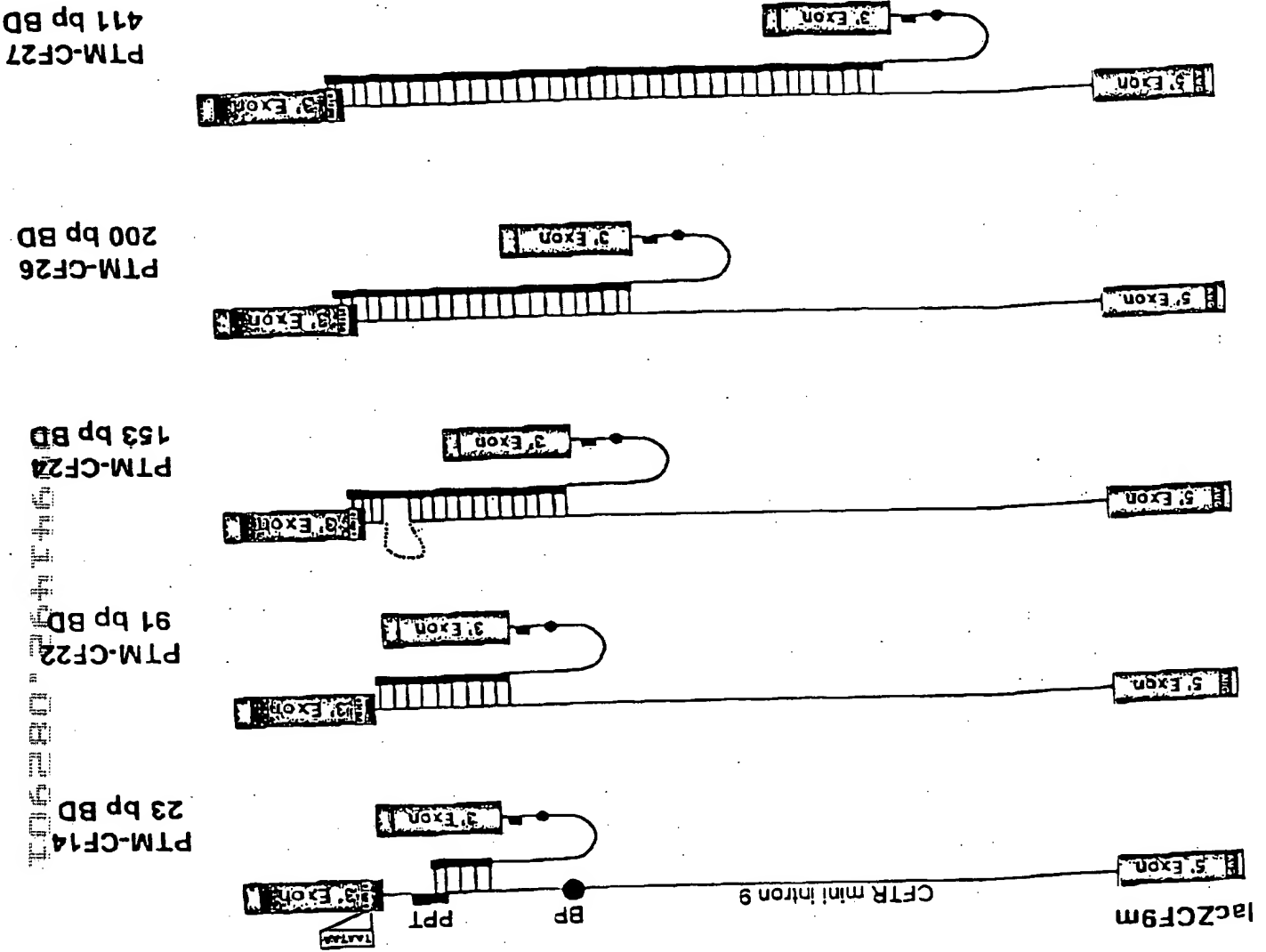


Figure 37B



C

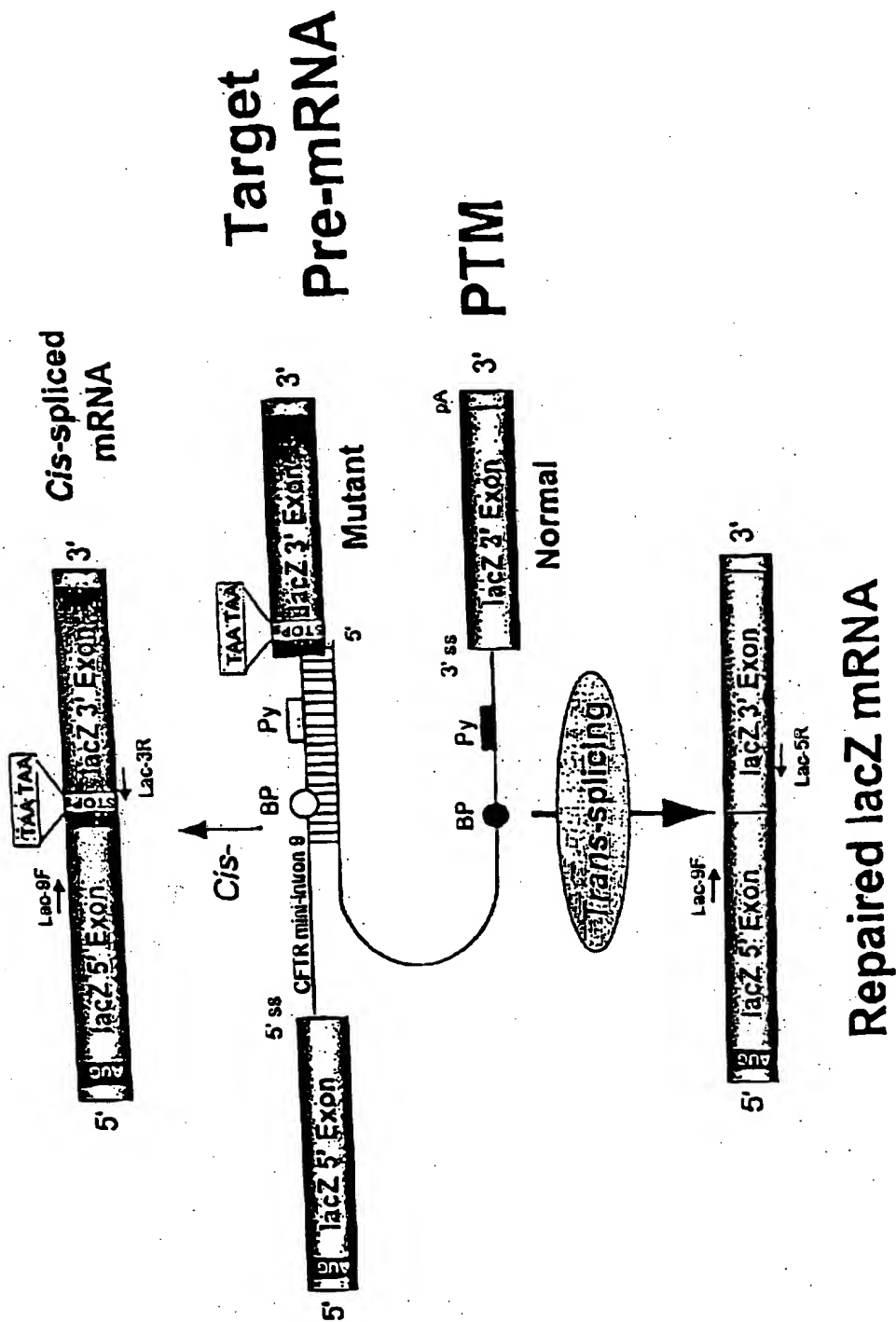


Figure 37C

A

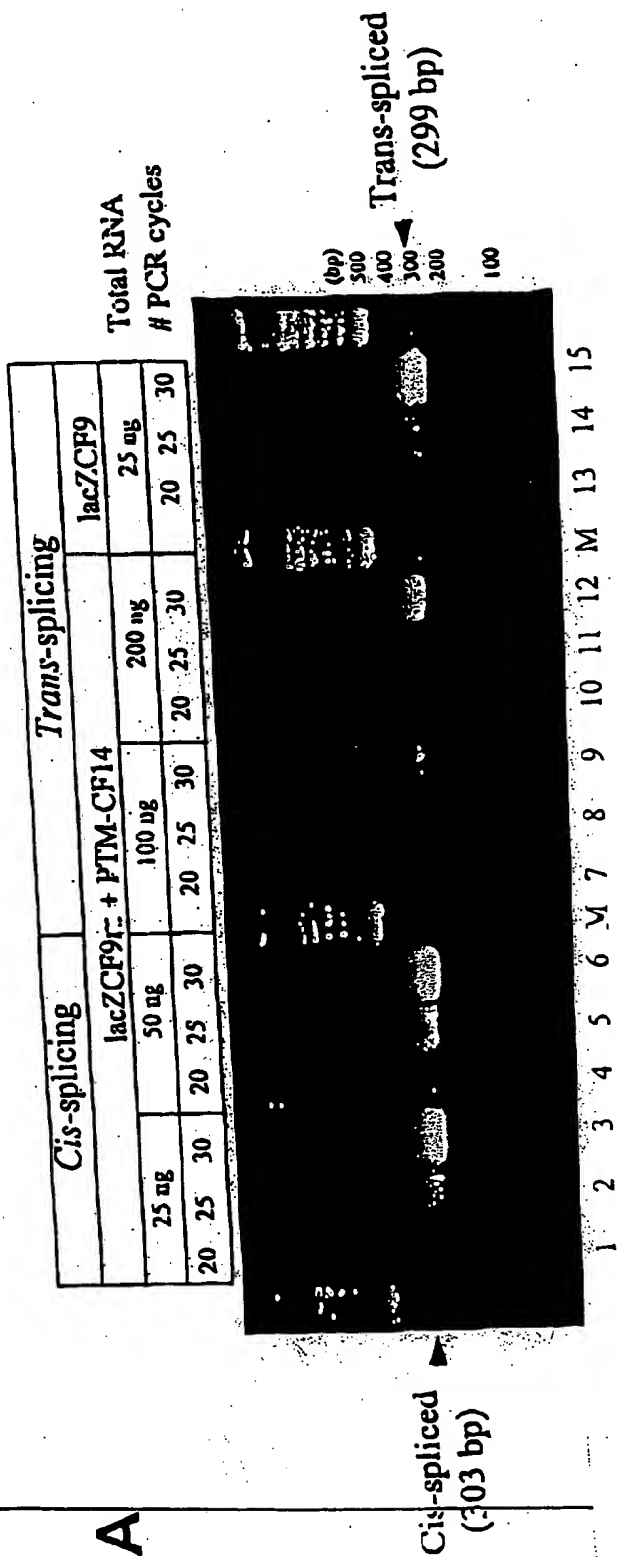
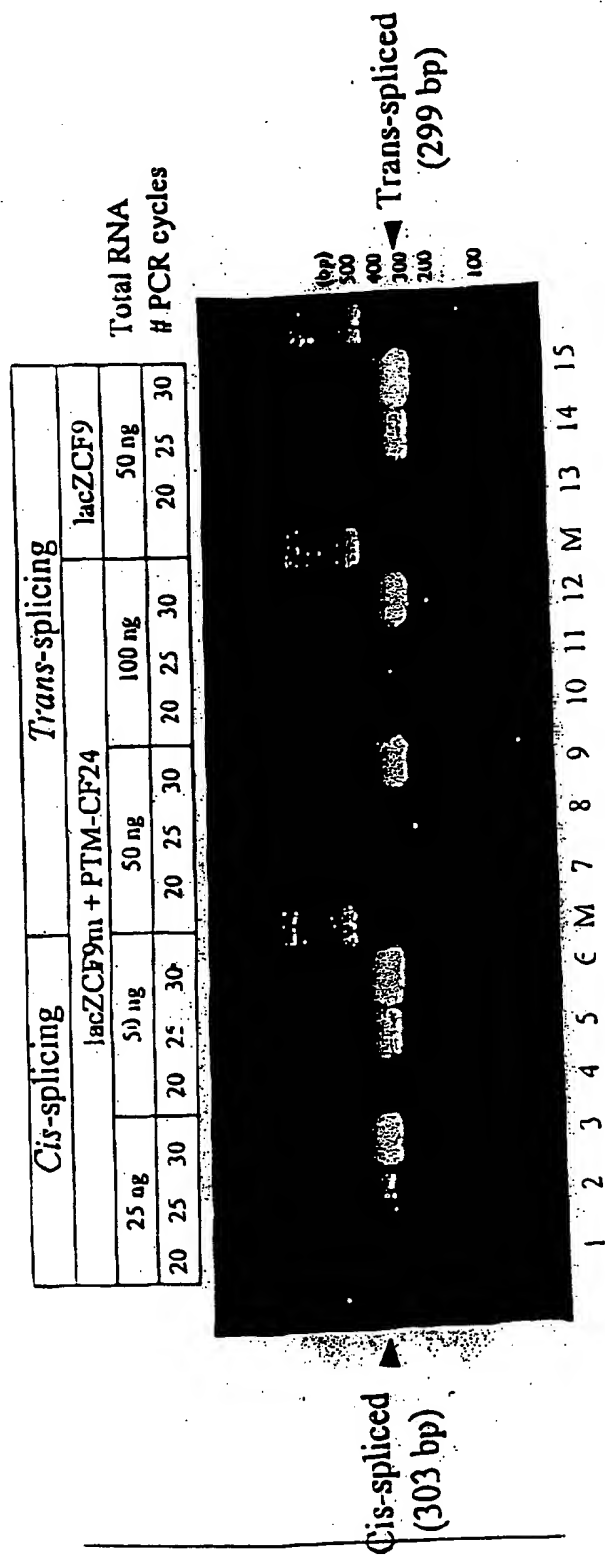


Figure 38A



[illegible]

00

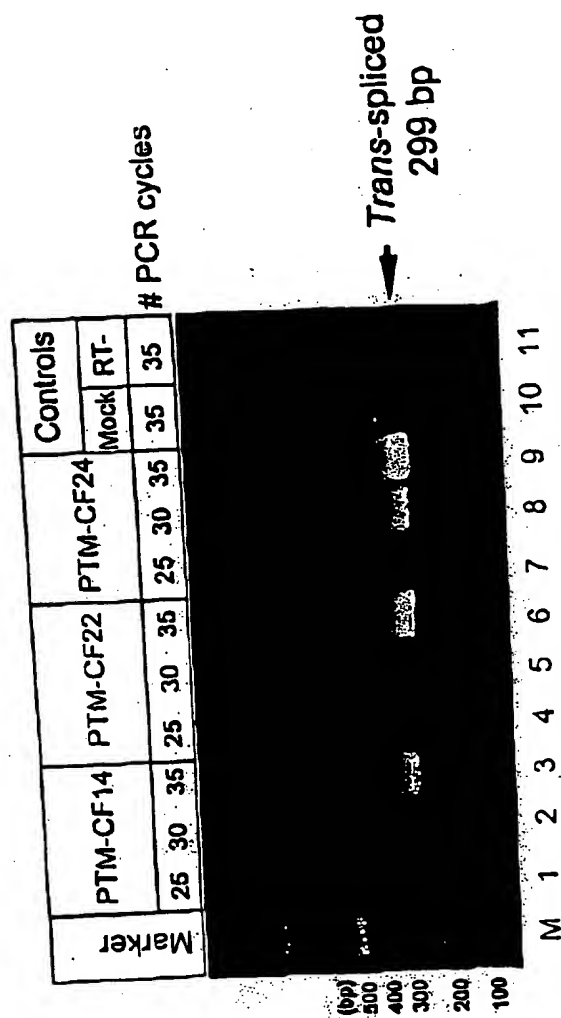


Figure 38B

51 of 68

106280" 26414650

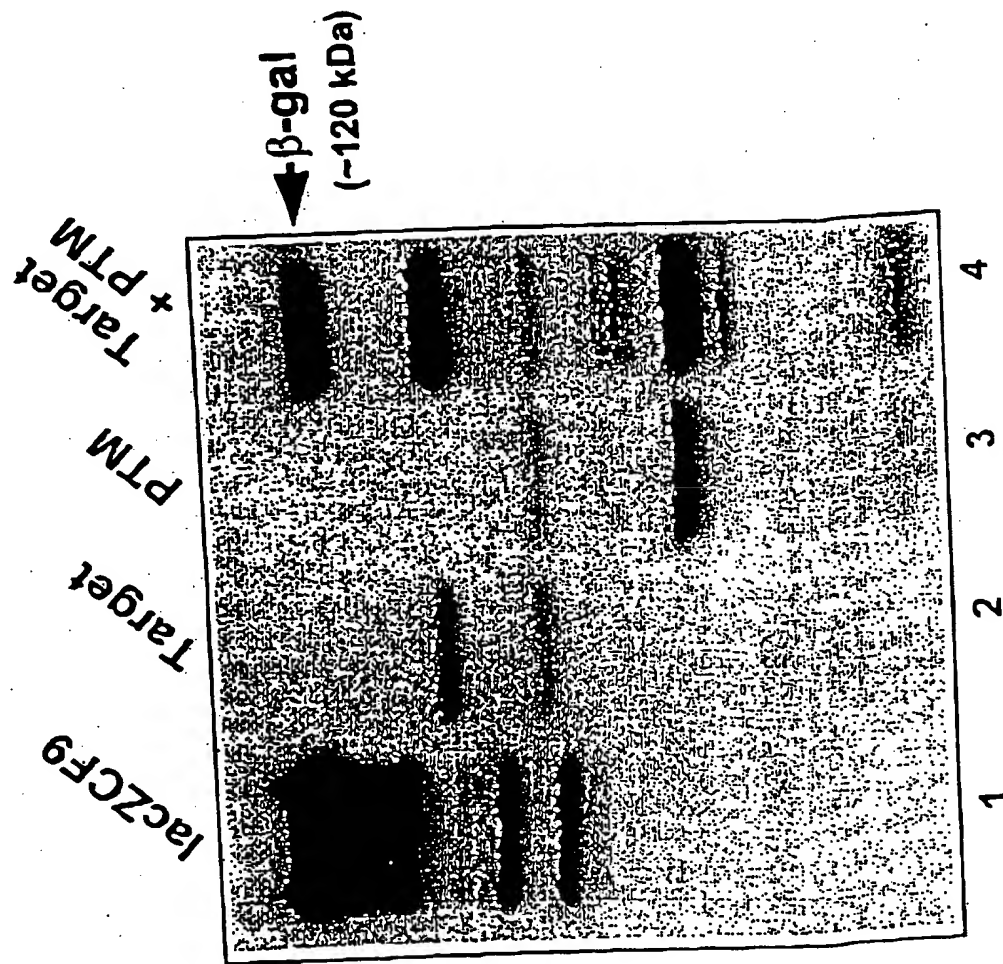


Figure 39



05941492-082901

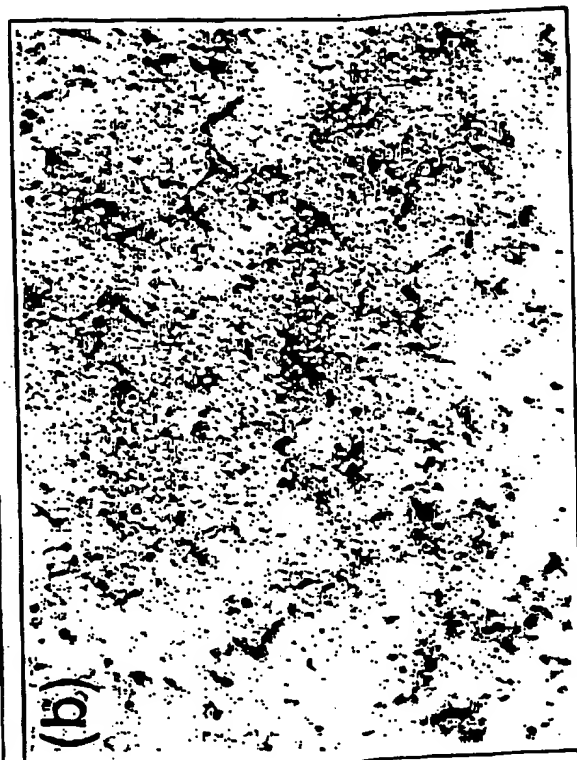
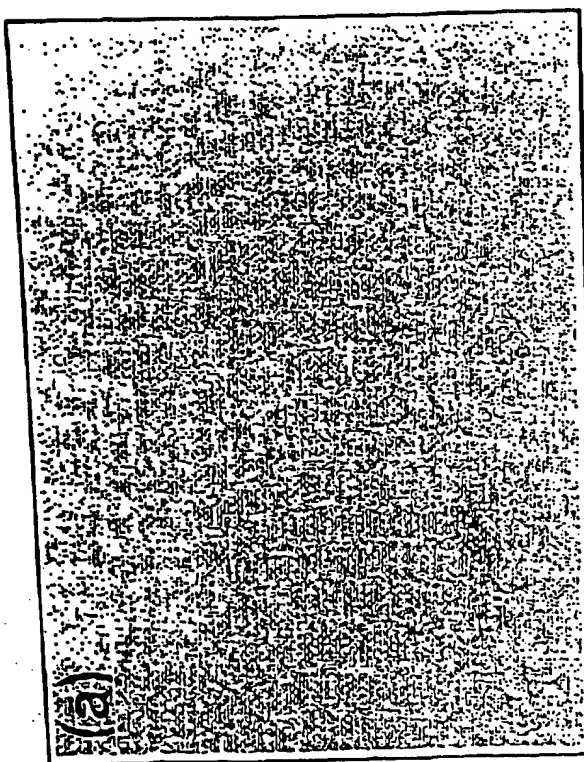


Figure 40A

53 of 89

106280" 264T450

B

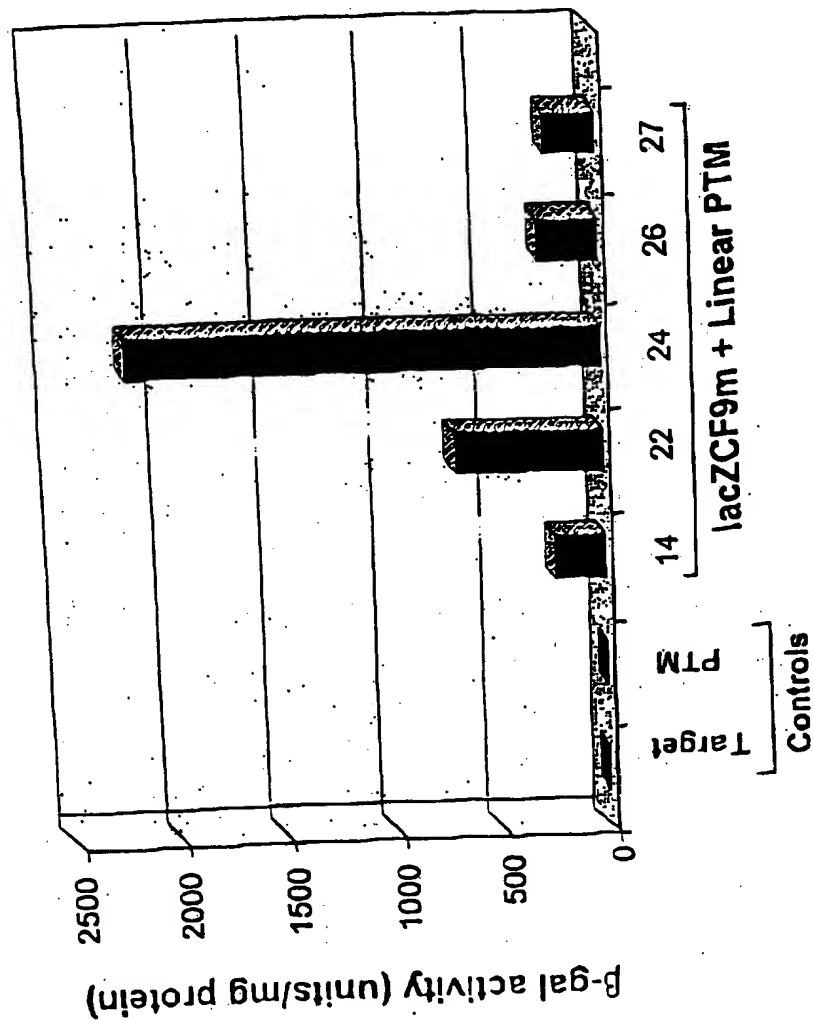


Figure 40B

54 of 89

FO6280"264T450

C

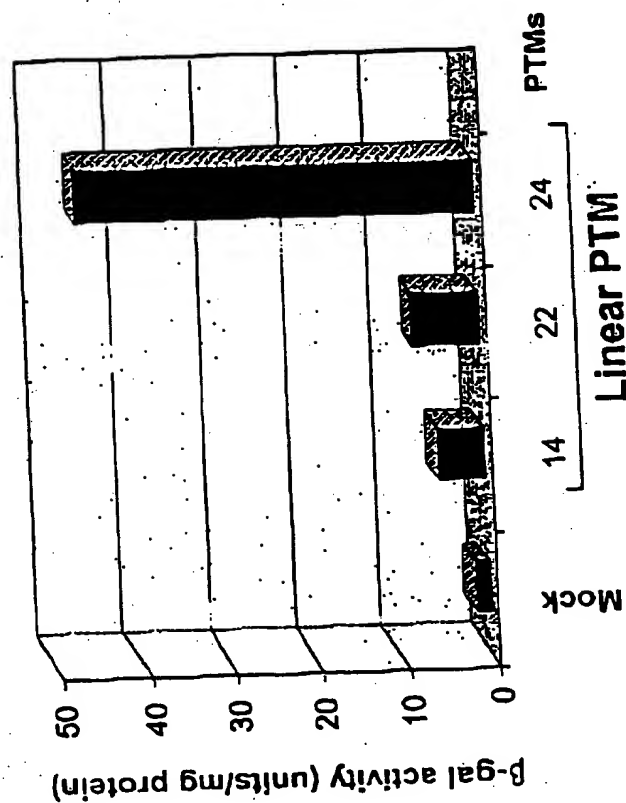


Figure 40C

55 of 89

A

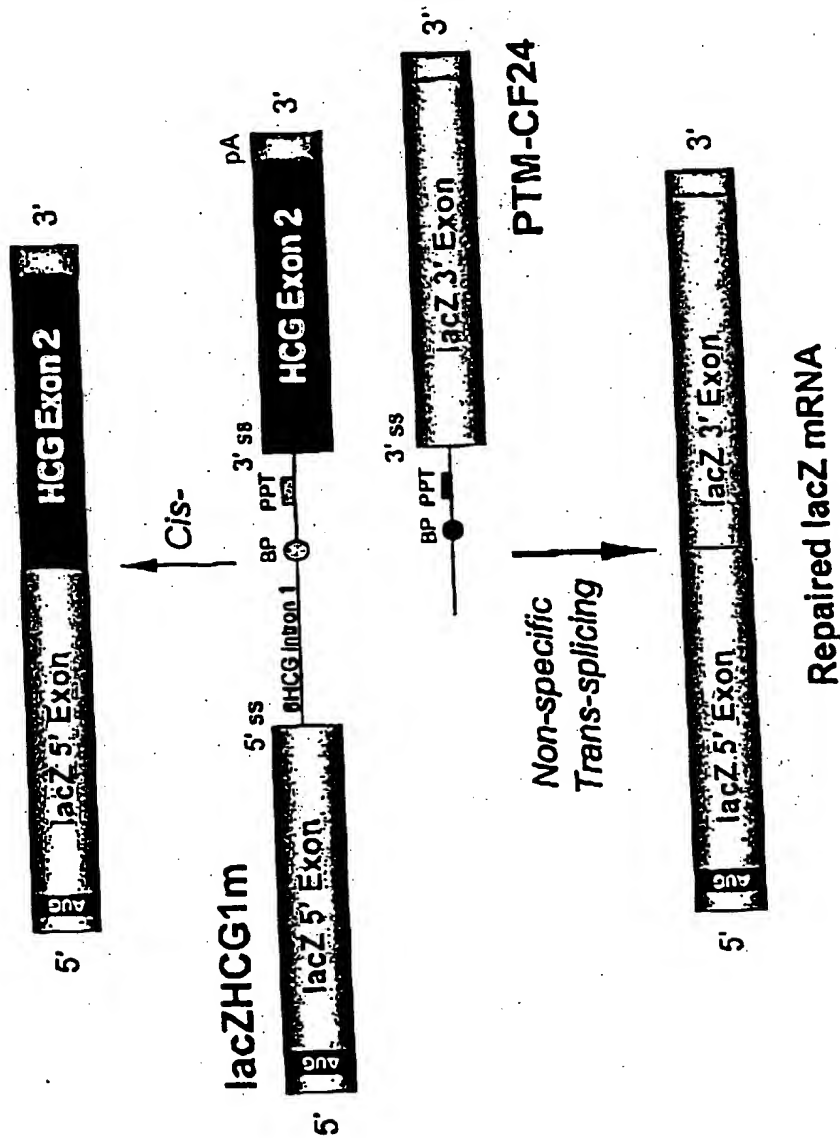


Figure 41A

56 of 89

106280-264T4660

B

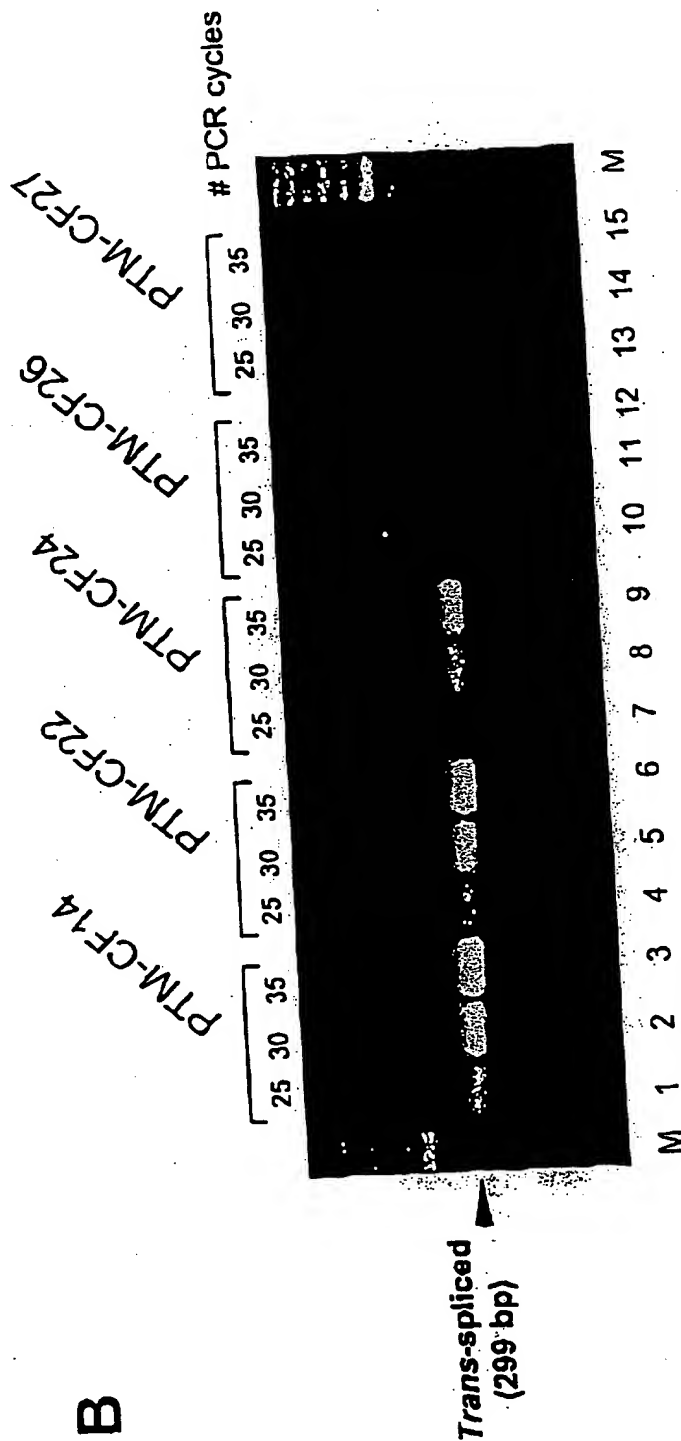


Figure 4(B)

C

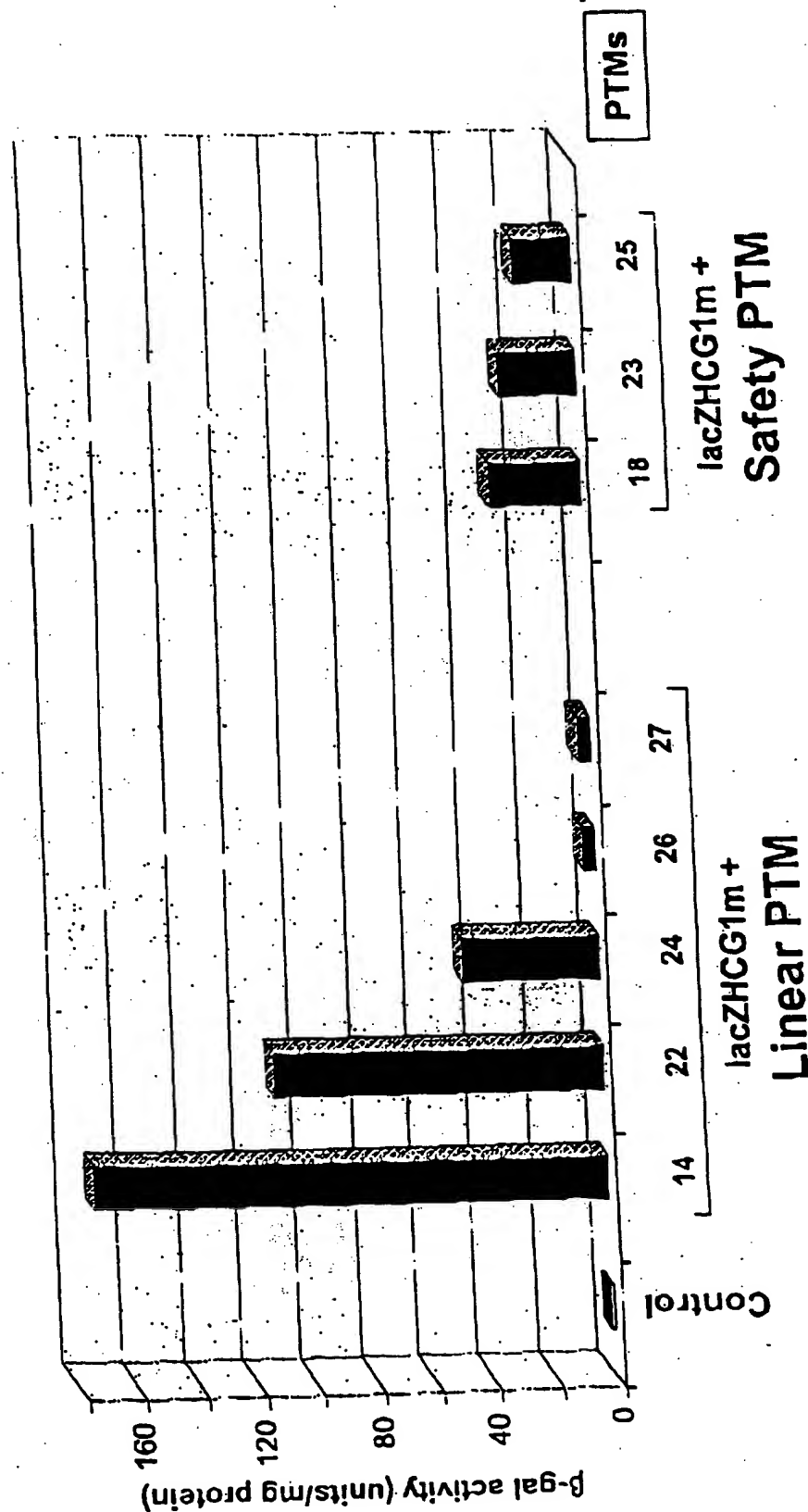


Figure 41C

58 of 89

## Exons 1-10

ATGCAGAGGTCGCCTCTGGAAAAGGCCAGCGTTGTCTCCAACTTTTTTTCAGCTGGACCAGACCAATTTTGAGGAAAG  
 GATACAGACAGCGCCTGGAATTGTCTAGACATATACCAATCCCTTCTGTTGATTCTGCTGACAATCTATCTGAAAAATT  
 GGAAAGAGAATGGGATAGAGAGCTGGCTTCAAAGAAAAATCCTAACTCATTAAATGCCCTTCGGCGATGTTTTTCTGG  
 AGATTTATGTTCTATGGAATCTTTTATATTTAGGGGAAGTCACCAAGCAGTACAGCCTCTCTTACTGGGAAGAATCA  
 TAGCTTCCTATGACCCGGATAACAAGGAGGAACGCTCTATCGCGATTTATCTAGGCATAGGCTTATGCCCTTCTCTTTAT  
 TGTGAGGACACTGCTCCTACACCCAGCCATTTTTTGGCCTTCATCACATTGGAATGCAGATGAGAATAGCTATGTTTAGT  
 TTGATTATAAGAAGACTTTAAAGCTGTCAAGCCGTGTTCTAGATAAAATAAGTATTGGACAACCTTGTTAGTCTCCTTT  
 CCAACAACCTGAACAAATTTGATGAAGGACTTGCAATTGGCACATTTTCGTGTGGATCGCTCCTTTGCAAGTGGCACTCCT  
 CATGGGGCTAATCTGGGAGTTGTTACAGGCGTCTGCCTTCTGTGGACTTGGTTTCCTGATAGTCCTTGCCCTTTTTTCAG  
 GCTGGGCTAGGGAGAATGATGATGAAGTACAGAGATCAGAGAGCTGGGAAGATCAGTGAAAGACTTGTGATTACCTCAG  
 AAATGATCGAGAACATCCAATCTGTTAAGGCATACTGCTGGGAAGAAGCAATGGAAAAAATGATTGAAAACTTAAGACA  
 AACAGAACTGAAACTGACTCGGAAGGCAGCCTATGTGAGATACTTCAATAGCTCAGCCTTCTTCTCTCAGGGTTCTTT  
 GTGGTGTTTTTATCTGTGCTTCCCTATGCACTAATCAAAGGAATCATCTCCGGAAAAATATTACCACCATCTCATTCT  
 GCATTGTTCTGCGCATGGCGGTCACTCGGCAATTTCCCTGGGCTGTACAAACATGGTATGACTCTCTTGAGCAATAAA  
 CAAAATACAGGATTTCTTACAAAAGCAAGAATATAAGACATTGGAATATAACTTAACGACTACAGAAGTAGTGATGGAG  
 AATGTAACAGCCTTCTGGGAGGAGGGATTGGGGAATTATTGAGAAAGCAAAACAAAACAATAACAATAGAAAAACTT  
 CTAATGGTGATGACAGCCTCTTCTTCACTAATTTCTCACTTCTTGGTACTCCTGTCTGAAAGATATTAATTTCAAGAT  
 AGAAAGAGGACAGTTGTTGGCGGTTGCTGGATCCACTGGAGCAGGCAAGACGAGCTTCTCATGATGATCATGGGCGAG  
TTAGAACCAAGTGAAGGCAAGATCAAACTTCCGGCCGCATCAGCTTTTGAGCCCAATTCAGTTGGATCATGCCCGGTA  
CCATCAAGGAGAACATAATCTTGGCGTCAGTTACGACGAGTACCGCTATCGCTCGGTGATTAGGCCCTGTCAAGTTGGA  
 GGAG

## Trans-splicing domain

GTAAGATATCACCGATATGTGTCTAACCTGATTGGGCTTCGATACGCTAAGATCCACCGG  
TCAAAAAGTTTTACATAATTTCTTACCTCTTCTTGAATTCATGCTTTGATGACGCTTCTGTATCTATATTCATCATTG  
GAAACACCAATGATATTTCTTAAATGGTGCCTGGCATAATCCTGGAAAACTGATAACACAATGAAATTTCTCCACTGT  
GCTTAATTTTACCCTCTGAATTTCCATTTCTCCATAATCATCATTACAACCTGAACTCTGGAAATAAAACCCATCATT  
ATTAACCTCATTATCAAATCACGCT

SCANNED, #

Figure 42

153 bp PTM24 Binding Domain:

Nhe I

153 bp BD underlined

GCTAGC-GACGAAGCGCCCTCACGCTCAGGATTCACTTGCCTCCAATTATCATCCTAAGCAGAAGTGTATA

TTCTTATTTGTAAAGATTCTATTAACTCATTTGATTCAAATAATTTAAATACTTCCCTGTTTCACCTACTCTGCTATGC

Sac II

AC-CCGCGG

68 7 09

Figure 43A



Trans-splicing domain  
AATAATGACGAAGCCGCCCTCACGCTCAGGATTCACCTTGCCCTCAATTATCATCCTAAGCAGAAGTGTATATTCTTA  
TTTGTAAGATTCTATTAACTCATTTGATTCAAAATATTTAAATACTTCTGTTCACCTACTCTGCTATGCACCCGC  
GGAACATTATTATAACGTTGCTCGAATACTAAGTGGTACCTCTTCTTTTTTTTGATATCTGCAG

Exons 10-14

ACTTCACTTCTAATGATGATTATGGGAGAACTGGAGCCTTCAGAGGGTAAAATTAAAGCACAGTGGGAAGAATTTCACTCT  
GTTCTCAGTTTTTCTGGATTATGCTTGGCACCATTAAAGAAAATATCATCTTTGGTGTTTCTATGATGAATATAGATA  
CAGAAGCGTCATCAAAGCATGCCAACTAGAAGAGGACATCTCCAAGTTTGACAGAGAAAGACAATATAGTTCCTGGAGAA  
GGTGAATCACACTGAGTGGAGGTCAACGAGCAAGAATTTCTTTAGCAAGAGCAGTATACAAAGATGCTGATTGTATT  
TATTAGACTCTCCTTTTGGATACCTAGATGTTTTAAGAGAAAAGAAATATTTGAAAGCTGTGTCTGTAACTGATGGC  
TAACAAAACCTAGGATTTTGGTCACTTCTAAAATGGAACATTTAAAGAAAGCTGCACAAAATATTAATTTTGCATGAAGGT  
AGCAGCTATTTTATGGGACATTTTCAAGACTCCAAATCTACAGCAGACTTTAGCTCAAACCTCATGGGATGTGATT  
CTTTTCGACCAATTTAGTGCAGAAAGAAATCTCAATCCTAAGTGACACTTACACCGTTTCTCATTAGAAGGAGATGC  
TCCTGTCTCCTGGACAGAAACAAAAAAACAATCTTTTAAACAGACTGGAGAGTTTGGGGAAAAAAGGAAGAATTCATT  
CTCAATCCAATCAACTCTATACGAAAATTTTCCATTGTGCAAAAGACTCCCTTACAAATGAATGGCATCGAAGAGGATT  
CTGATGAGCCTTTAGAGAGAAGGCTGTCTTAGTACCAGATTCTGAGCAGGGAGAGGCGATACTGCCTCGCATCAGCGT  
GATCAGCACTGGCCCCACGCTTCAGGCACGAAGGAGGCAGTCTGTCTGAACCTGATGACACTCAGTTAACCAGAGT  
CAGAACATTCACCGAAAGACACAGCATCACGAAAGTGTCACCTGCGCCCTCAGGCAACTTGACTGAACTGGATA  
TATTTCAAGAAGGTTATCTCAAGAACTGGCTTGGAAATAAGTGAAGAAATTAACGAAGAAGACTTAAAGGAGTGCTT  
TTTTTGATGATATGGAGAGCATACCAGCAGTGACTACATGGAACACATACCTTCGATATATTACTGTCCACAAGAGCTTA  
ATTTTTGTGCTAATTTGGTGCTTAGTAATTTTTCTGCGAGAGGTGGCTGCTTCTTTGGTTGTGCTGTGGCTCCTTGGAA  
ACACTCCTCTTCAAGACAAAGGGAATAGTACTCATAGTAGAAATAACAGCTATCGAGTGATTATCACCAGCACCAGTTC  
GTATTATGTGTTTTACATTTACGTGGGAGTAGCCGCACATTTGCTTGCTATGGGATTTCTCAGAGTCTACCAGTGGT  
CATACTCTAATCAGAGTTCGAAAATTTTACACCAAAAATTTTACATTTCTGTTCTTCAAGCACCTATGTCAACCCTCA  
ACAGCTTGAAGCAGGTGGGATTCTTAATAGATTCTCCAAAGATATAGCAATTTTGGATGACCTTCTGCCTCTTACCAT  
ATTTGACTTCATCCAGTTGTTATTAATTGTGATTGGAGCTATAGCAGTTGTGCGAGTTTTACAACCCTACATCTTTGTT  
GCAACAGTGCCAGTGATAGTGGCTTTTATTATGTTGAGAGCATATTTCTCCAAACCTCACAGCACTCAAACAATGG  
AATCTGAAGGCAGGAGTCCAATTTTCACTCATCTTGTTACAAGCTTAAAGGACTATGGACACTTCGTGCTTCGGAGC  
GCAGCCTTACTTTGAAACTCTGTTCCACAAGCTCTGAATTTACATCTGCTGCCAATGGTTCTTGTTACCTGTCAACACTG  
CGCTGGTTCCAAATAGAACTAGAAATGATTTTTGTCACTTTCTTTCATTGCTGTTACCTTCATTTCCATTTTAAACAAG  
GAGAAGGAGAAGGAAGAGTTGGTATTATCCTGACTTTAGCCATGAATATCATGAGTACATTGCAGTGGGCTGTAAACTC  
CAGCATAGATGTGGATAGCTTGATGCGATCTGTGAGCCGAGTCTTTAAGTTCAATTGACATGCCAACAGAAGGTAAACCT  
ACCAAGTCAACCAACCATACAAGAAATGGCCAACTCTCGAAAGTTATGATTATTGAGAATTCAACGTGAAGGAAGATG  
ACATCTGGCCCTCAGGGGGCCAAATGACTGTCAAAGTCTCACAGCAAAATACACGAAGGTGGAAATGCCATATTAGA  
GAACATTTCTTCTCAATAAGTCTGTAGCTGGAGGTGGCGCTCTTGGGAAGAAGTGGATCAGGGAAGAGTACTTTGTTA  
TCAGCTTTTTTGGAGACTACTGAACACTGAAGGAGAAATCCAGATCGATGGTGTGTCTTGGGATTCAATAACTTTGCAAC  
AGTGGAGGAAGCCTTTGGAGTGATACCAAGAAAGTATTTATTTTTCTGGAACATTTAGAAAAAAGCTGGATCCCTTA  
TGAAACAGTGGAGTGATCAAGAAATATGGAAAGTTGCAGATGAGGTTGGGCTCAGATCTGTGATAGAACAGTTTCTGGG  
AAGCTTGACTTTGTCTTGTGGATGGGGCTGTGTCTTAAGCCATGGCCACAAGCAGTGTGATGTGCTGGCTAGATCTG  
TTCTCAGTAAGGCGAAGATCTGTGCTGTGATGAACCCAGTGCTCATTGGATCCAGTAACTACCAAAATAATTAGAAG  
AACTCTAAAACAAGCAATTTGCTGATTGCAAGTAATCTCTGTGAACAAGGATAGAAGCAATGCTGGAATGCCAACAA  
TTTTTGGTTCATAGAAGAGAAACAAGTGCGGCAGTACGATTCCATCCAGAACTGCTGAACGAGAGGAGCCTCTTCCGGC  
AAGCCATCAGCCCTCCGACAGGTTGAAGCTCTTCCCCACCGGAATCAAGCAAGTGAAGTCTAAGCCCCAGATTGC

Histidine tag      Stop

TGCTCTGAAAGAGGAGACAGAAGAAGAGGTGCAAGATACAAGGCTTCATCATCATCATCATTAG

Figure 43B



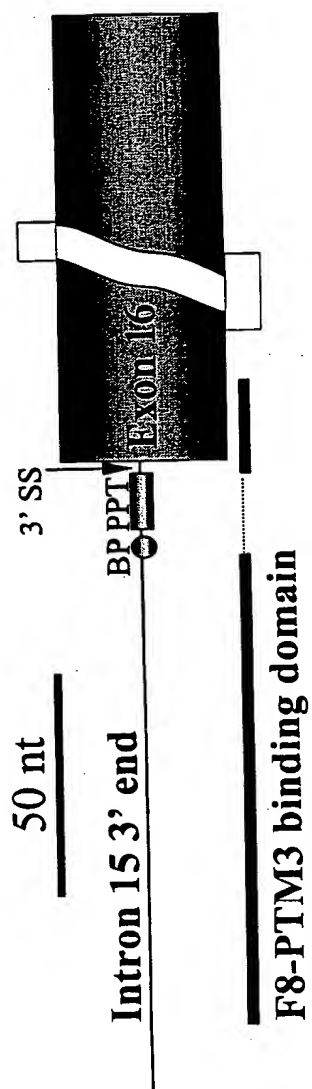


Figure 44B

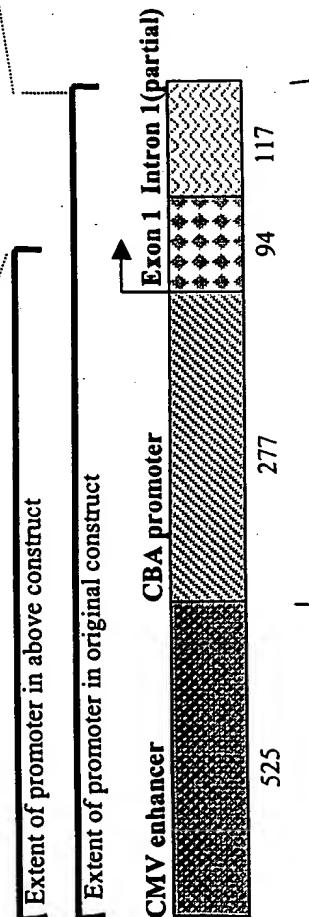
Figure 44C

**NN**nucleotide changes are shown in blue  
**Boxed** = CAT box, TATA box  
**Boxed + Arrow** = Transcription Start  
**Oval** = Downstream elements  
**Bold** = Binding domain  
**Italicized** = Spacer+PPT+BP+AG dinucleotide

Ncol  
 CCATGGTGCACGTTAGCCCCACGTTCTGCTTCACTCTCCCCCATCTCCCCCCCCTCCCCACCCCCCAATT  
 TTTGTATTATTATTATTTTTTAATTATTTTGTGCAGCGATGGGGGGGGGGGGGGGGGGGGGGGGCGCCAGGC  
 GGGGGCGGGCGGGCGAGGGGGCGGGGGCGAGGGCGGAGAGGTGCGGCGGAGCCTAATCAAGCGGCGCG  
 CTCCGAAAGTTTCCTTTTATGGCGAGCGCGGGCGGGCGGGCGGGCGGGCGGGCGGGCGGGCGGGCGG  
 EGAGTCGTGCGACGCTGCCTTCGCCCCGTCGCGAACCCTCGAGCTTACCTGAACTAATTTTTTAGAA  
 E13 ..... XhoI  
 HindIII  
 TATTAAATCCTAAGCTTTTATATCTCTATCCCTCTATCTTTTGTCTCTCTATCCAAATTTTATTAACTTAGA  
 Sall  
 CTTTAAAGAAACTTATGAGAAAAATTTCCGGGGAACATTATTATAACGTTGCTCGAAATACTAAGTGTAC  
 EcoRV  
 PstI  
 CTCTCTCTTTTGTGATATCTCTGCAG

Sequence not included in construct

CGCGCGCCCTCGCGCGCCCGCGCGCTCTGACTGACCGCGTTACTCCACACAGTCAG  
CGGGCGGGACGGCCCCCTCTCTCCGGGCTGTAAATTAGCGCTTGGTTTAATGACGGCT  
TGGTGGTCTGGTCTGGCTGCAAGCCCTTGAGGGGCTCCGGGAGGAATTCGTA

$$\begin{aligned} \mathbf{F13} + \mathbf{F2} &= 235 + 106 = 341 \text{ bp} \\ \mathbf{F13} + \mathbf{F4} &= 235 + 315 = 550 \text{ bp} \end{aligned}$$


Chicken Beta Actin Promoter (including exon 1 and part of intron 1)

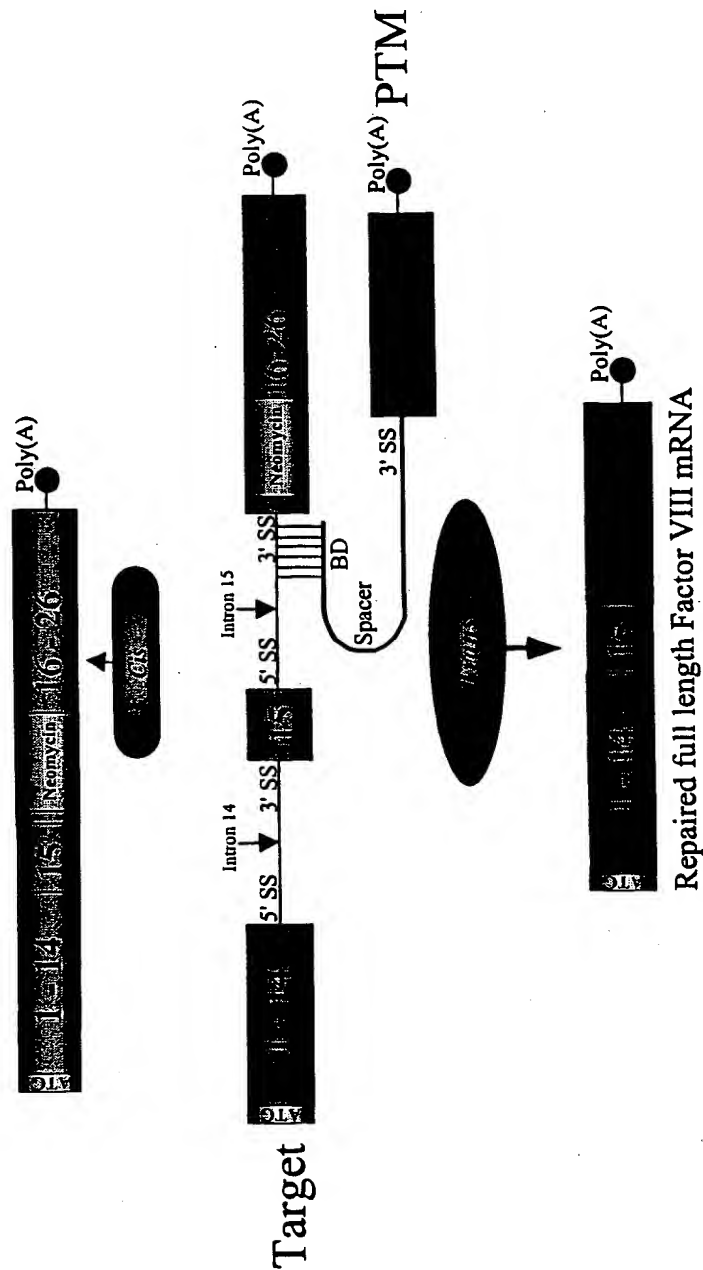
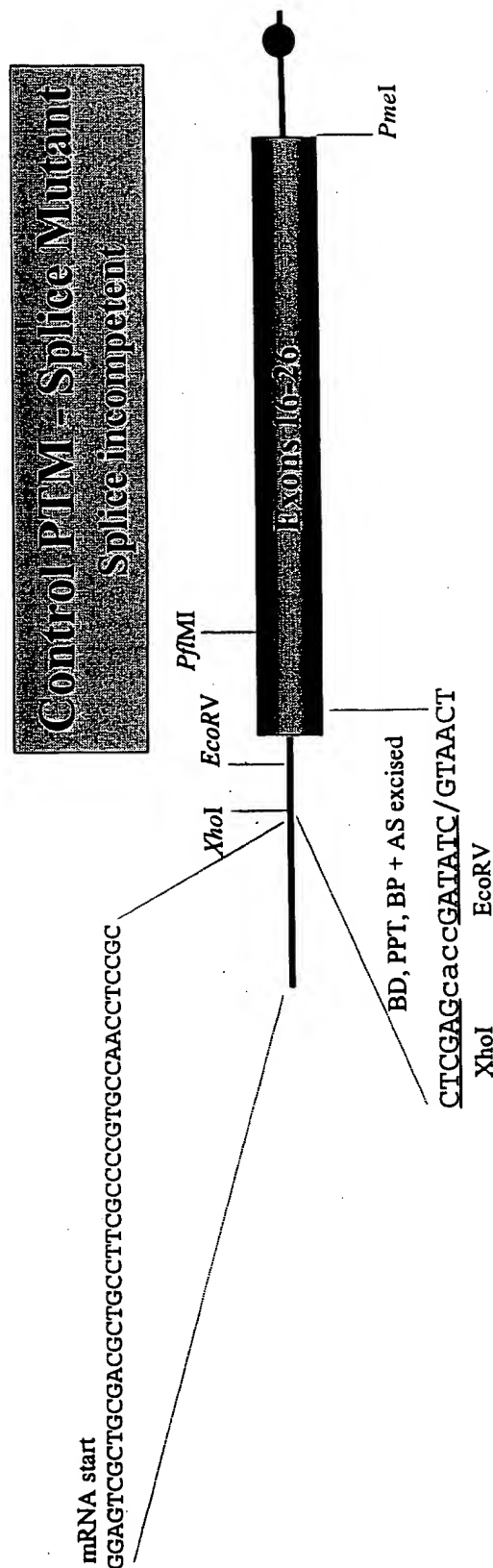


Figure 44D

Figure 45



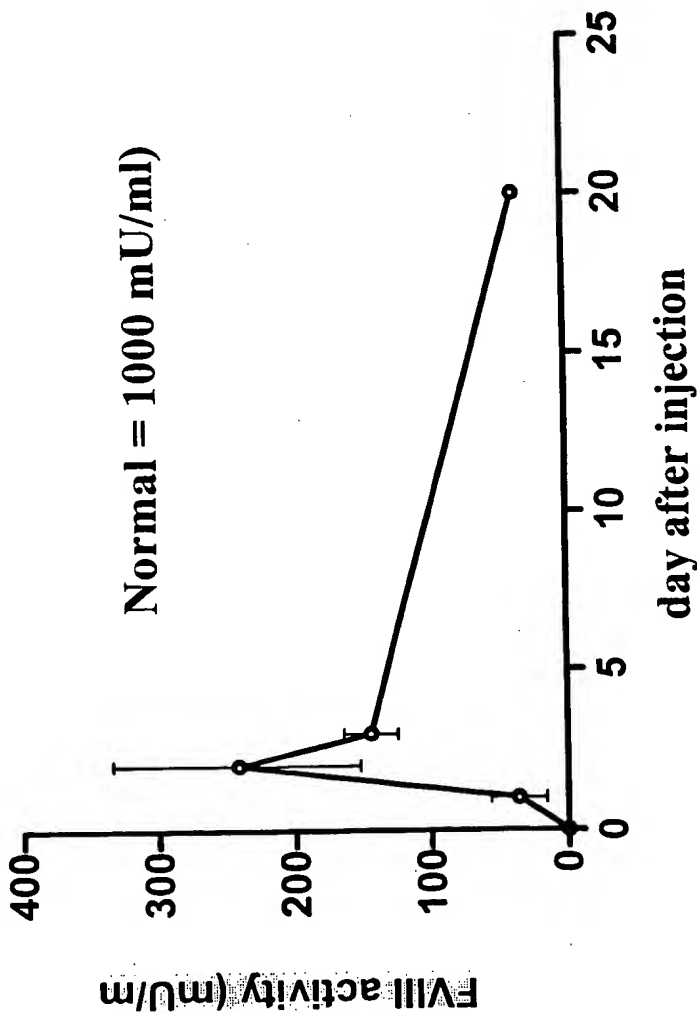
Method:

- Excise TSD and part of exon 16 with XhoI and PflMI and ligate in a PCR product that:
- 1) eliminates the TSD and splice acceptor site
  - 2) inserts EcoRV adjacent to exon 16
  - 3) restores the coding for exon 16

# Repair of Factor VIII

*Preliminary results from one experiment*

FVIII activity in Exon 16 FVIII-KO mice  
after IV PTM-FVIII intraportal infusion  
(100ugDNA)(n=3)



## METHODS

Inject plasmid intraportally



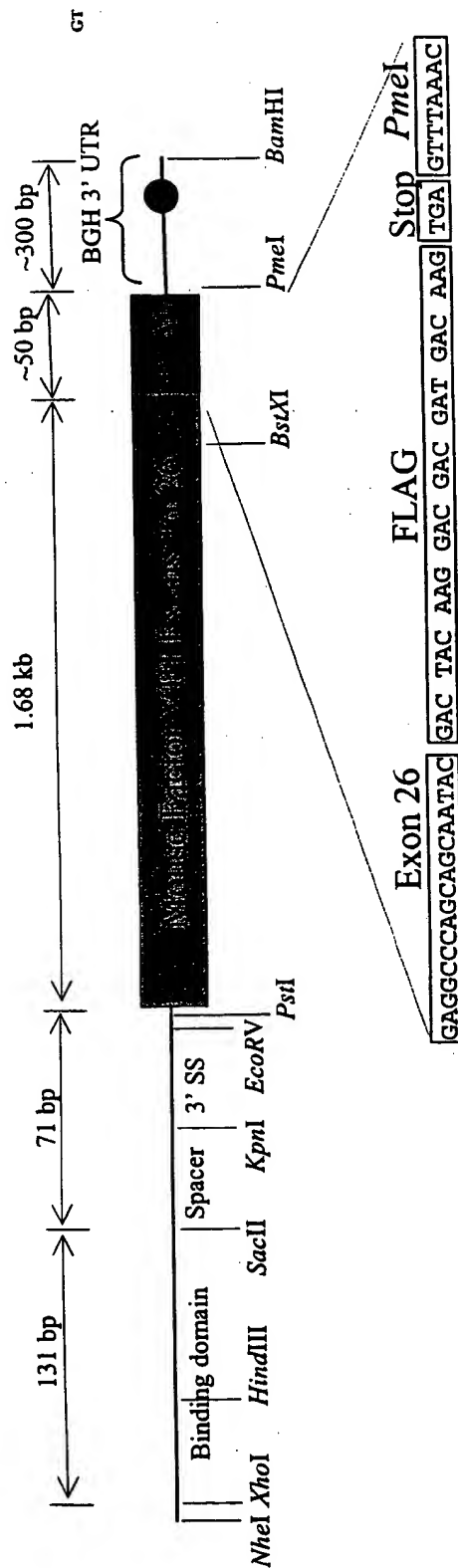
Sample blood (1, 2, 3, 20 d)



Assay for factor VIII activity

Figure 46

Detailed structure of a mouse factor VIII PTM containing normal sequences for exons 16-26 and a C-terminal FLAG tag. BGH = bovine growth hormone 3' UTR; Binding domain = 125 bp.

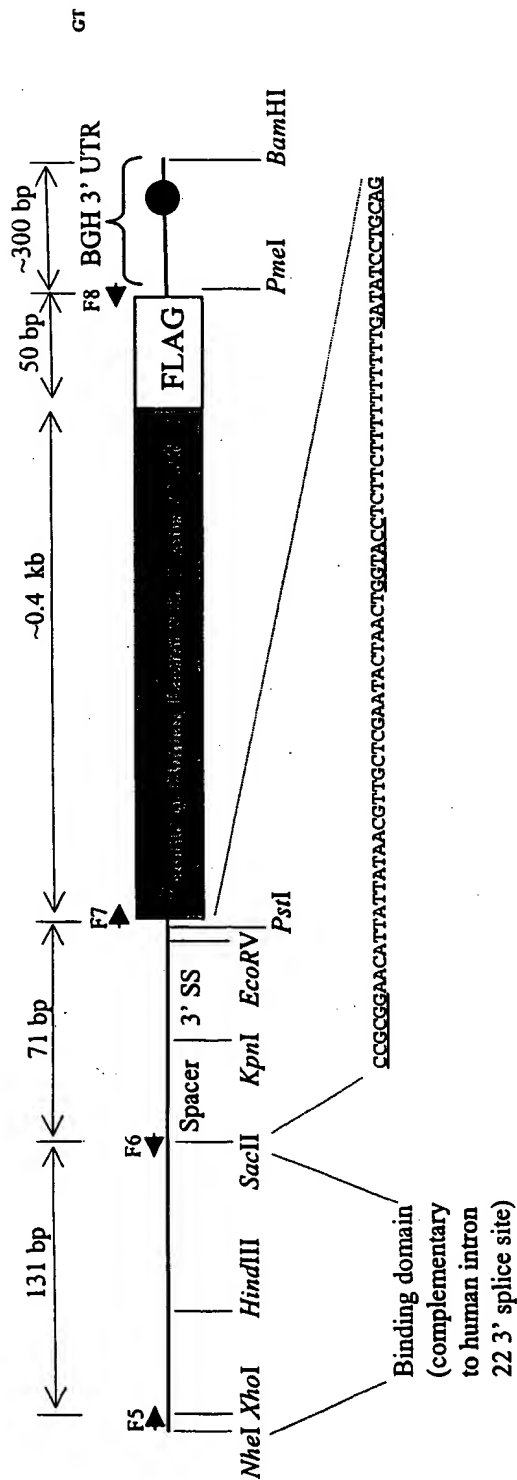


**REFERENCE FOR DESIGN OF FLAG TAG**

Brann T, Kayda D, Lyons RM, Shirley P, Roy S, Kaleko M, Smith T.  
Adenoviral vector-mediated expression of physiologic levels of human factor VIII in nonhuman primates.  
Hum Gene Ther 1999 Dec 10;10(18):2999-3011  
Genetic Therapy, Inc., a Novartis Company, Gaithersburg, MD 20878, USA.  
Epitope-tagged B domain-deleted human factor VIII cDNA (flagged FVIII) was evaluated in nonhuman primates.

Figure 47A





FLAG = C-terminal tag to be used to detect repaired factor VIII protein.

Figure 47B

# Transcription Map of HPV-16

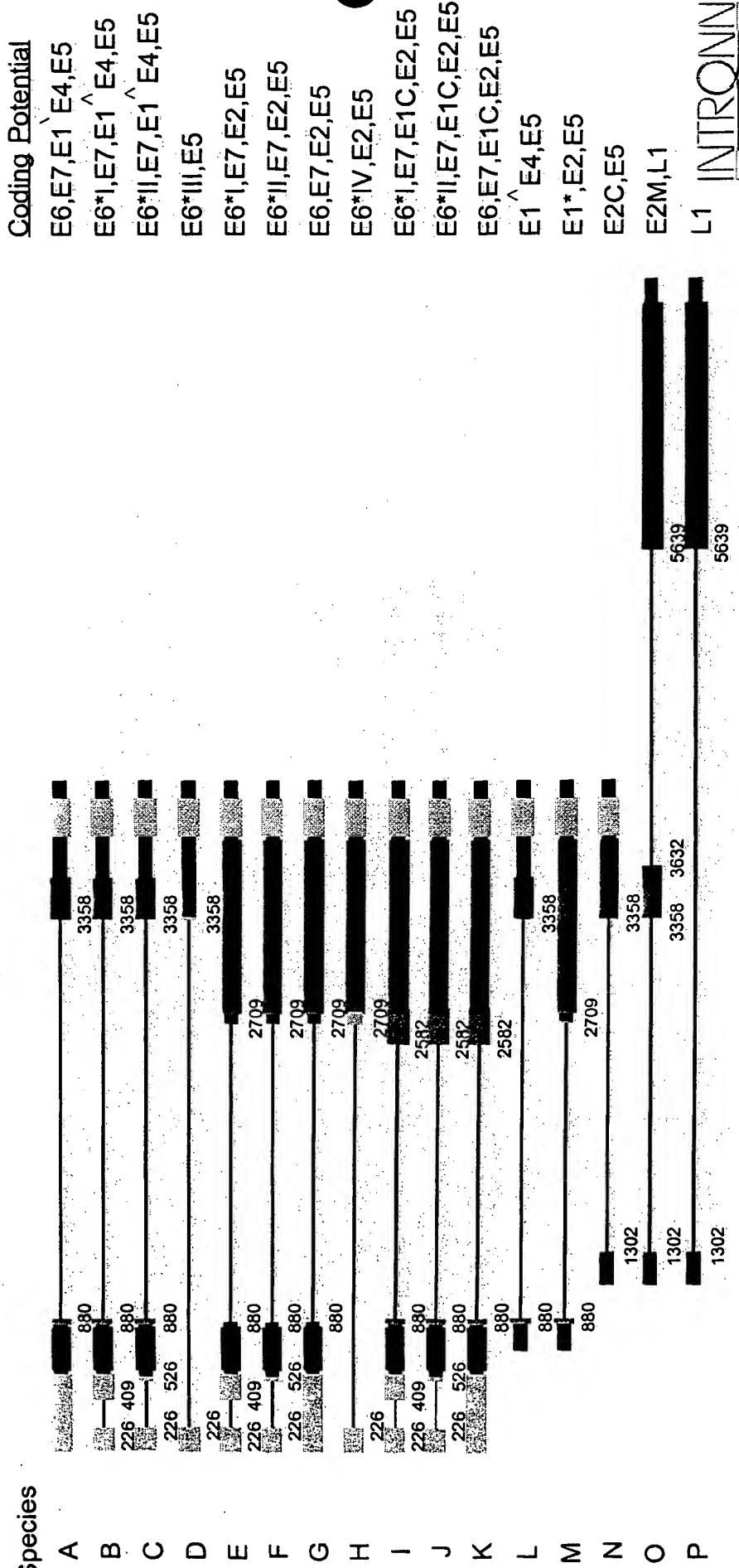
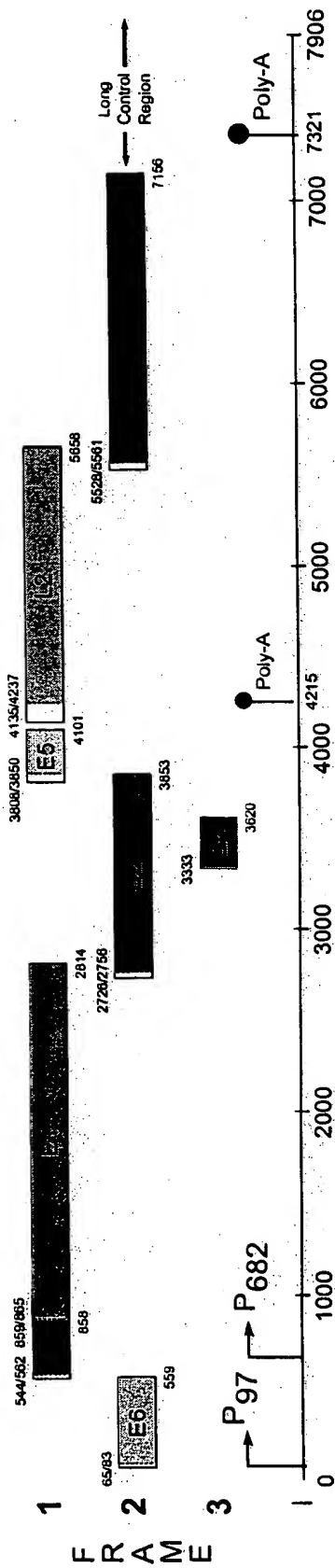


FIGURE 48



72 of 89



五十五



# PTM Design

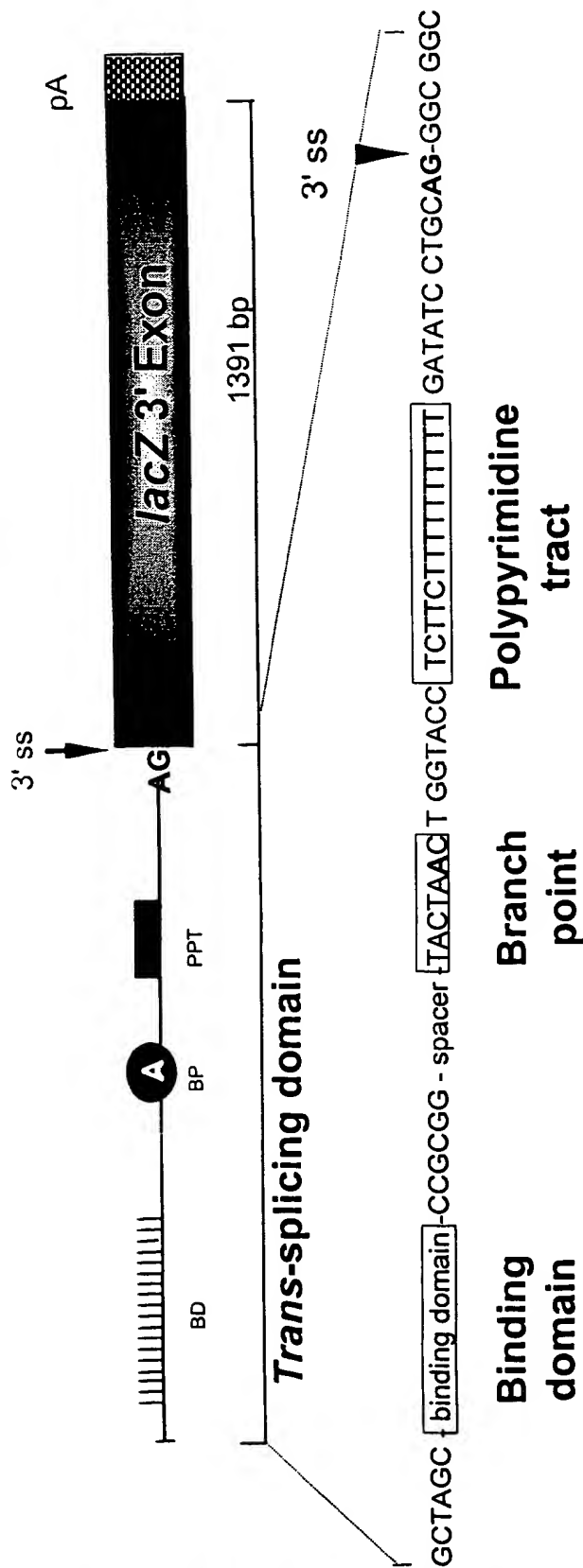
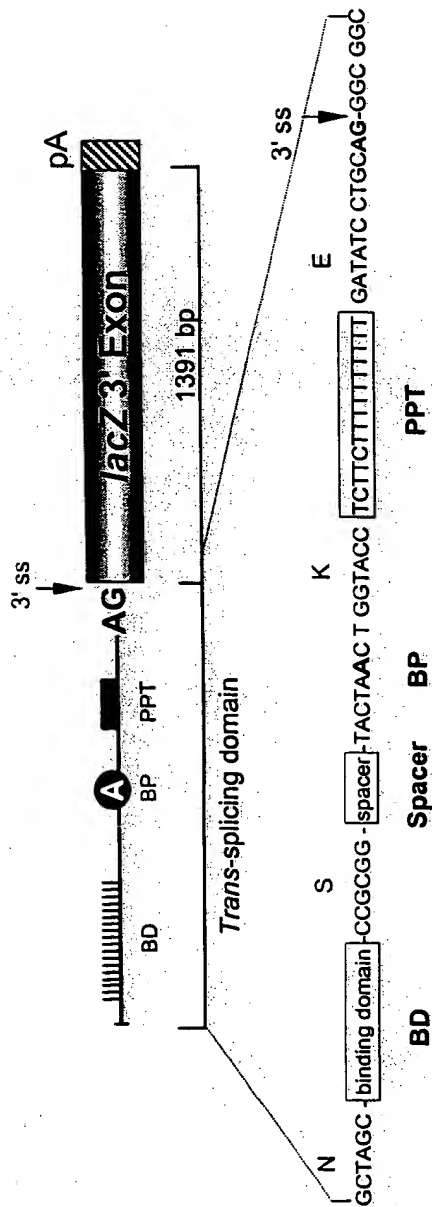


FIGURE 5/

13 Feb 89

HPV-PTM1 with 80 bp binding domain targeted to 3' ss at 409 :



**Binding domain sequence:** CAGTTAATAC ACCTAATTAA CAAATCACAC AACGCTTTGT TGTATTGCTG  
TTCTAATGTT GTTCCATACA CACTATAACA

HPV-PTM2 with 149 bp binding domain targeted to 3' ss at 409:

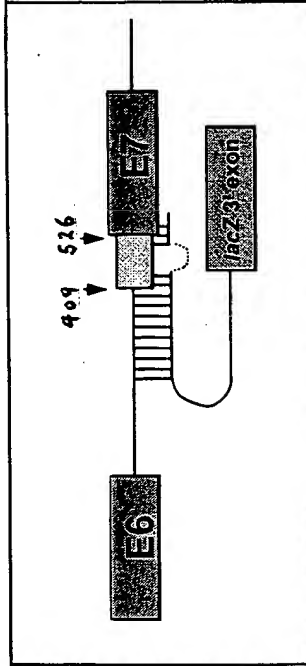


**Binding domain sequence:** CAGTTAATAC ACCTAATTAA CAAATCACAC AACGCTTTGT TGTATTGCTG  
TTCTAATGTT GTTCCATACA CACTATAACA ATAATGTCTA TACTCACTAA  
TTTTAGAATA AAACCTTTAA CATTATCAC ATACAGCATA TCGATTCCC

## Binding Domains of HPV-PTM3 and 4

**HPV-PTM3 Binding domain** (covers both 3' ss at 409 and 526; has 53 bp bubble)

GATGATCTGCAACAAGACATACATCGACCGGTCCA (53 nt bubble) CTTACGGACACAGTGGCTTTTGAC  
AGTTAATACACCTAATTAAACAAATCACACAAACGGTTTGTGTATTGCAGTTCTTAATGTTGCCATACACACTA  
TAACAAAT



**HPV-PTM4 Binding domain** (covers both 3' ss at 409 and 526; has 76 bp bubble)

GATGATCTGCAACAAGAC (76 nt bubble) GACACAGTGGCTTTTGACAGTTAATACACCTAATTAAACAAATC  
ACACAAACGGTTTGTGTATTGCAGTTCTTAATGTTGCCATACACACTATAACAAT

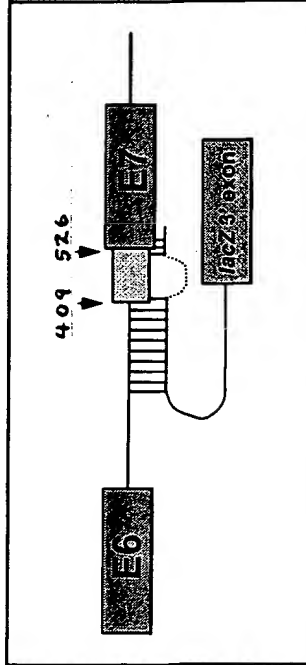


FIGURE 5.3



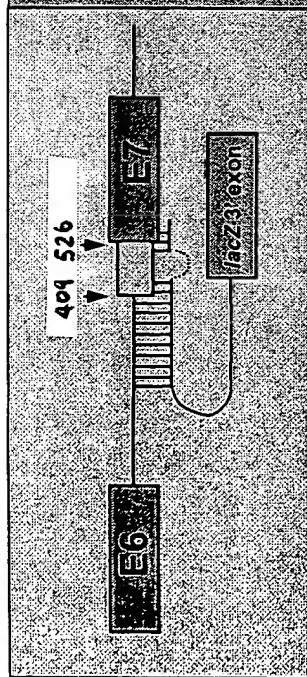
# HPV-PTM5 and 6

**HPV-PTM5**, Binding domain (140 nt, has 53 nt bubble, covers 3'ss at position 409 and 526)

GATGATCTGCAACAAGACATACATCGAC**CCGT**TCCA. CTTGAGGACACAGTGGCTTTTGACAGTTAATACACCTAATTAAACAATCACACAACGGT  
TTGTTGTATTGCAGTTCTAATGTTGTTCCATACACACTATAACA

CCGT

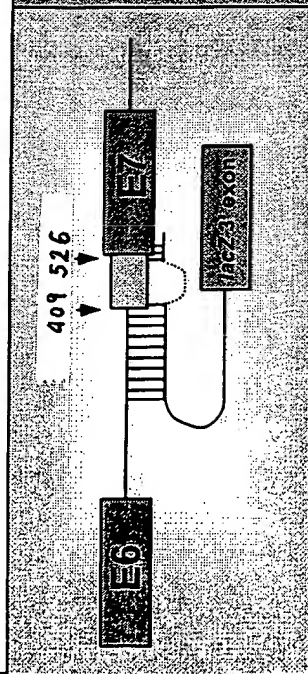
CCGT



**HPV-PTM6**, Binding domain (117 nt, has 76 nt bubble, covers 3'ss at position 409 and 526)

GATGATCTGCAACAAGAC. GACACAGTGGCTTTTGACAGTTAATACACCTAATTAAACAATCACACAACGGT**CCGT**TGTTGTATTGCAGTTCT  
AATGTTGTTCCATACACACTATAACA

CCGT



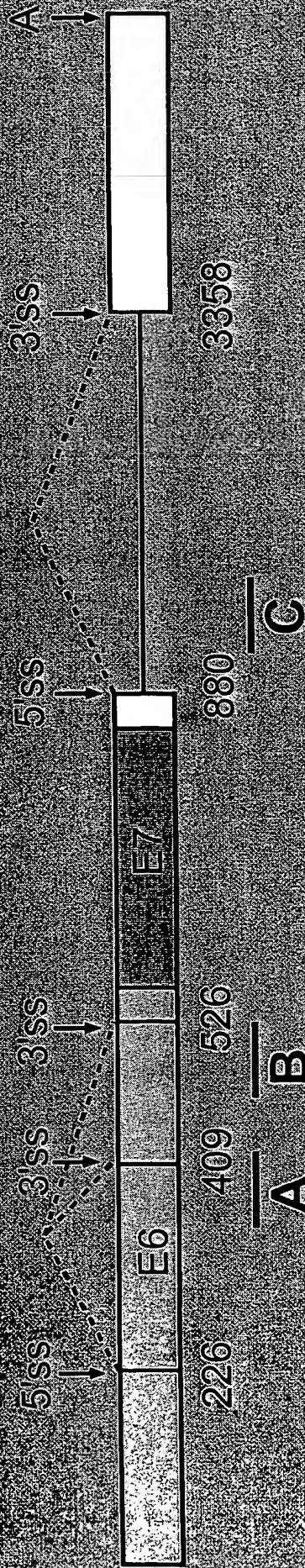
Note: Nucleotides in bold are modified to prevent PTMs cryptic splicing

FIGURE 54

INTRONN



# Positions of HPV-PTM Targeting Domains

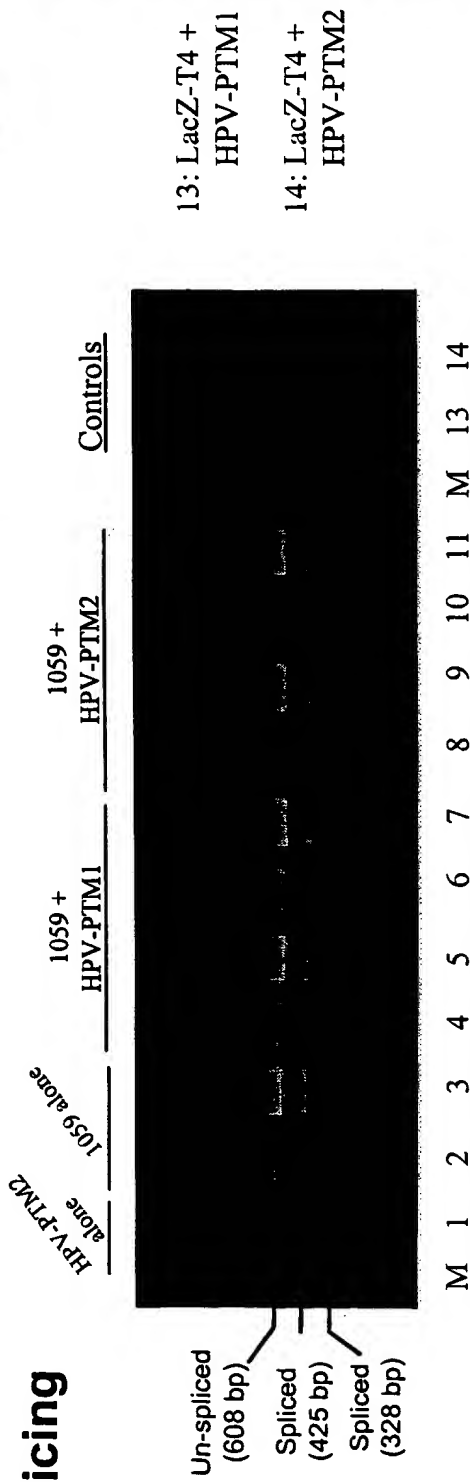


**Binding Domain**

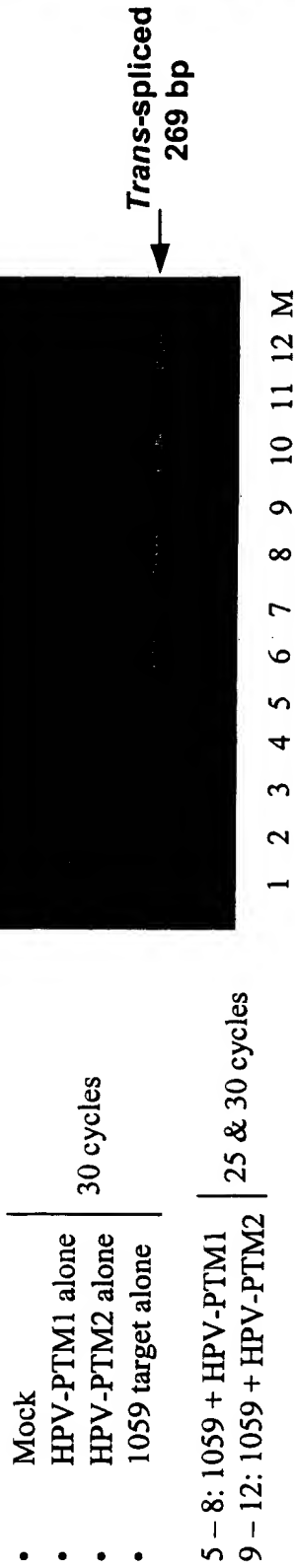
<u>PTM</u>	<u>Region</u>	<u>Size (nt)</u>
HPV-PTM1	A	80
HPV-PTM2	A	149
HPV-PTM5	A+B	140
HPV-PTM6	A+B	117
HPV-PTM8	C	104
HPV-PTM9	C	174

# Trans-splicing Efficiency of HPV-PTMs in 293T Cells

## Cis-splicing



## Trans-splicing



RT-PCR Analysis of total RNA



# Trans-splicing between target pre-mRNA and PTM is accurate (293T cells)

0.1 μM D16

E6

lacZ 3' exon

Lac6R

Sequencing  
Primer

Trans-spliced  
Chimeric mRNA

E6 of HPV-16R

TS junction

lacZ 3' exon

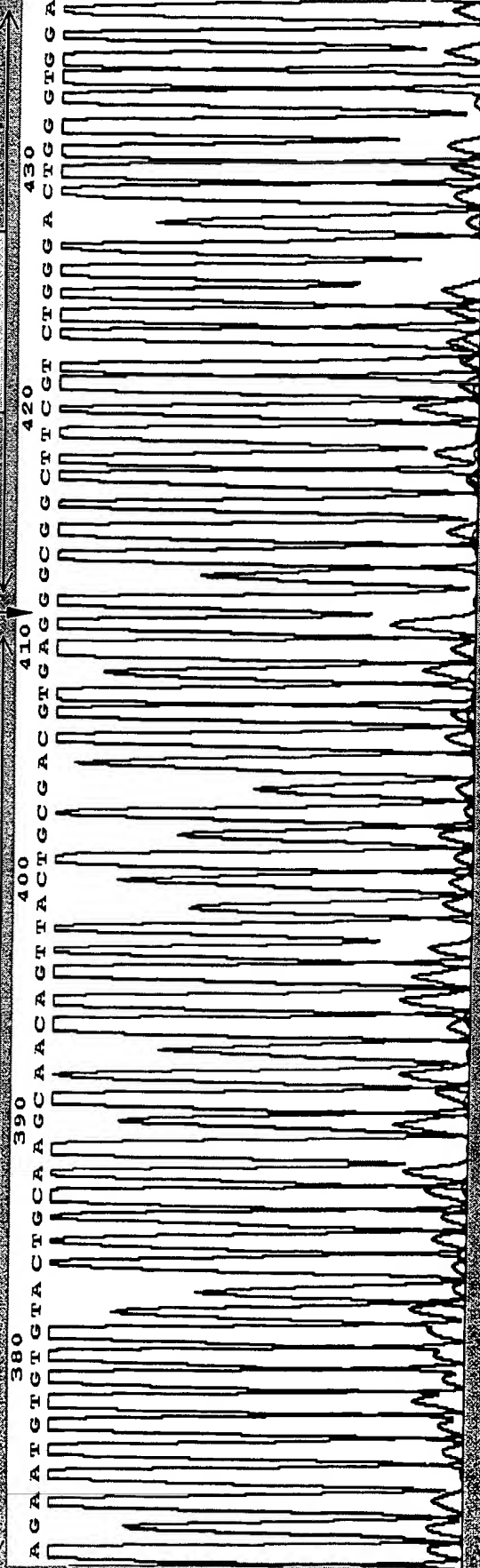
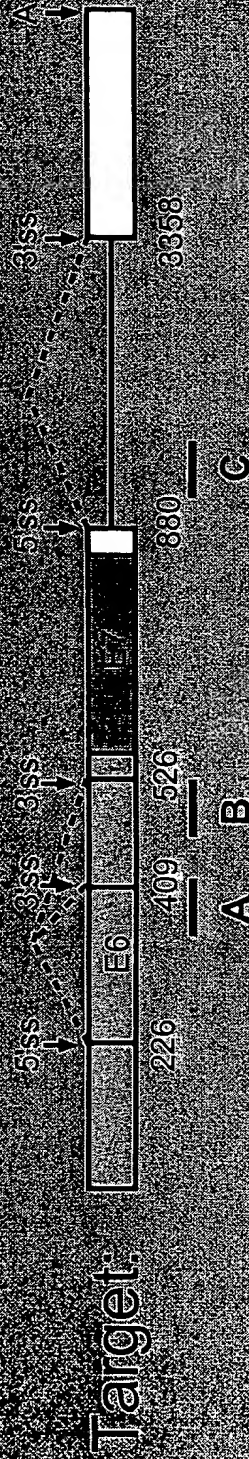


FIGURE 57

INTRONN

79 of 84

# Trans-splicing in 293 Cells (Co-transfections)



PTM	<u>Binding Domain</u>		<u>% trans-spliced</u>	
	<u>Region</u>	<u>Size (nt)</u>	<u>226 sd</u>	<u>880 sd</u>
HPV-PTM1	A	80	69	0.6
HPV-PTM2	A	149	45	0.9
HPV-PTM5	A+B	140	55	0.8
HPV-PTM5ΔBP/PPT	A+B	140	0.5	0.2
HPV-PTM6	A+B	117	59	1
HPV-PTM8	C	104	7	37
HPV-PTM9	C	174	14	22
CF-PTM27	CF intron	411	0	0

Quantification of *trans*-splicing efficiency using real-time QRT-PCR



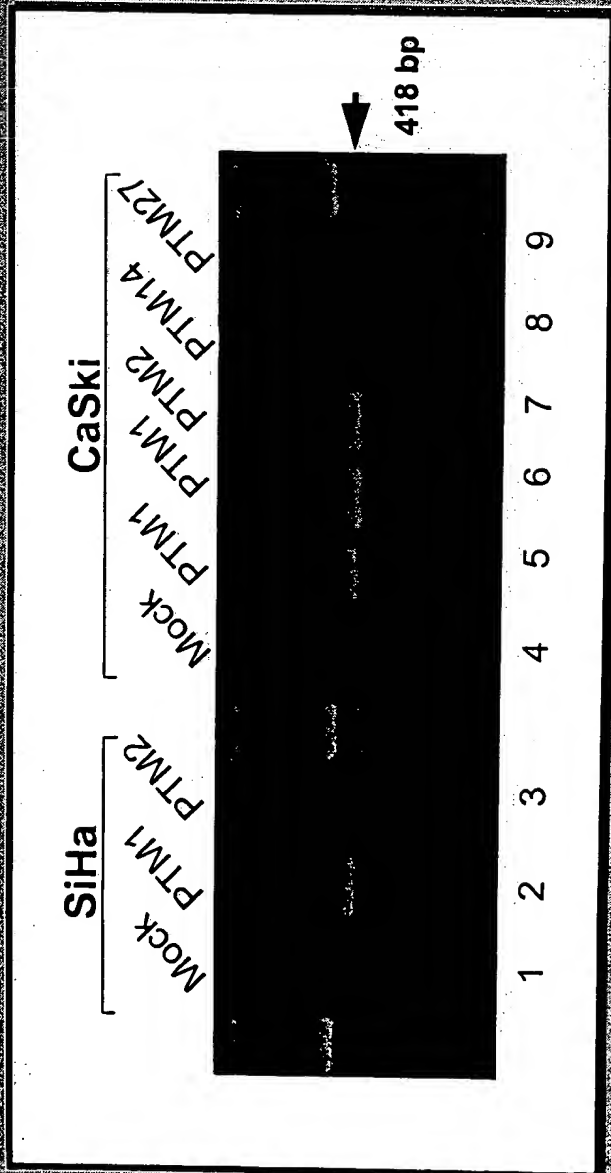
# Trans-splicing into Endogenous HPV Pre-mRNA Target in SiHa & CaSki Cells

## RT-PCR Conditions

- Total RNA: 400 ng/rxn
- Primer's: oJMD15 + Lac16R
- # Cycles : 35
- Expected product : 418 bp

## Details

- PTM1, PTM2 : HPV targeted, specific
- PTM14 : CF targeted, non-specific, has 23 bp BD
- PTM14 : CF targeted, non-specific, has 411 bp BD



SiHa : Single copy  
CaSki : ~ 400-500 copy!!

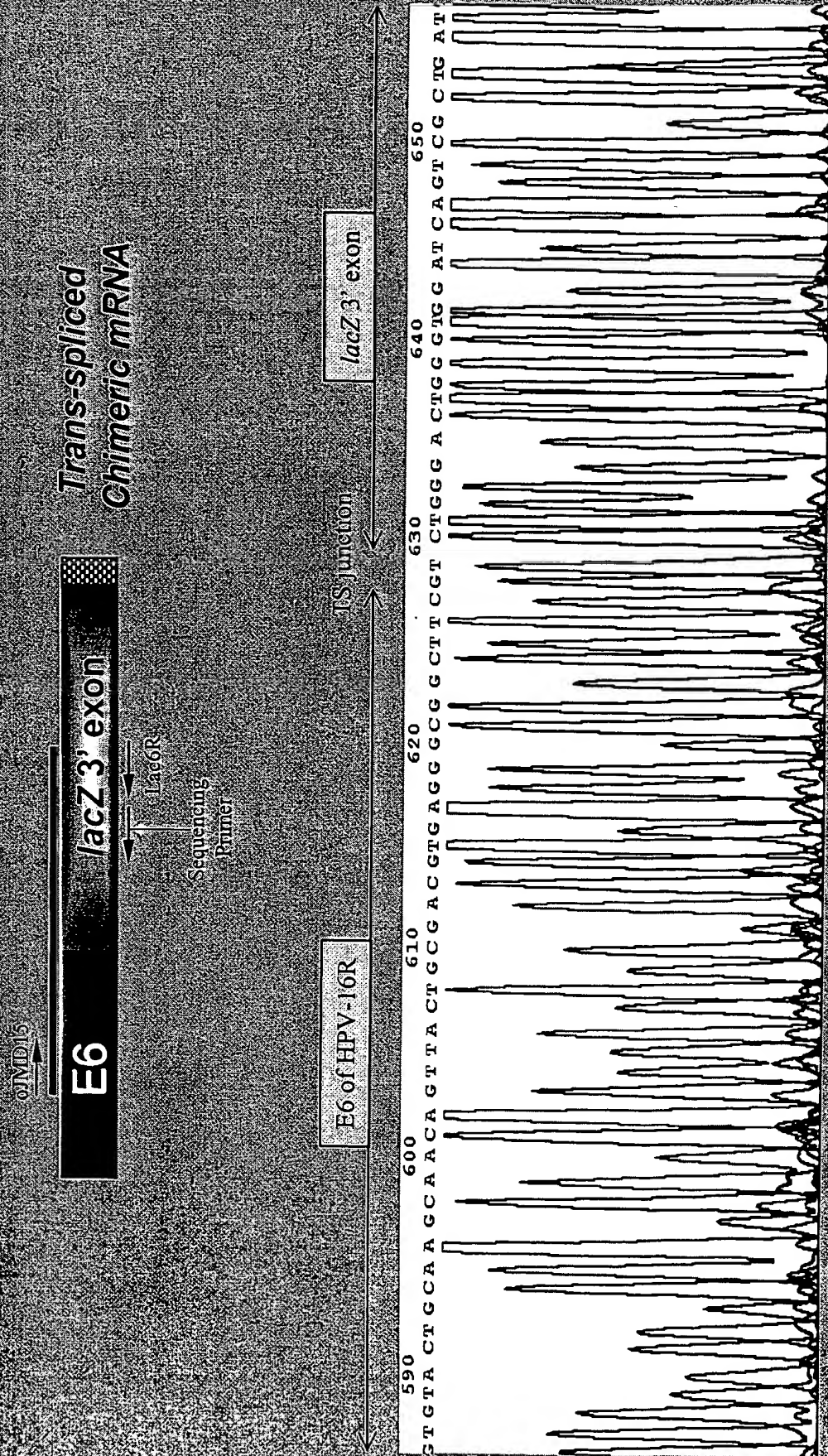


RT-PCR Analysis of total RNA

FIGURE 59

INTRONN

# Accurate Trans-splicing of HPV-PTM1 in Si Ha Cells (Endogenous target pre-mRNA)



INTRONN

FIGURE 60



# Trans-splicing in SiHa Transfections

(Endogenous target)

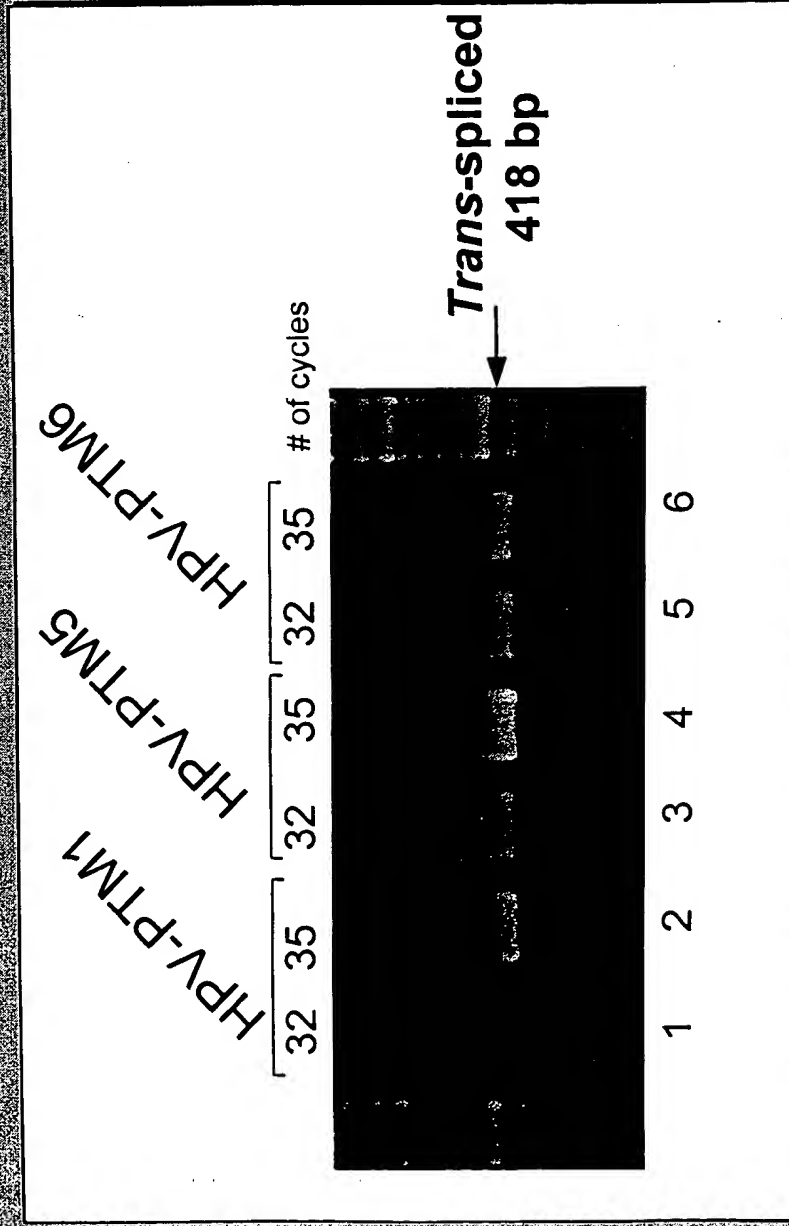
<u>PTM</u>	<u>% <i>trans</i>-spliced</u>
pcDNA3.1	0
HPV-PTM1	0.16
HPV-PTM5	0.12
HPV-PTM6	0.11
CF-PTM27	0

Quantification of *trans*-splicing efficiency using real-time QRT-PCR

INTRONN

FIGURE 6

# Trans-splicing Efficiency of HPV-PTM1, 5, & 6 in SiHa Cells



- SiHa cells transfected with 1.5 µg plasmid DNA, LipoPlus
- RNA isolated after 48 hr
- Total RNA: 500 ng/Rxn
- Primers: oJMD15 + Lac16R
- Expected product: 418 bp

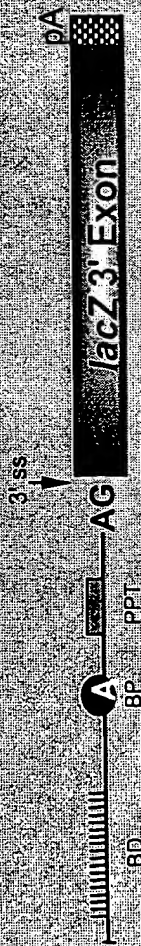
1, 3, 5, : 32 cycles

2, 4, 6, : 35



# Deletion of polypyrimidine tract abolishes *trans*-splicing

HPV-PTM5 (has all the elements)



HPV-?PPT (PPT has been deleted)



? PPT mutant      HPV-PTM5      Total RNA (μg)

200      400      200      400



## Methods:

- SiHa cells transfected with 1.5 μg of plasmid DNA
- Total RNA isolated after 48 hr and analyzed by RT-PCR (30 cycles)

Primers: oJMD15+Lac6R  
Expected product: 269 bp

Lanes 1 & 2: RNA from cells transfected with HPV-? PPT (mutant); No trans-splicing detected

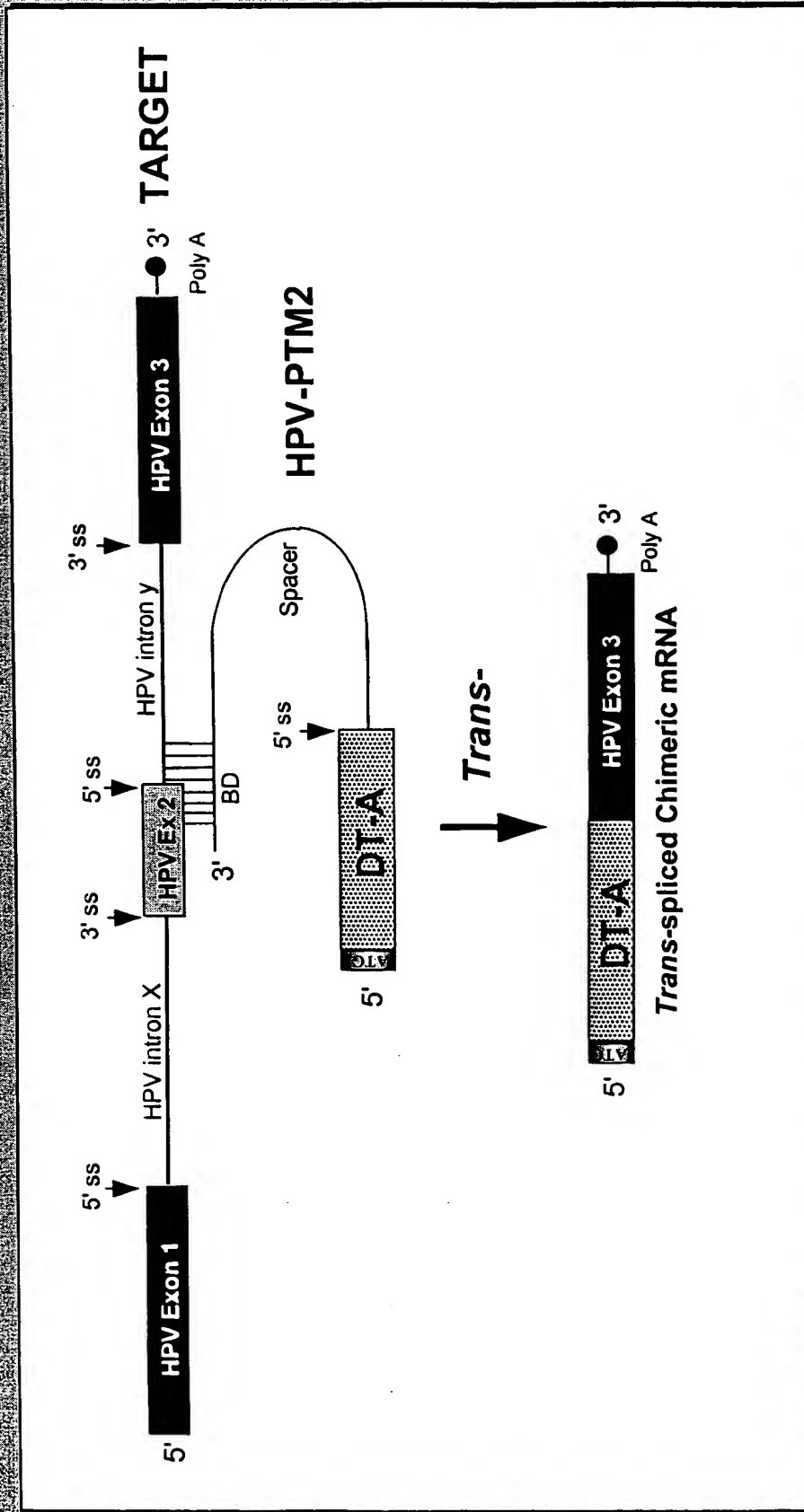
Lanes 3 & 4: RNA from cells transfected with HPV-PTM5 plasmid; trans-splicing Detected (269 bp product)

INTRON

FIGURE 63

85 of 89

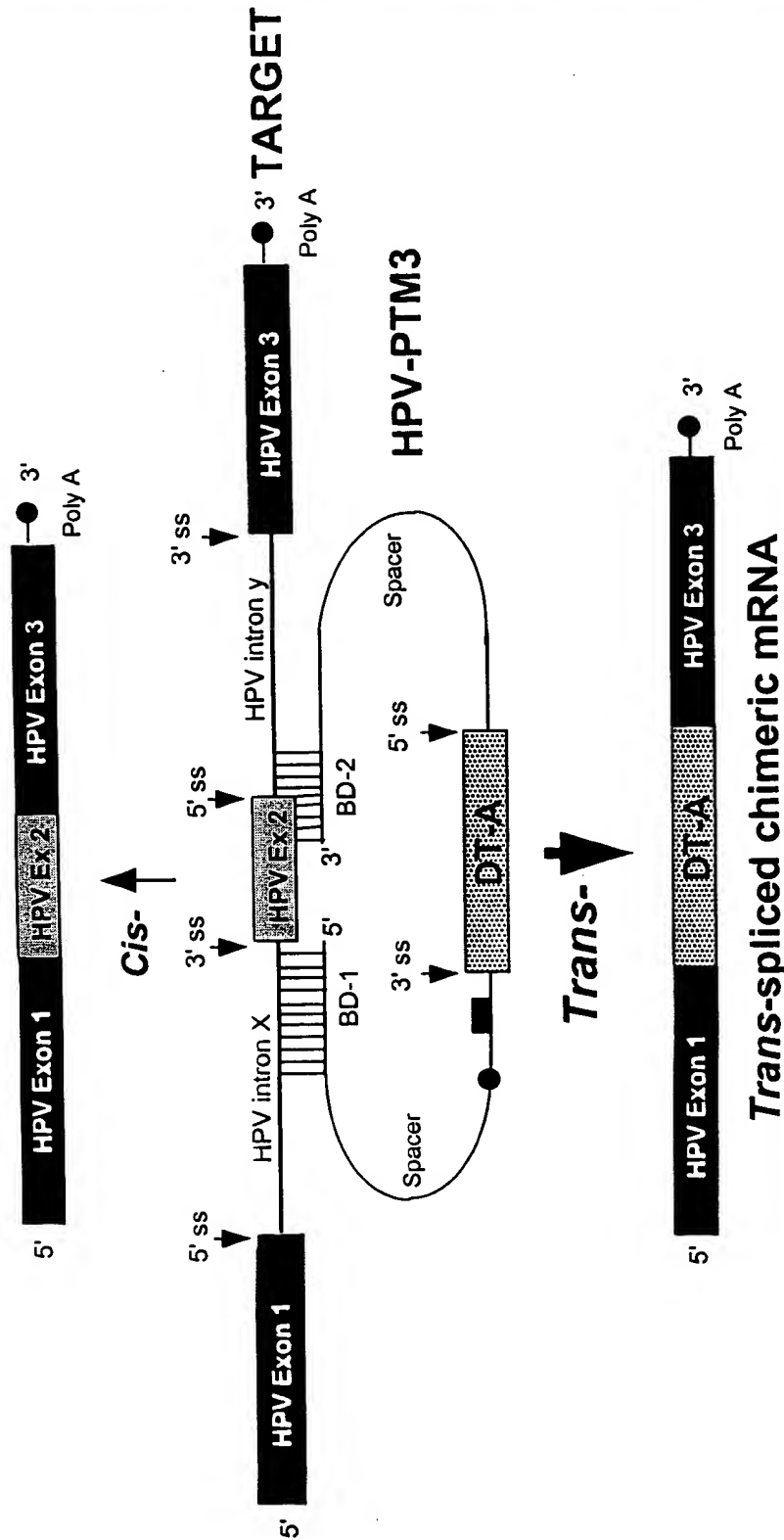
# SMaRT Strategy by 5' Exon Replacement



Schematic diagram of a PTM binding to the 5' splice site of the HPV mini-gene target

hr to 9.8

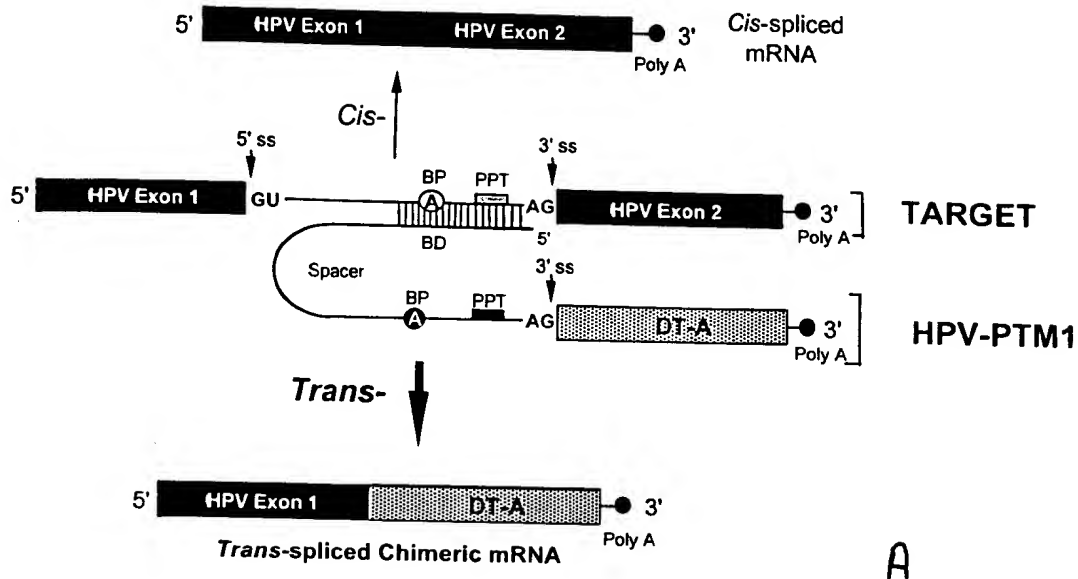
# Double Trans-splicing



Schematic diagram of a double trans-splicing PTM binding to the 3' and 5' splice sites of the HPV mini-gene target

INTRONN

**SMaRT Strategy by 3' Exon Replacement:** Schematic diagram of a PTM binding to the 3' splice site of the HPV mini-gene target



**SMaRT Strategy by 5' Exon Replacement:** Schematic diagram of a PTM binding to the 5' splice site of the HPV mini-gene target

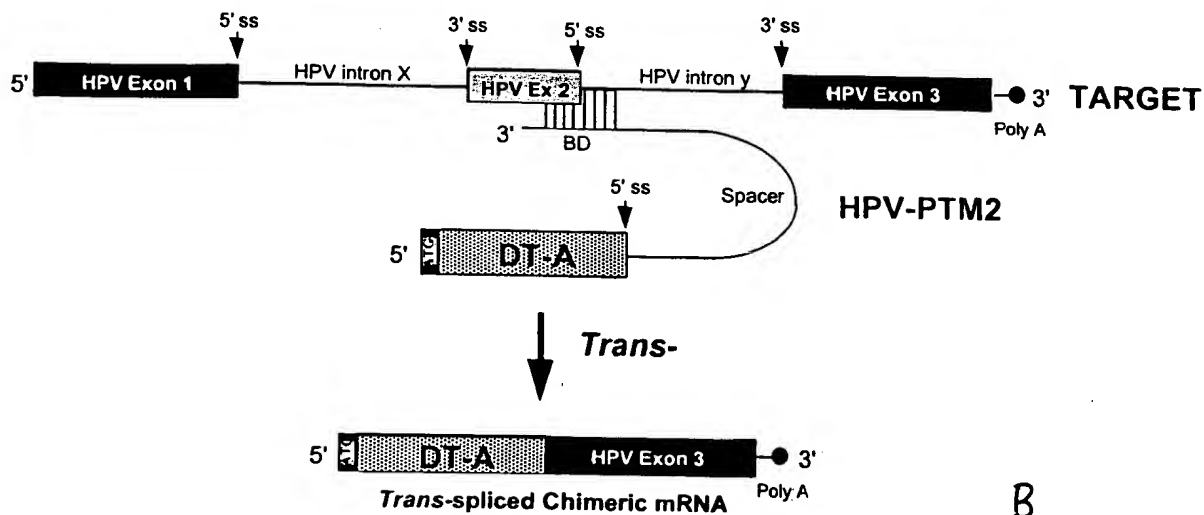


FIGURE 66



HPV-PTM3 (For Internal exon replacement)

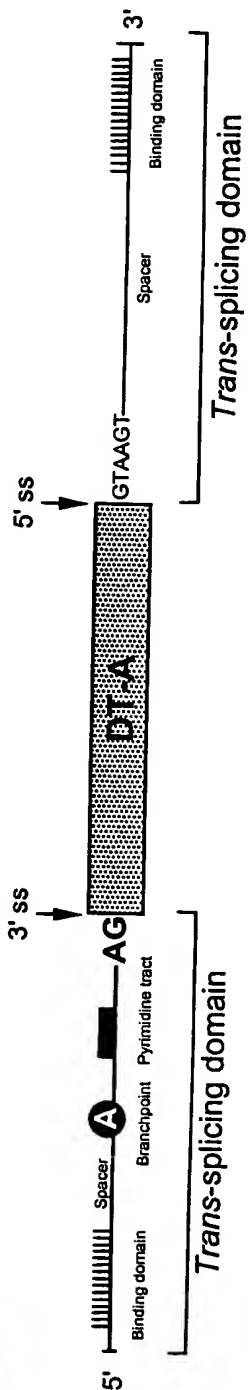


FIGURE 67